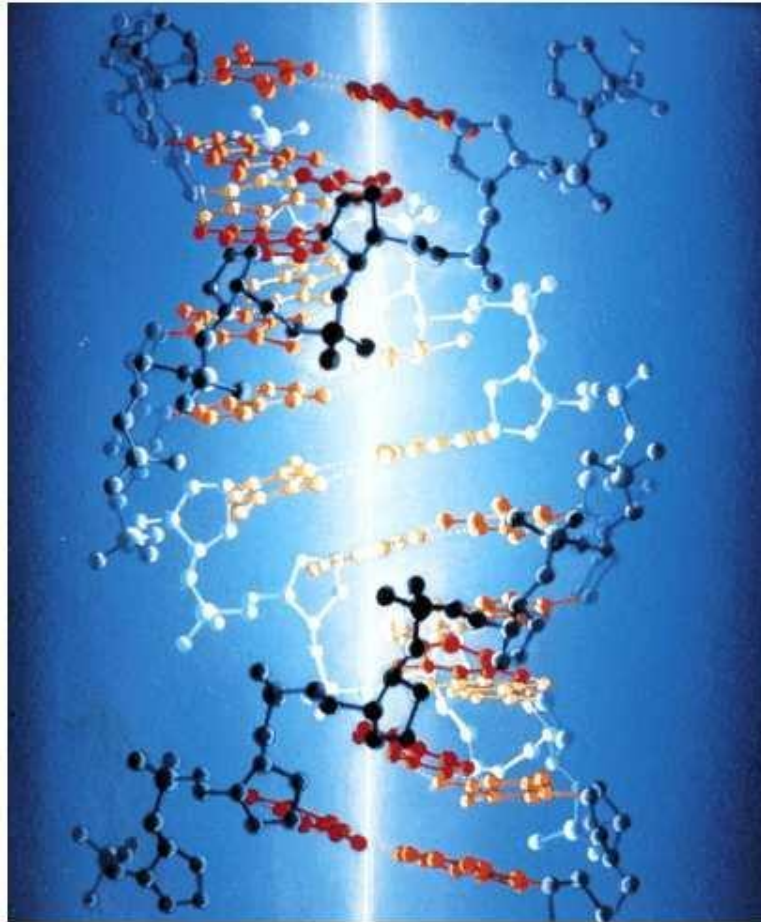


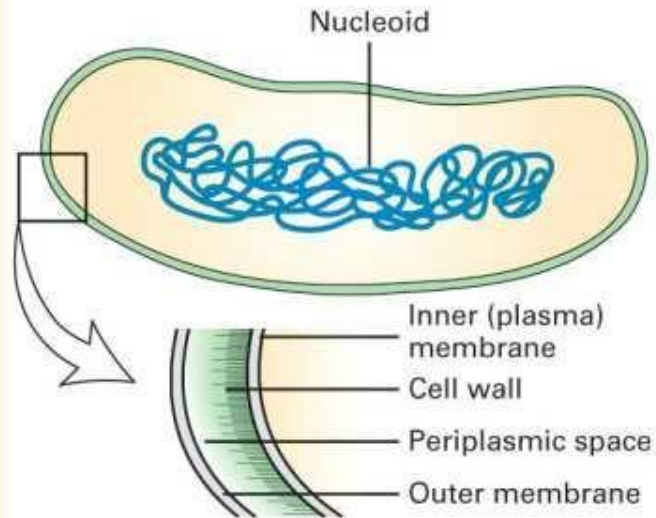
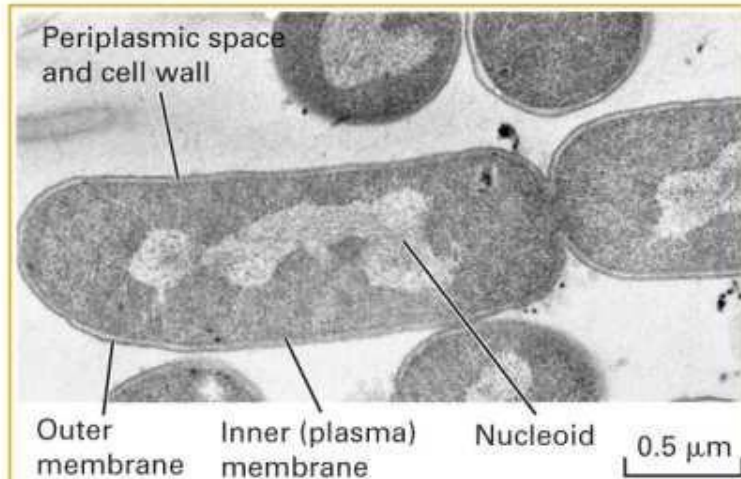
DNA, RNA, Recombinant DNA Technology



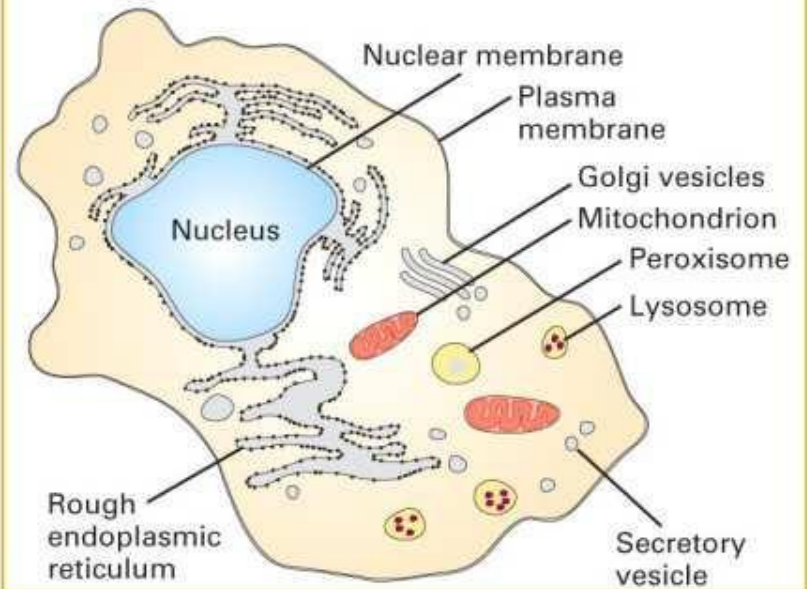
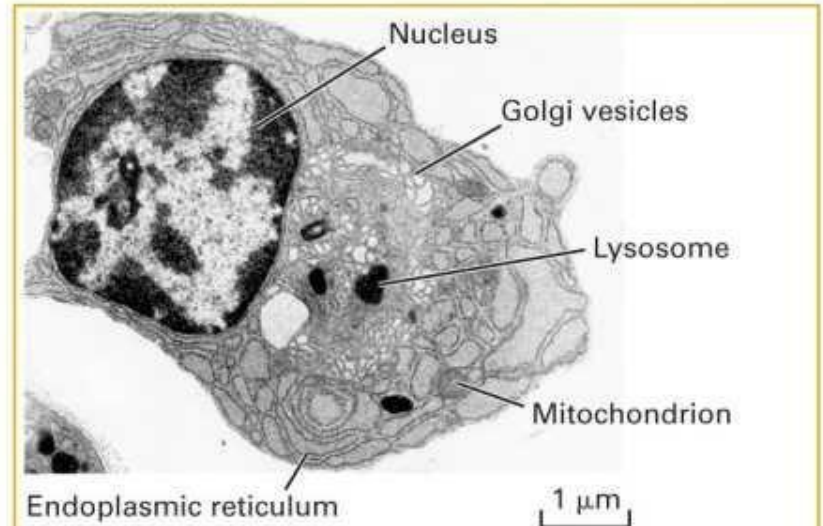
Chapter 3 Opener Fundamentals of Biochemistry, 2/e

“Metabolic pathways” expanded

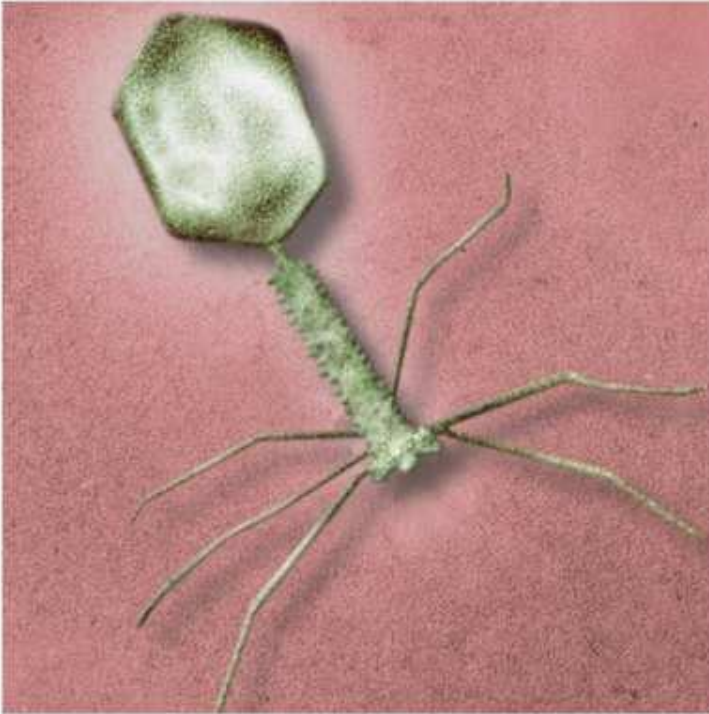
(a) Prokaryotic cell



(b) Eukaryotic cell



Model organisms: Cellular biology, biochemistry...
molecular biology



Viruses

Proteins involved in DNA, RNA,
protein synthesis

Gene regulation

Cancer and control of cell
proliferation

Transport of proteins and
organelles inside cells

Infection and immunity

Possible gene therapy approaches



Fruit fly (*Drosophila melanogaster*)

Development of the body plan

Generation of differentiated cell lineages

Formation of the nervous system, heart, and musculature

Programmed cell death

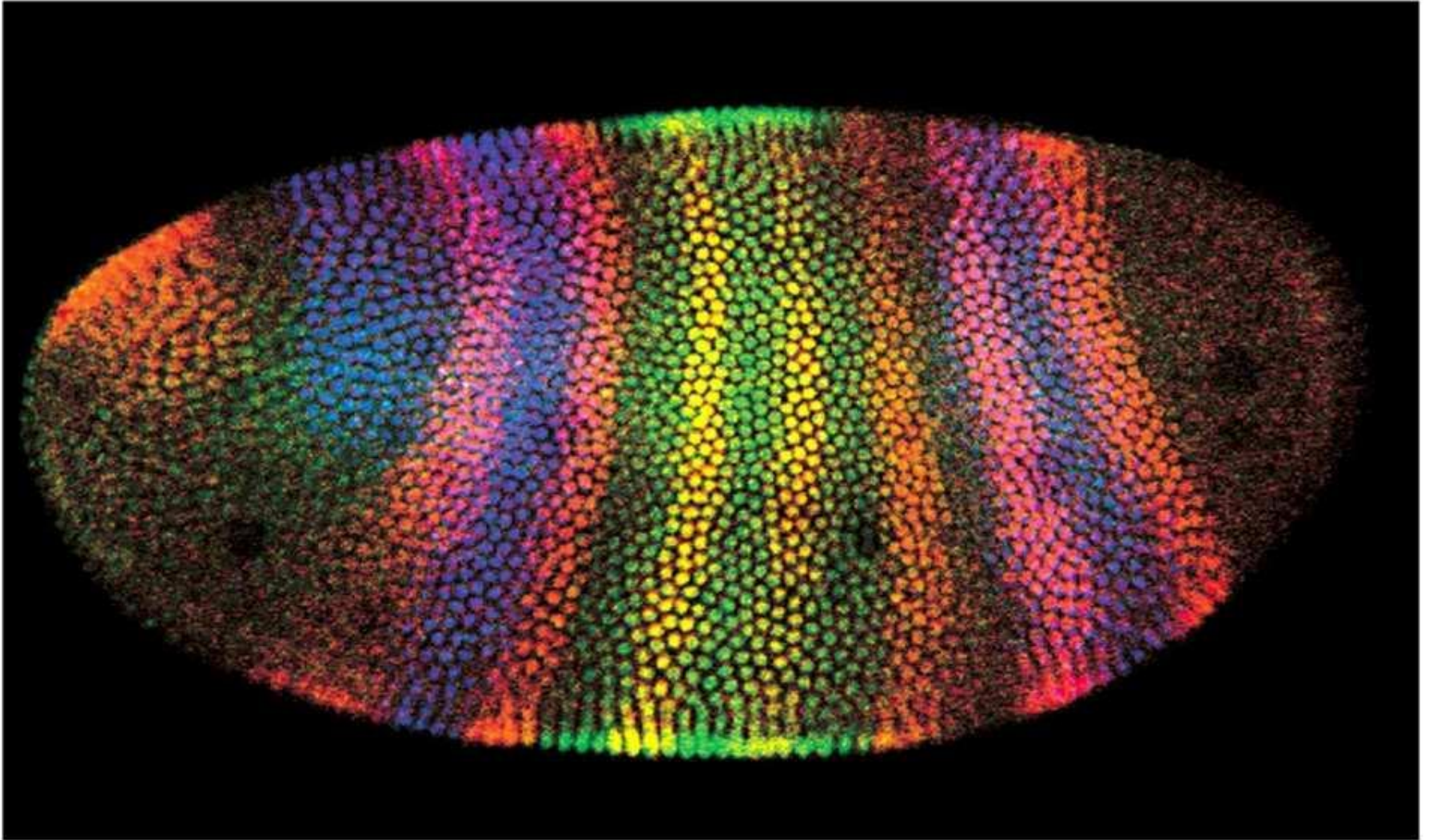
Genetic control of behavior

Cancer genes and control of cell proliferation

Control of cell polarization

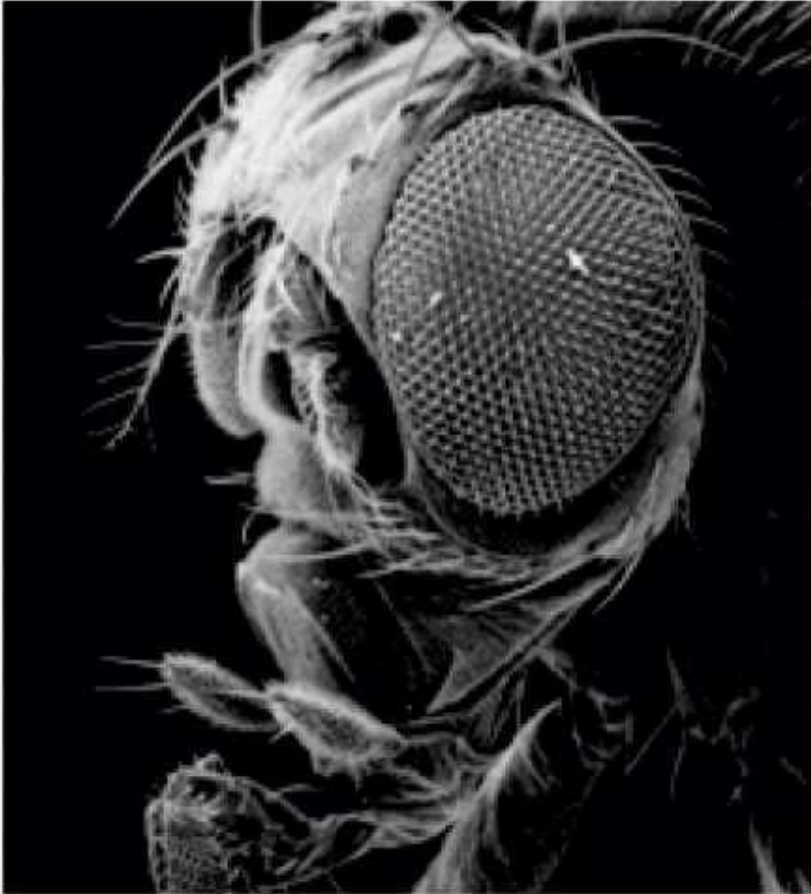
Effects of drugs, alcohol, pesticides

Developmental biology.....

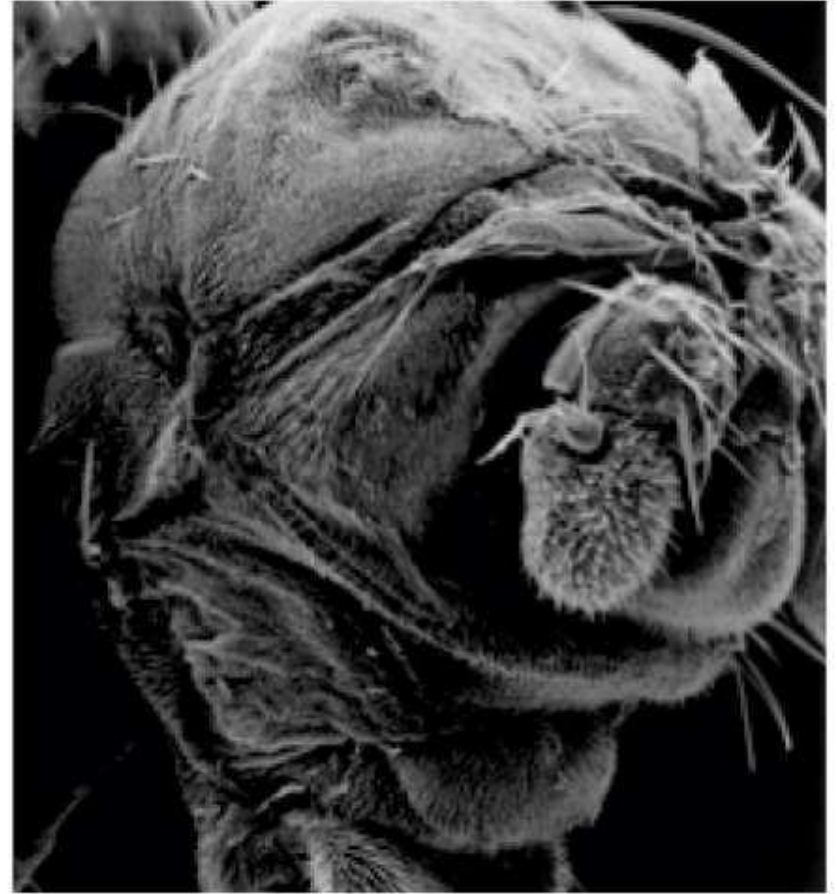


Fly mutation “eyeless”

(b)



(c)



‘The fly and you are not much different.’



Mice, including cultured cells

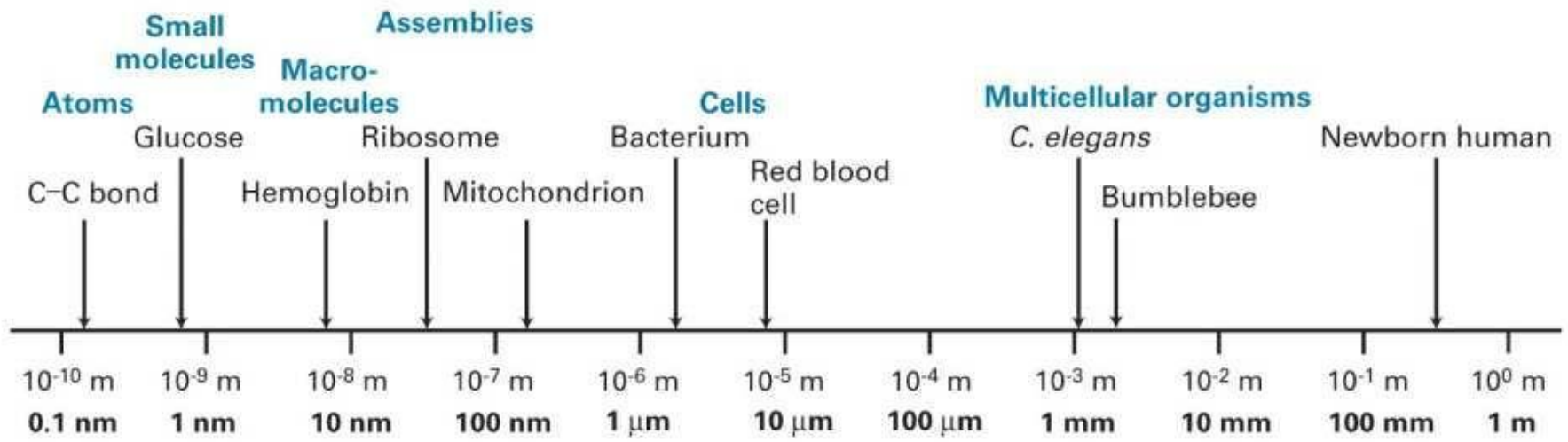
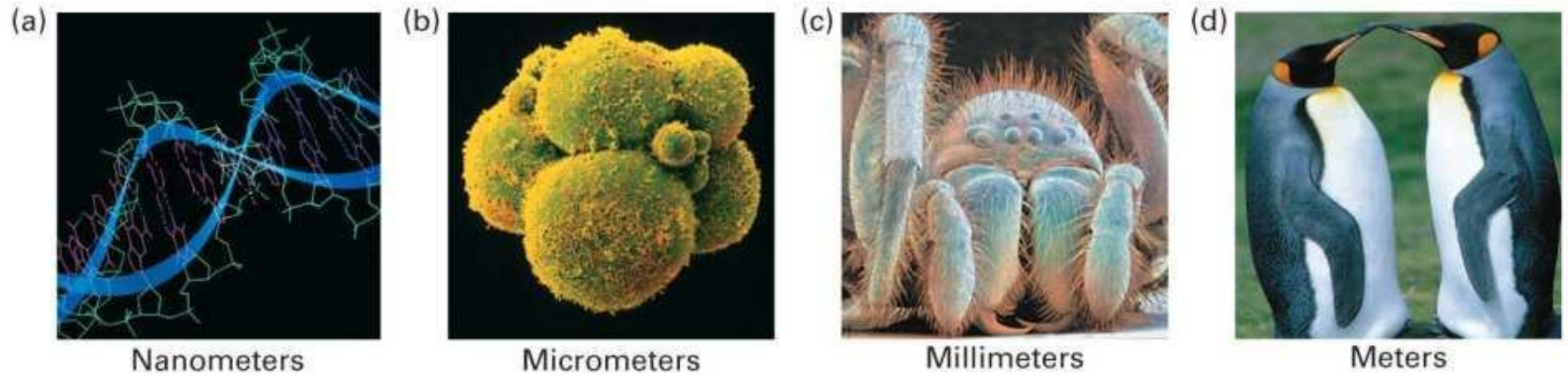
Development of body tissues
Function of mammalian immune system

Formation and function of brain and nervous system

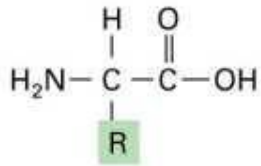
Models of cancers and other human diseases

Gene regulation and inheritance
Infectious disease

The 'first' science: "technology drives research drives technology dri..."

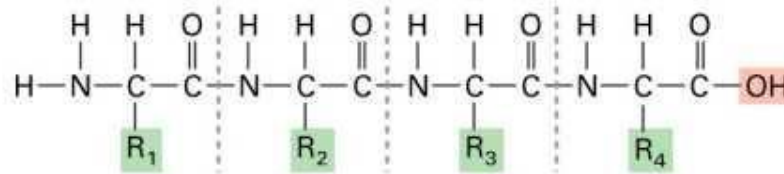


MONOMERS

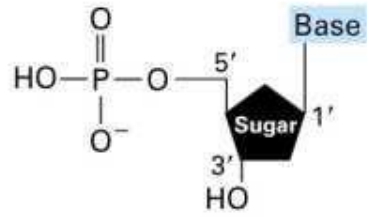
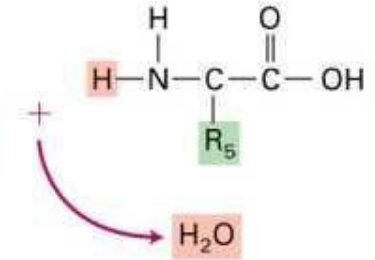


Amino acid

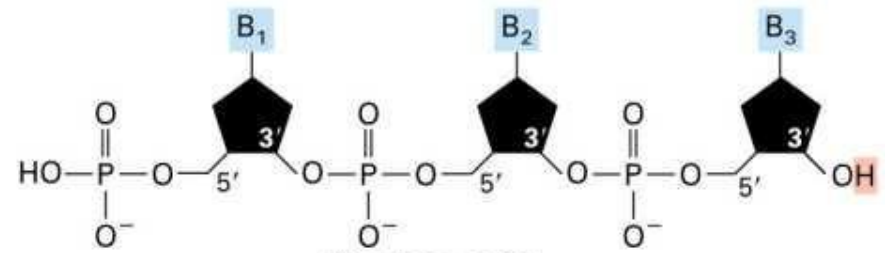
POLYMERS



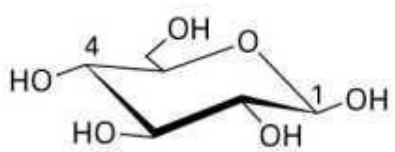
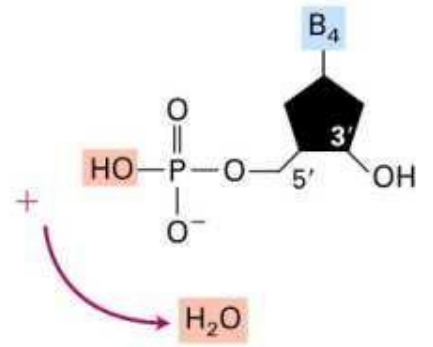
Polypeptide



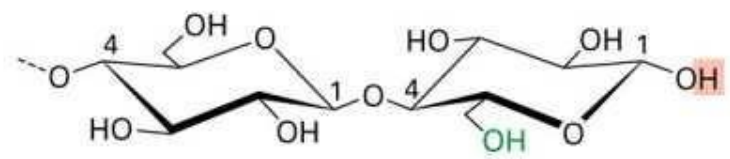
Nucleotide



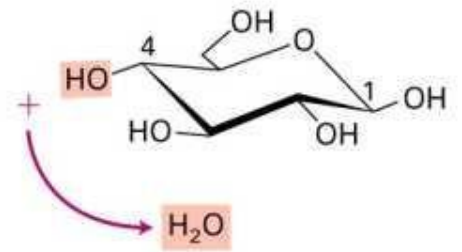
Nucleic acid



Monosaccharide

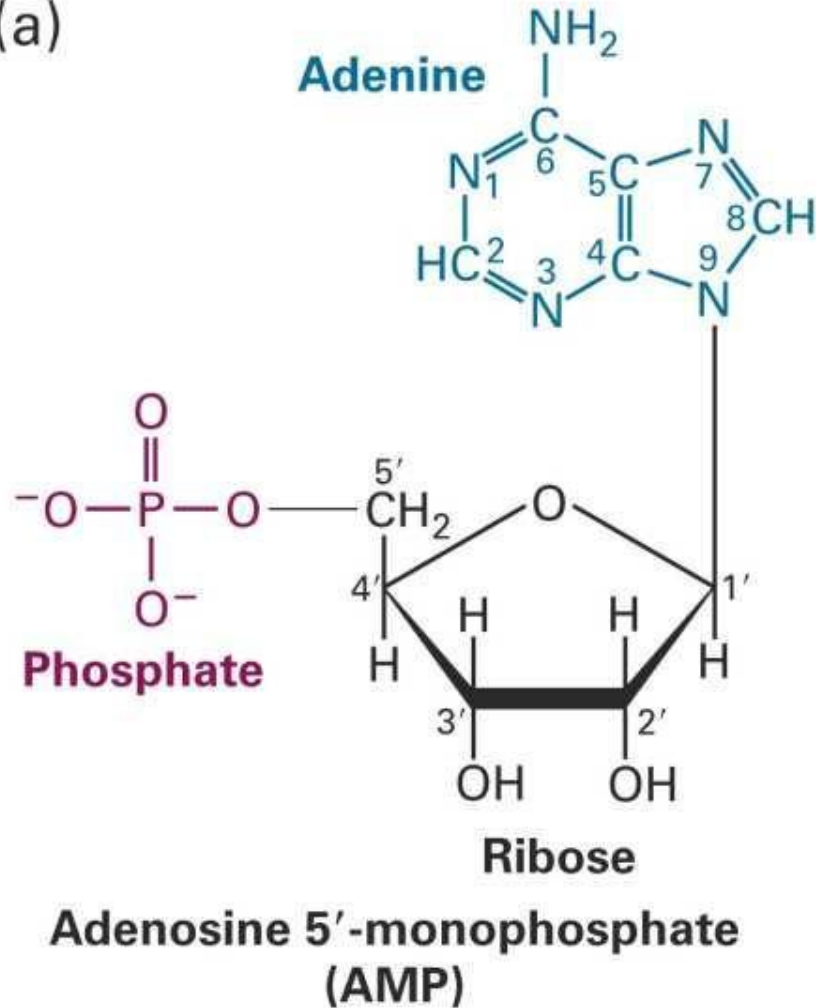


Polysaccharide

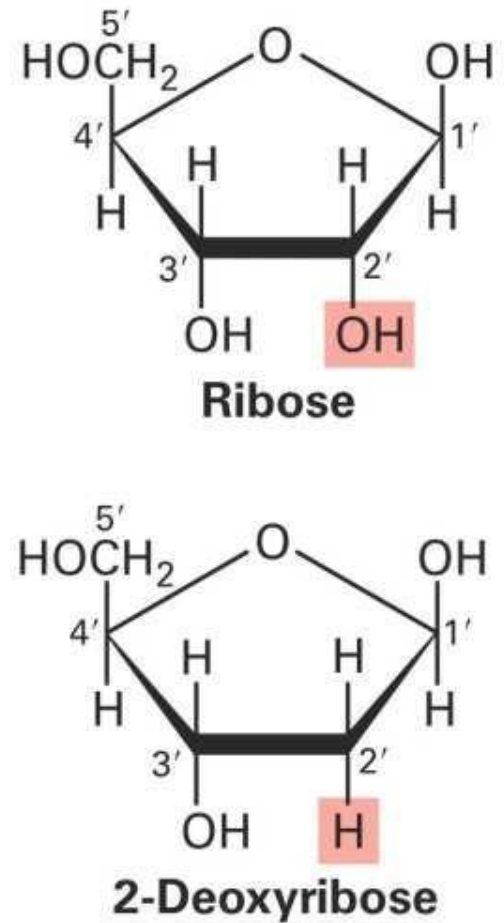


Nucleic Acids - DNA and RNA

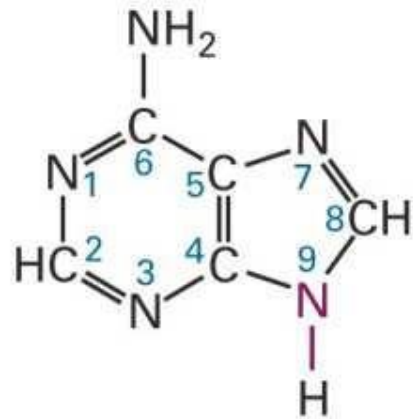
(a)



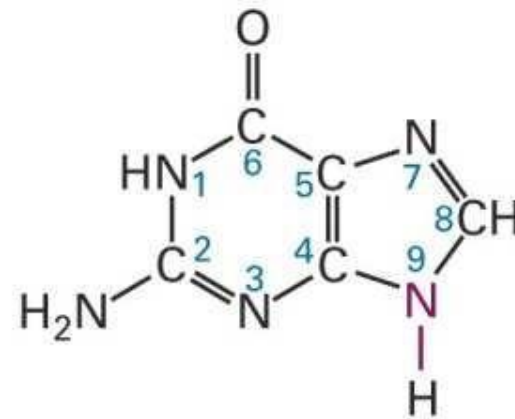
(b)



PURINES

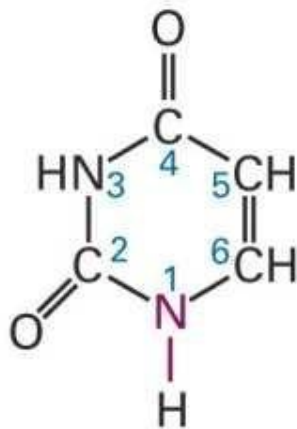


Adenine (A)



Guanine (G)

PYRIMIDINES



Uracil (U)

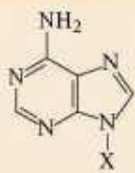
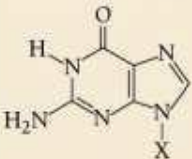
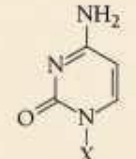
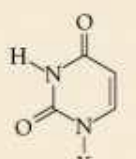
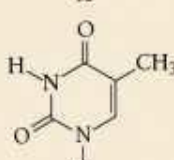


Thymine (T)



Cytosine (C)

Table 3-1 Names and Abbreviations of Nucleic Acid Bases, Nucleosides, and Nucleotides

Base Formula	Base (X = H)	Nucleoside (X = ribose ^a)	Nucleotide ^b (X = ribose phosphate ^a)
	Adenine Adc A	Adenosine Ado A	Adenylic acid Adenosine monophosphate AMP
	Guanine Gua G	Guanosine Guo G	Guanylic acid Guanosine monophosphate GMP
	Cytosine Cyt C	Cytidine Cyd C	Cytidylic acid Cytidine monophosphate CMP
	Uracil Ura U	Uridine Urd U	Uridylic acid Uridine monophosphate UMP
	Thymine Thy T	Deoxythymidine dThd dT	Deoxythymidylic acid Deoxythymidine monophosphate dTMP

DNA + RNA

DNA + RNA

DNA + RNA

RNA ←

DNA ←

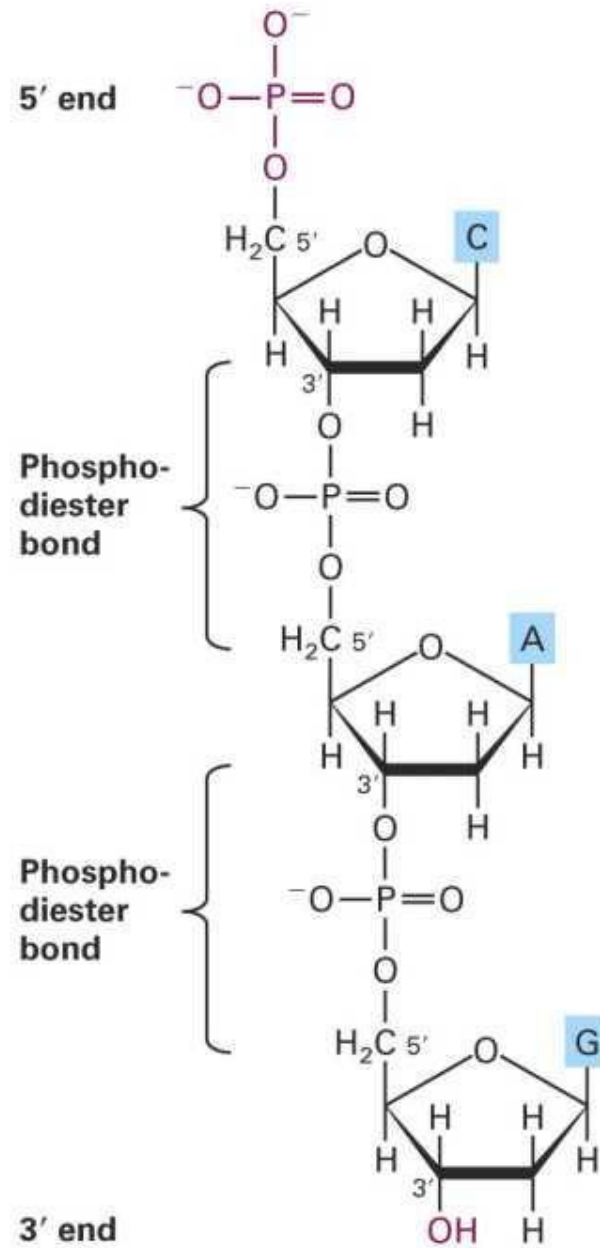
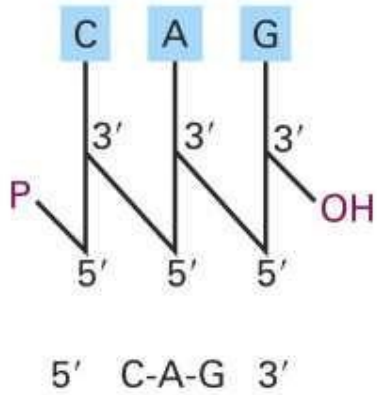
^aThe presence of a 2'-deoxyribose unit in place of ribose, as occurs in DNA, is implied by the prefixes "deoxy" or "d." For example, the deoxy-nucleoside of adenine is deoxyadenosine or dA. However, for thymine-containing residues, which rarely occur in RNA, the prefix is redundant and may be dropped. The presence of a ribose unit may be explicitly implied by the prefix "ribo."

^bThe position of the phosphate group in a nucleotide may be explicitly specified as in, for example, 3'-AMP and 5'-GMP.

Table 3-1 Fundamentals of Biochemistry, 2/e

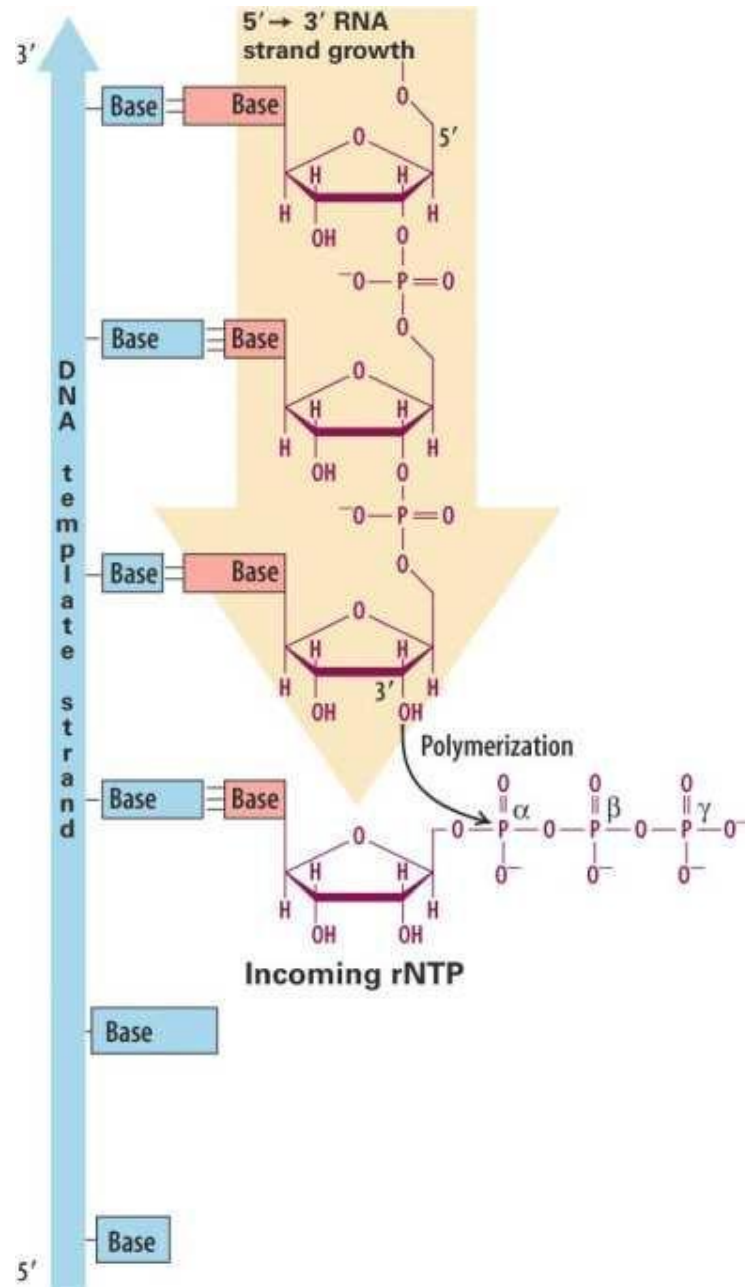
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DNA structure -> sequence

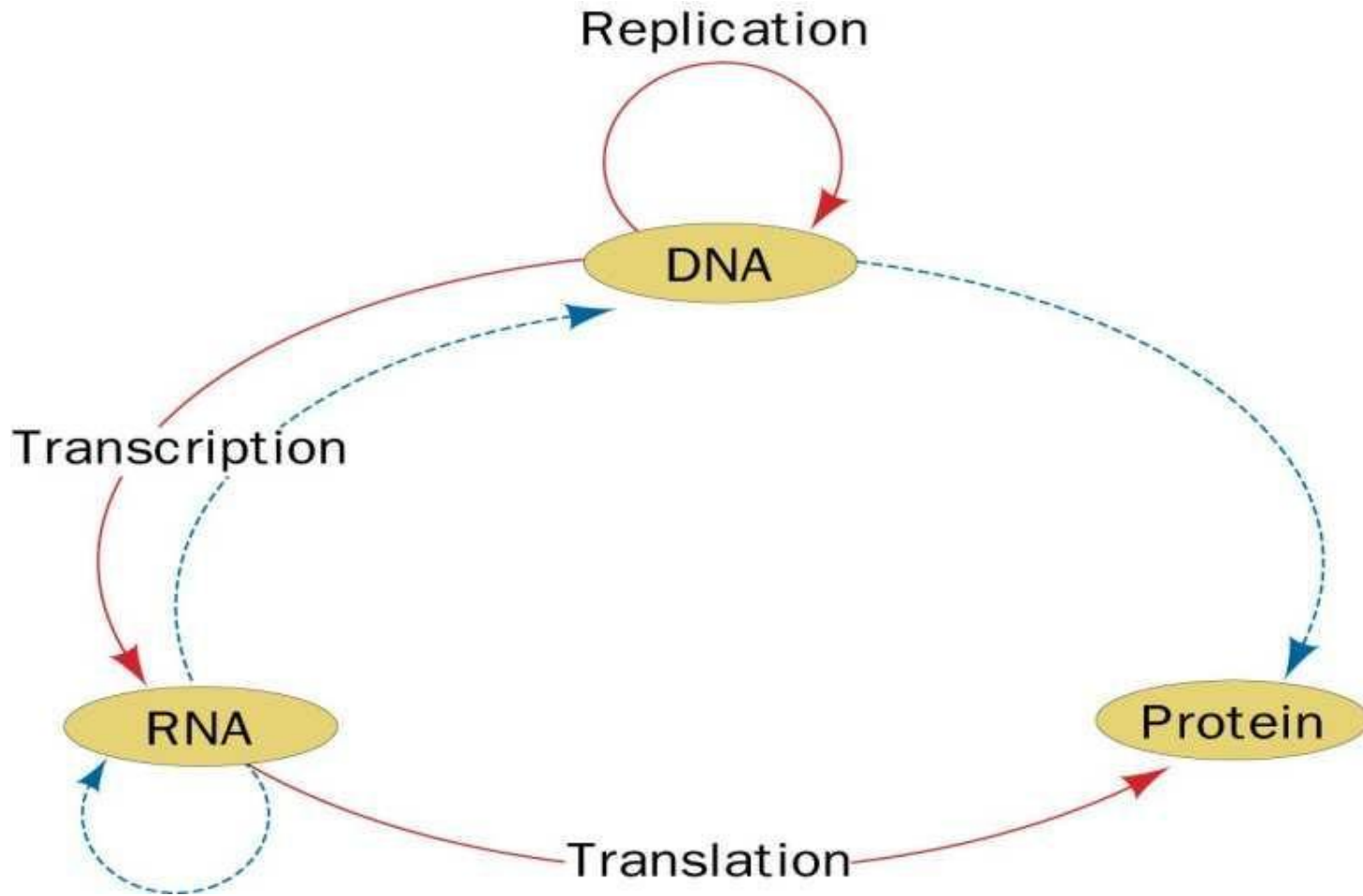


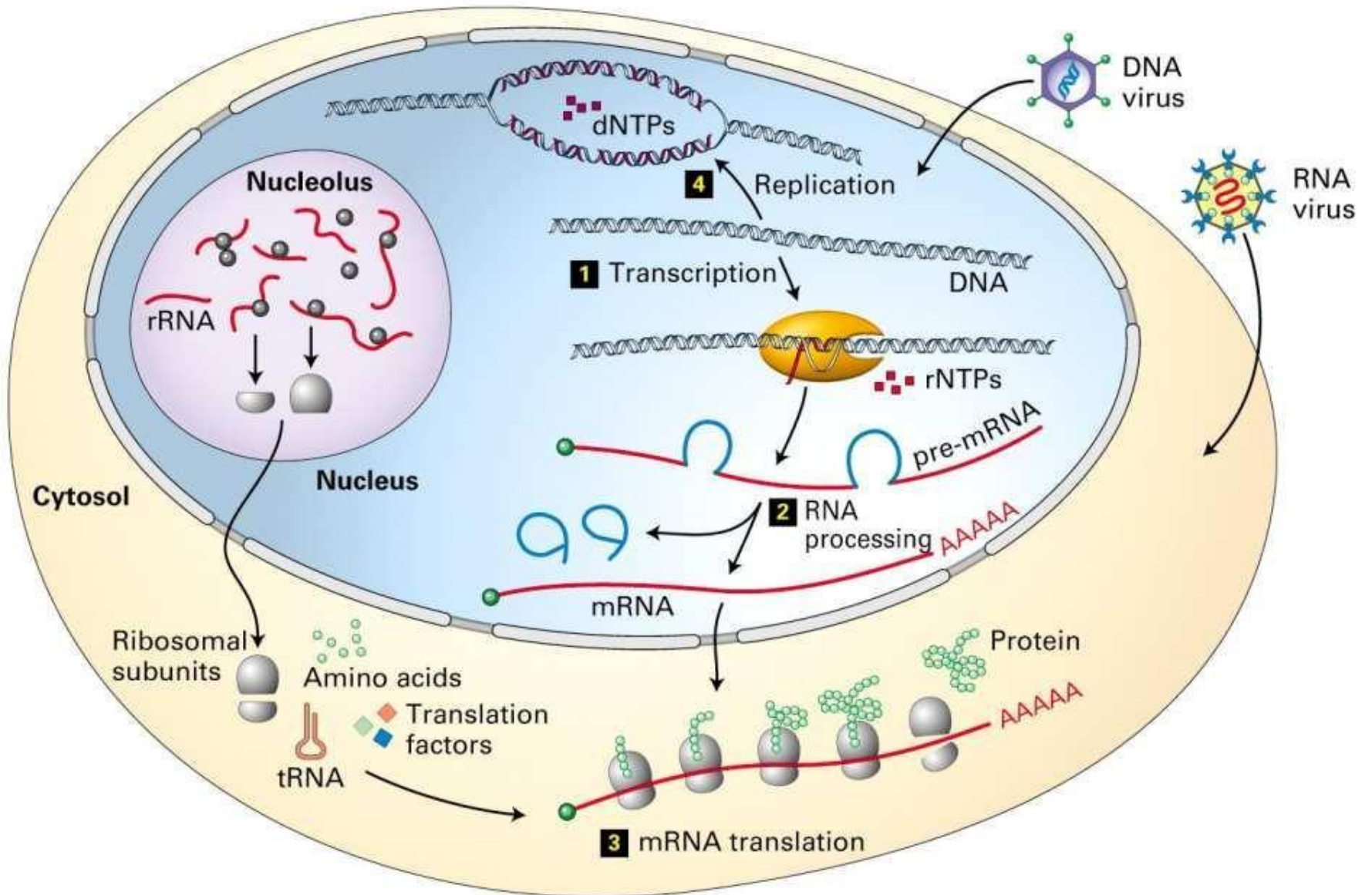
Polymerase reaction:

5' → 3'



The central dogma





Gene expression.

DNA 5' — A-G-A-G-G-T-G-C-T — 3'
3' — T-C-T-C-C-A-C-G-A — 5'



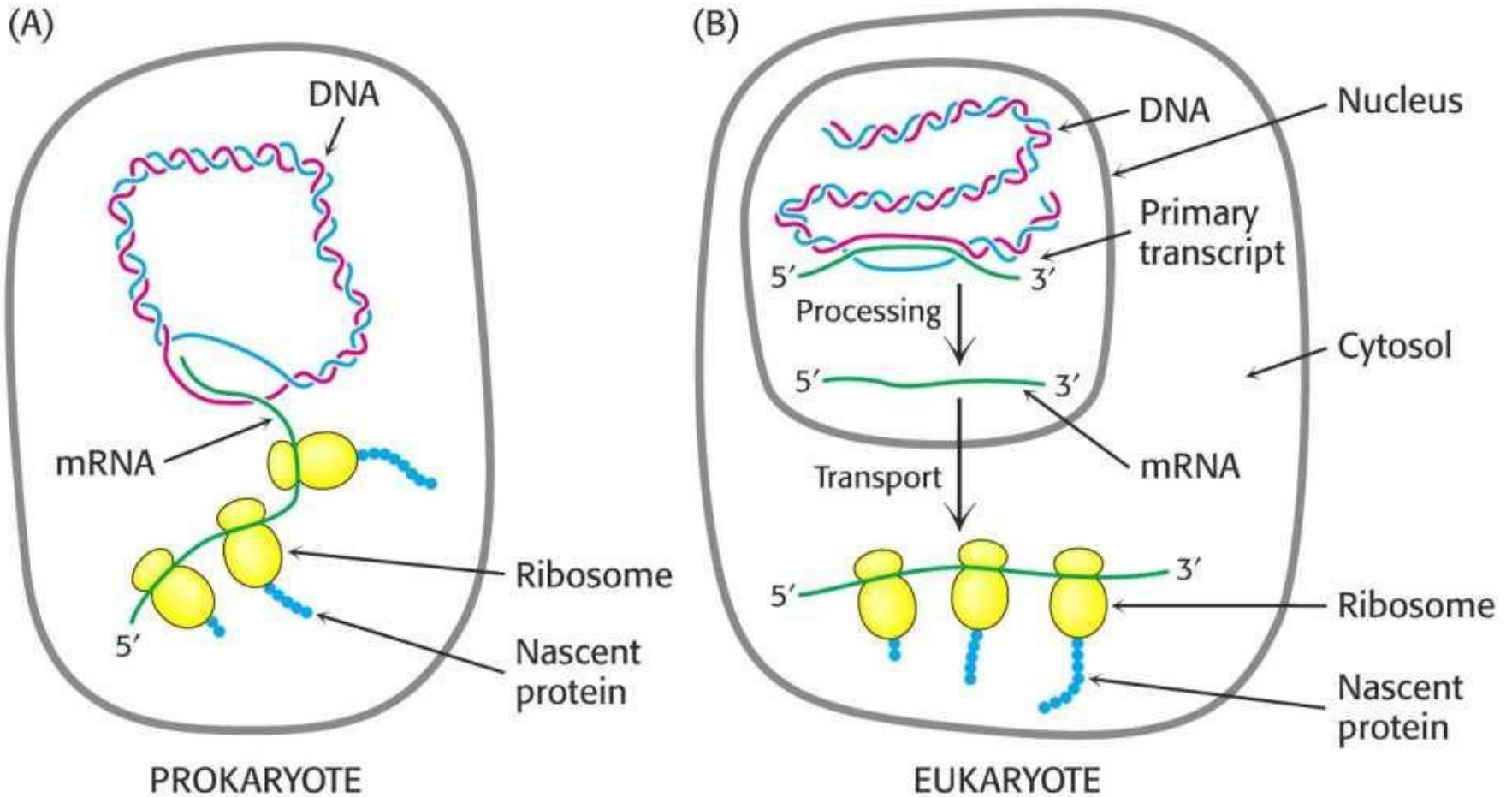
mRNA 5' — A-G-A-G-G-U-G-C-U — 3'
tRNAs U-C-U C-C-A C-G-A

┌───┐ ┌───┐ ┌───┐
| | | | | |
└───┘ └───┘ └───┘
Arginine Glycine Alanine

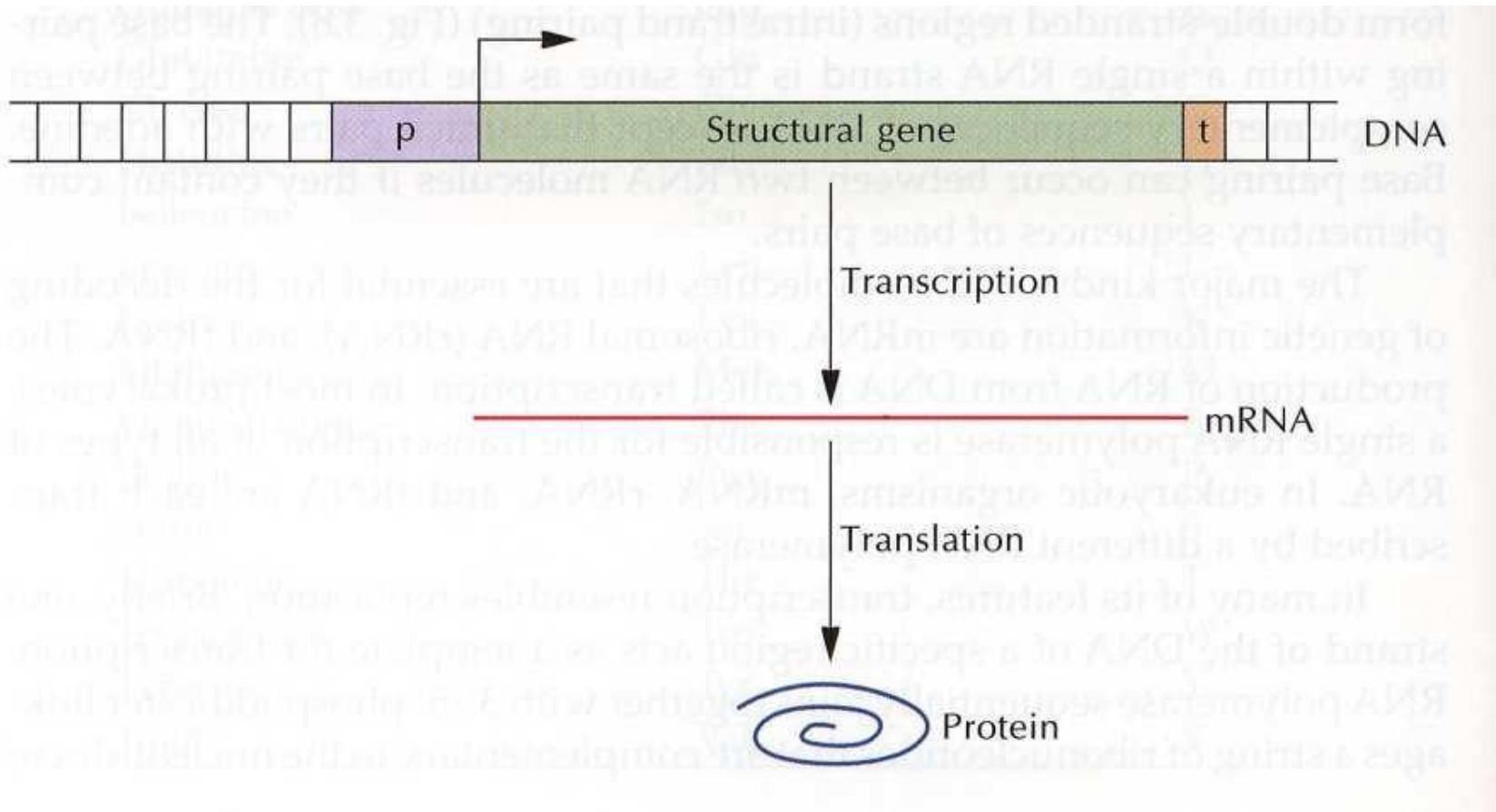


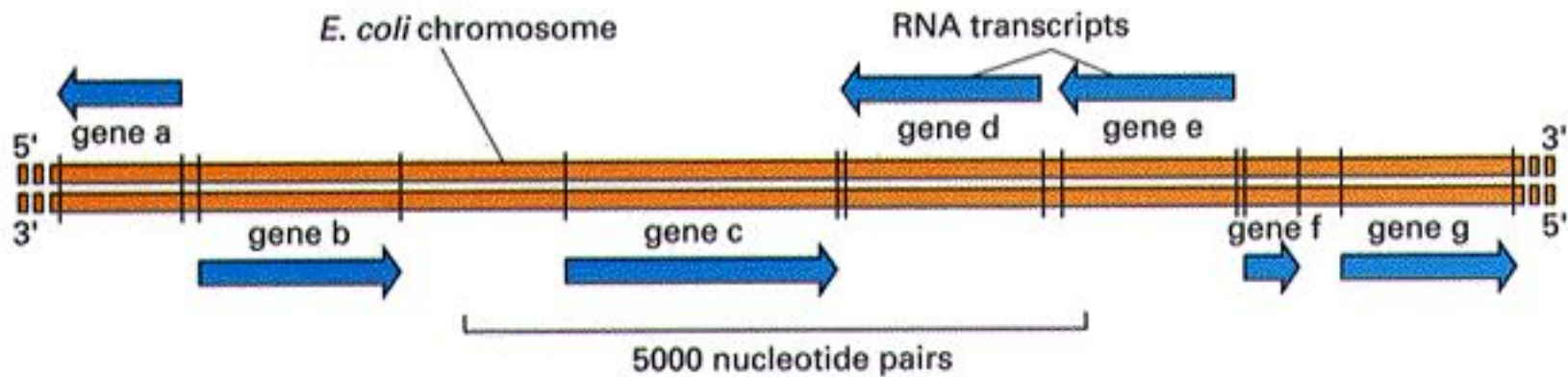
Polypeptide

—Arg—Gly—Ala—



What is a gene?







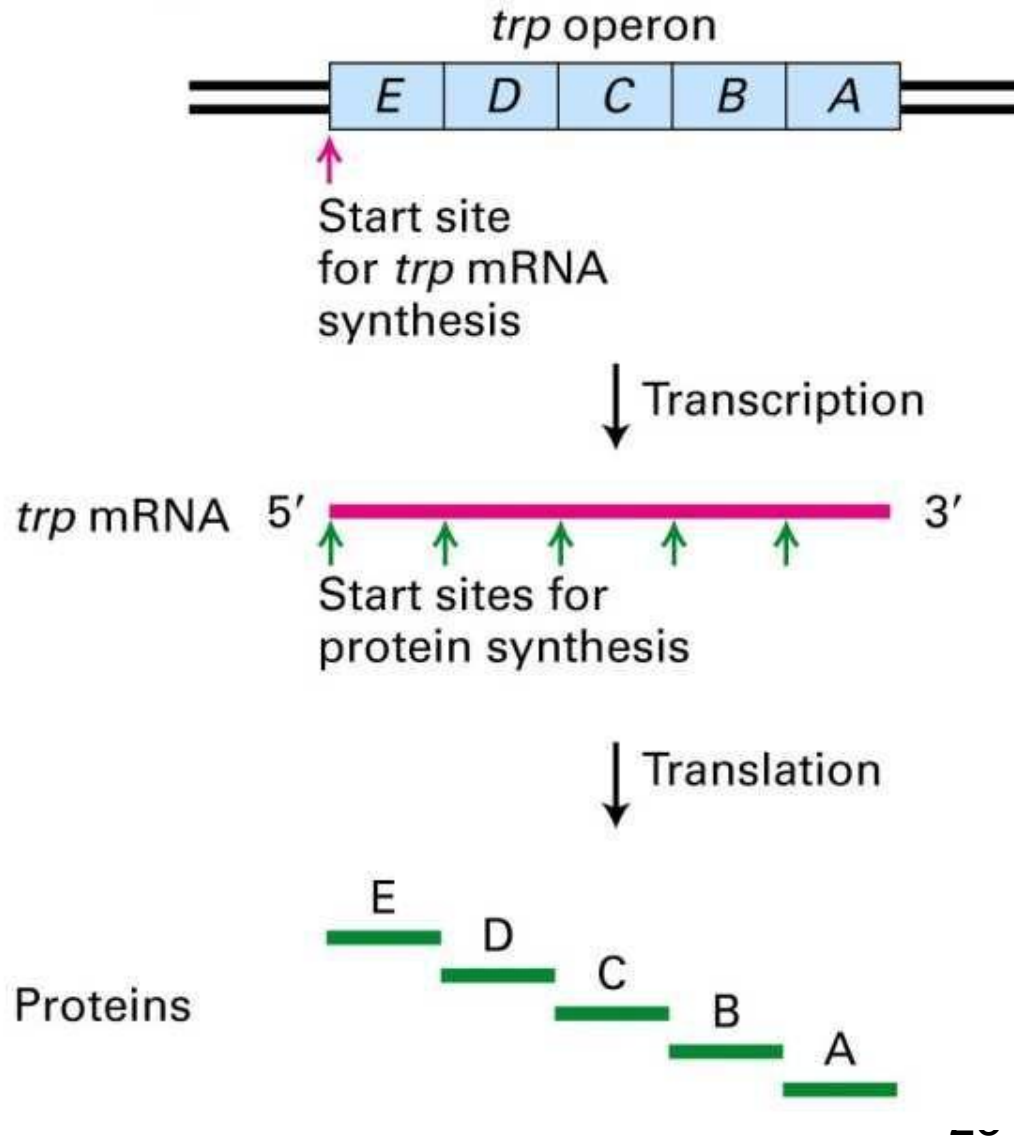
(a) Monocistronic



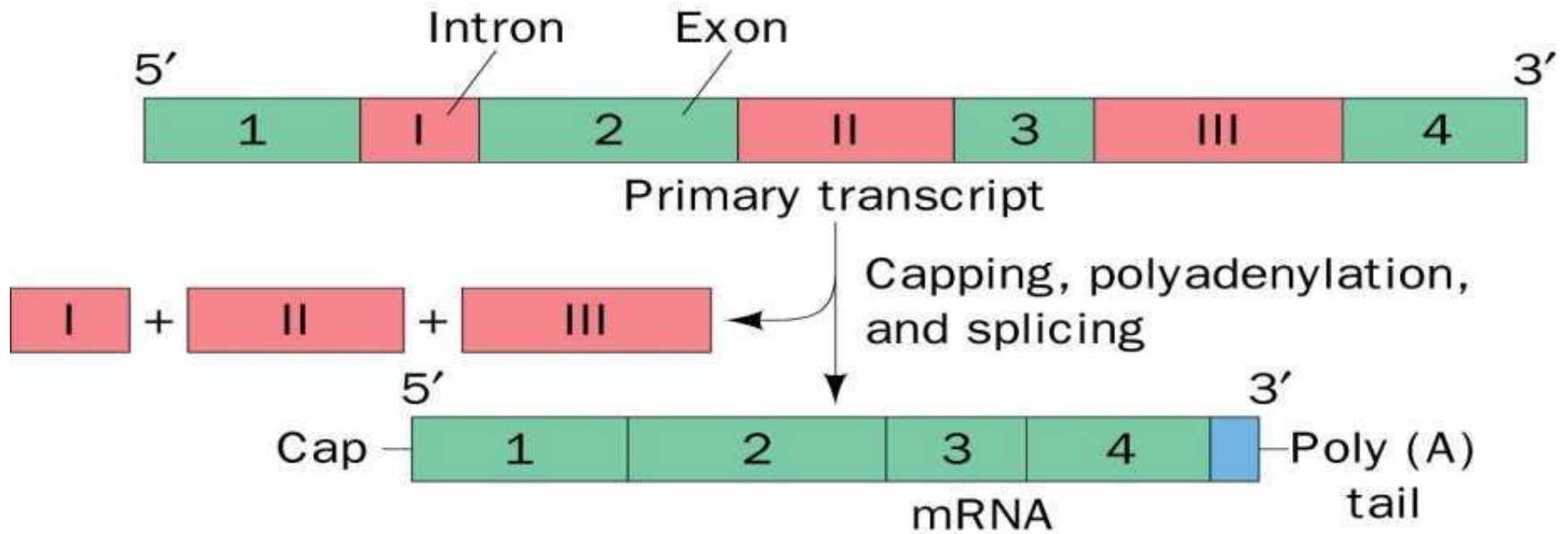
(b) Polycistronic

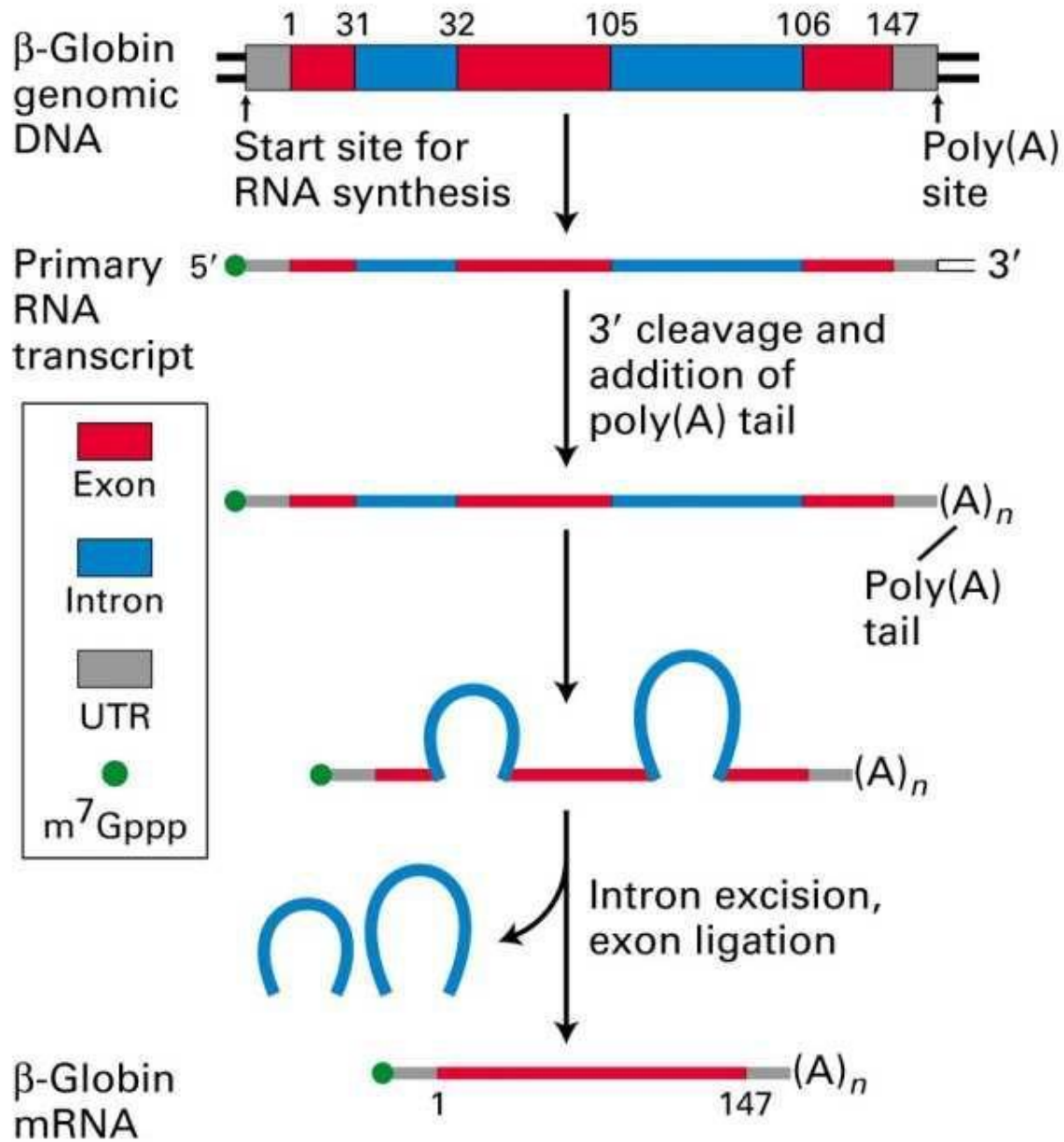
(a) Prokaryotes

E. coli genome



Eukaryotes - Intron-Exon concept





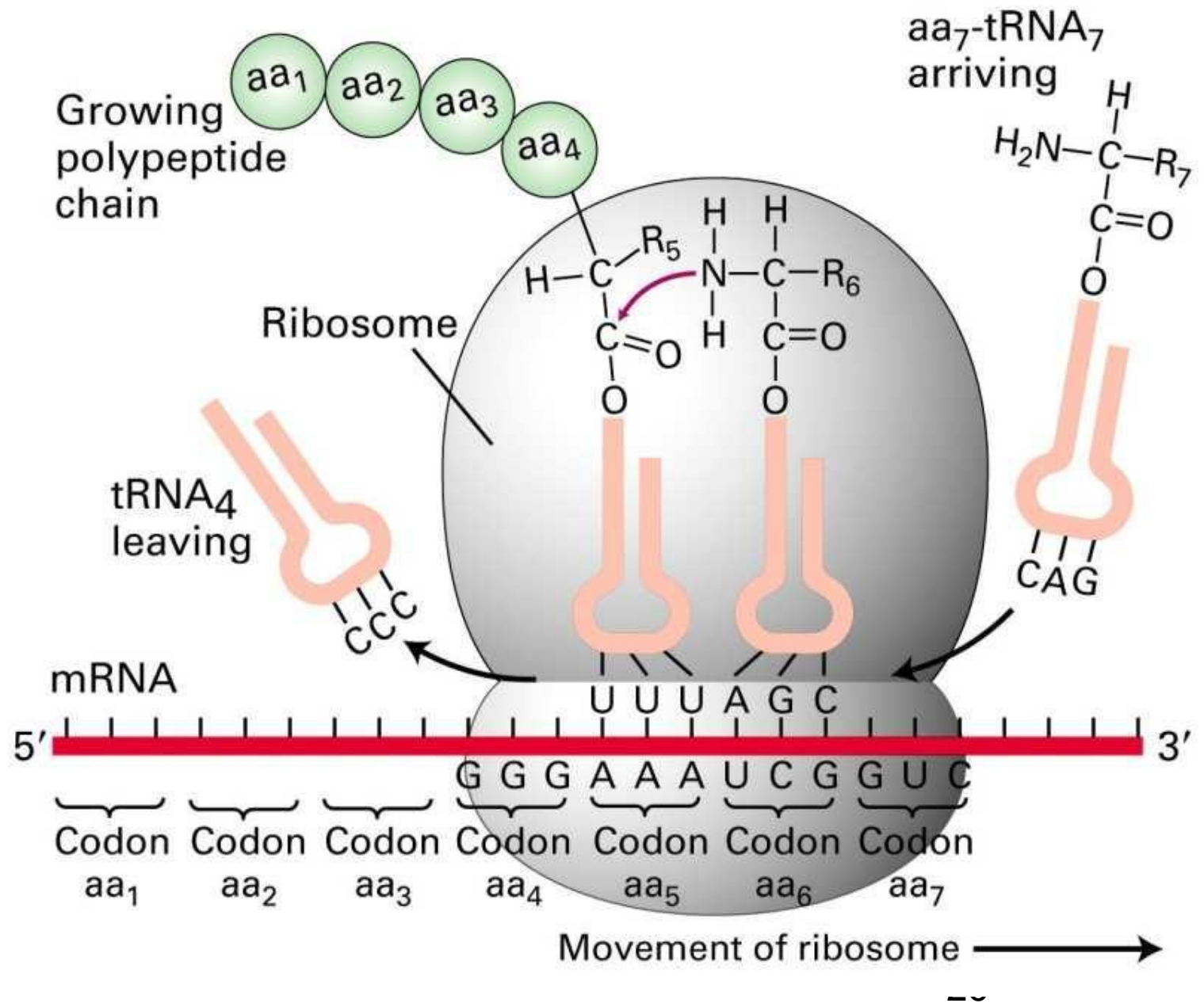
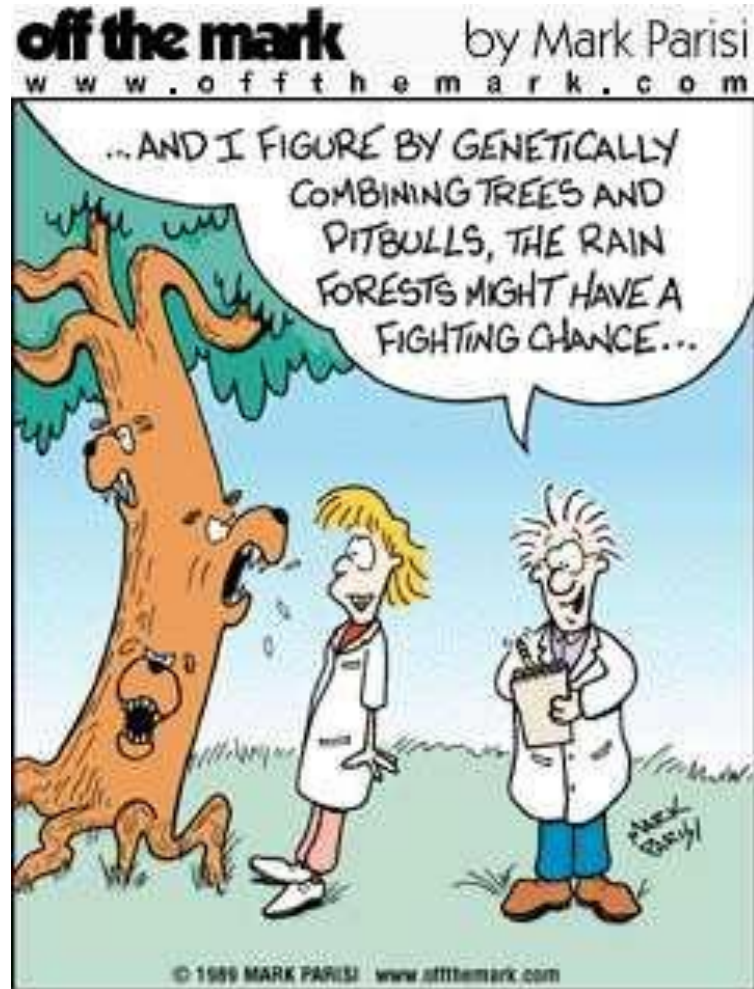


TABLE 5.4 The genetic code

First position (5' end)	Second position				Third position (3' end)
	U	C	A	G	
U	Phe	Ser	Tyr	Cys	U
	Phe	Ser	Tyr	Cys	C
	Leu	Ser	Stop	Stop	A
	Leu	Ser	Stop	Trp	G
C	Leu	Pro	His	Arg	U
	Leu	Pro	His	Arg	C
	Leu	Pro	Gln	Arg	A
	Leu	Pro	Gln	Arg	G
A	Ile	Thr	Asn	Ser	U
	Ile	Thr	Asn	Ser	C
	Ile	Thr	Lys	Arg	A
	Met	Thr	Lys	Arg	G
G	Val	Ala	Asp	Gly	U
	Val	Ala	Asp	Gly	C
	Val	Ala	Glu	Gly	A
	Val	Ala	Glu	Gly	G

Note: This table identifies the amino acid encoded by each triplet. For example, the codon 5' AUG 3' on mRNA specifies methionine, whereas CAU specifies histidine. UAA, UAG, and UGA are termination signals. AUG is part of the initiation signal, in addition to coding for internal methionine residues.

Recombinant DNA Technology



Definitions

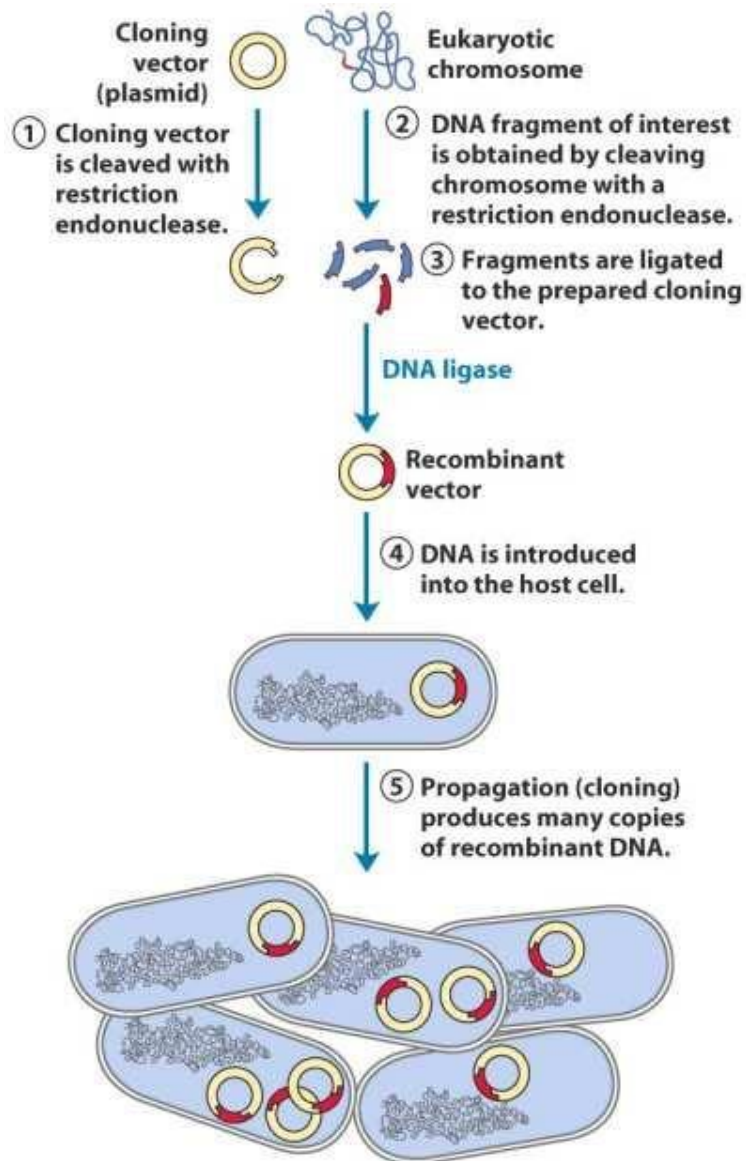
Recombinant DNA, a DNA construct created by fusing different fragments of DNA

Genetic Engineering, the deliberate alteration of DNA through the creation of recombinant DNA

Genetically Modified Organism, a living entity modified through genetic engineering

Transgenic, a genetically modified organism containing DNA from another source

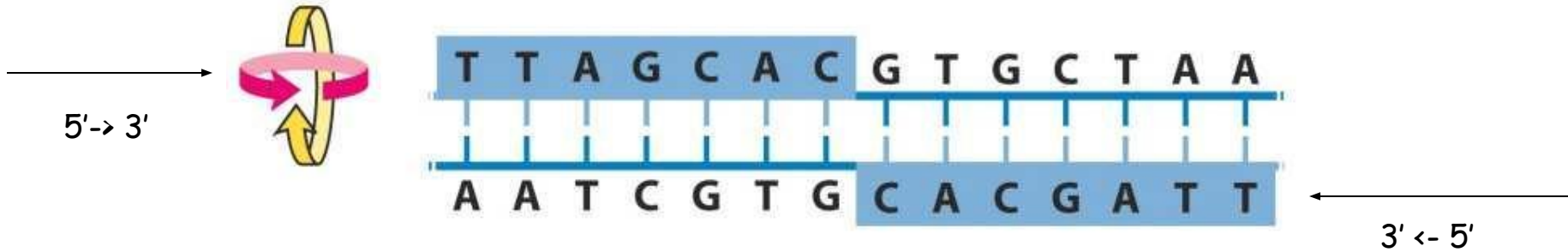
Recombinant DNA Technology



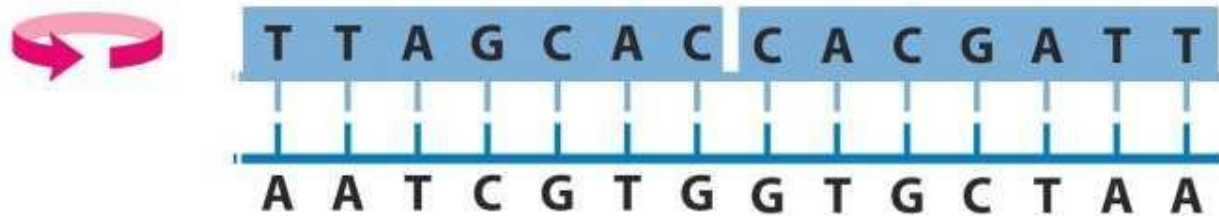
Clones -> Cells or organisms with identical DNA

Restriction endonucleases

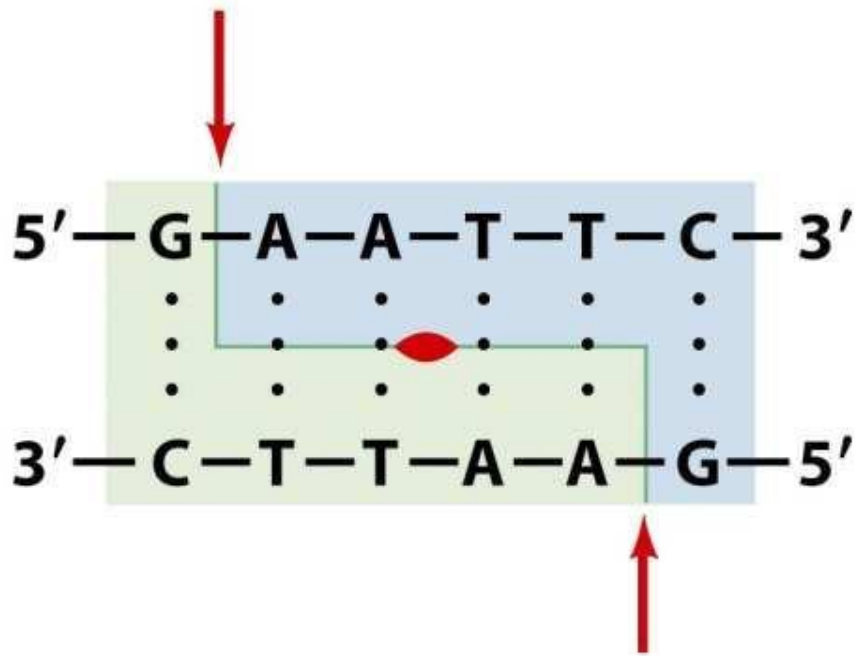
Palindrome



Mirror repeat

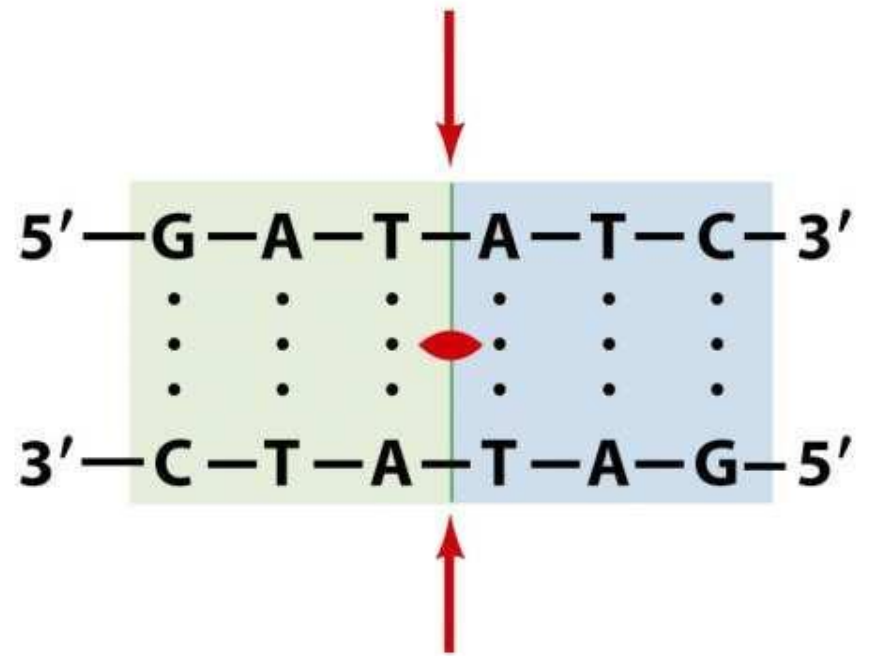


(a) *EcoRI*



 **Cleavage site**

(b) *EcoRV*



 **Twofold symmetry axis**

Figure 3-16 Fundamentals of Biochemistry, 2/e
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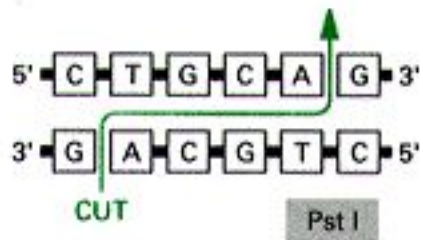
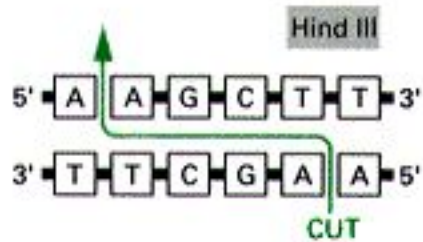
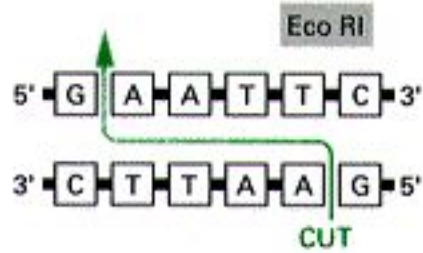
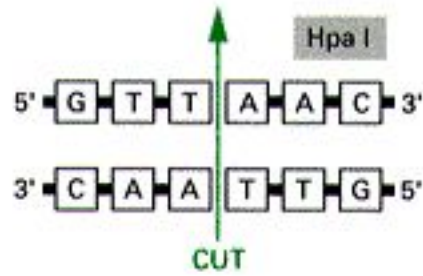


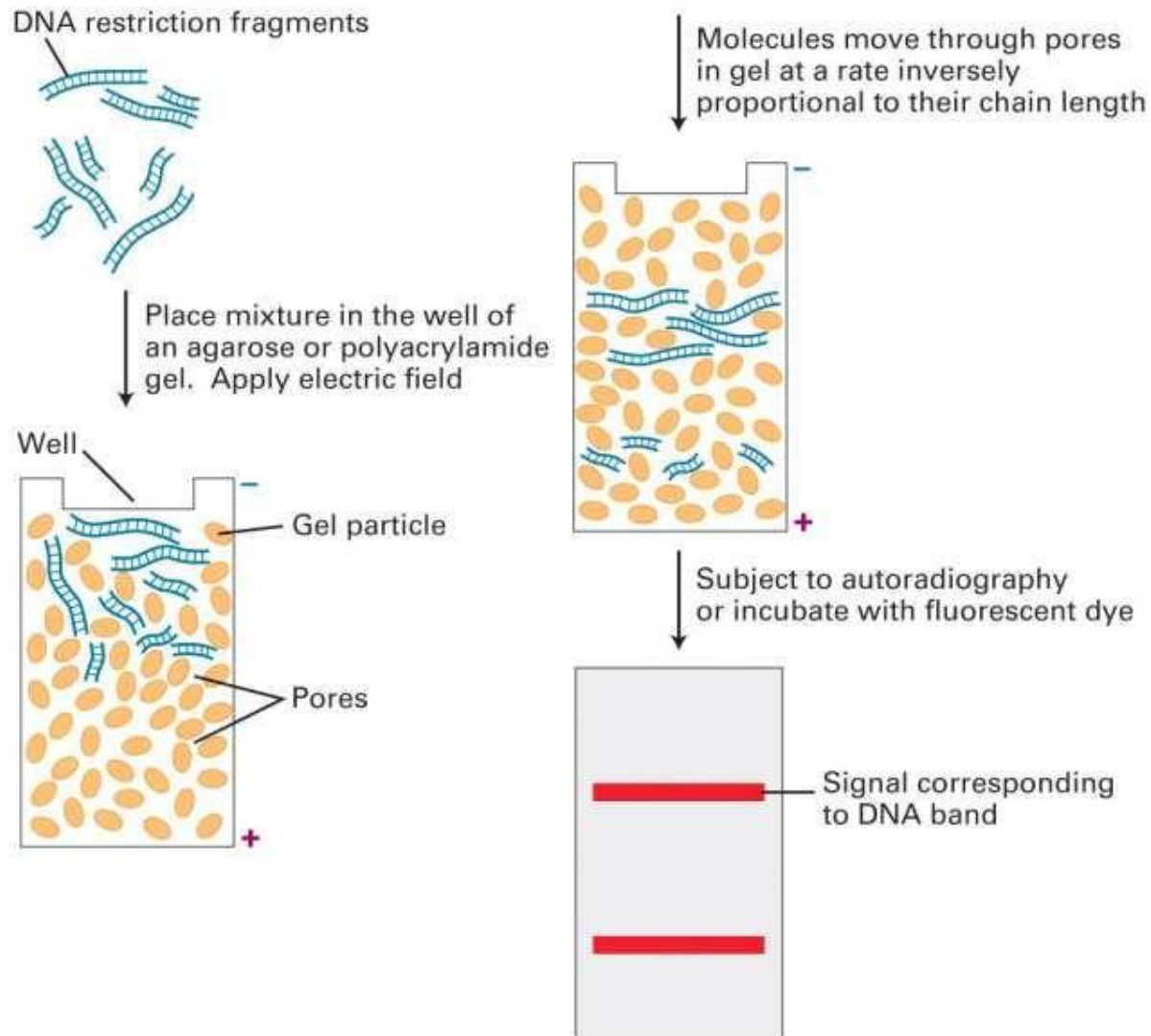
Table 3-2 Recognition and Cleavage Sites of Some Restriction Enzymes

Enzyme	Recognition Sequence ^a	Microorganism
<i>Alu</i> I	AG↓CT	<i>Arthrobacter luteus</i>
<i>Bam</i> HI	G↓GATCC	<i>Bacillus amyloliquefaciens</i> H
<i>Bgl</i> I	GCCNNNNN↓NGGC	<i>Bacillus globigii</i>
<i>Bgl</i> II	A↓GATCT	<i>Bacillus globigii</i>
<i>Eco</i> RI	G↓AATTC	<i>Escherichia coli</i> RY13
<i>Eco</i> RII	↓CC(↓)GG	<i>Escherichia coli</i> R245
<i>Eco</i> RV	GAT↓ATC	<i>Escherichia coli</i> J62 pLG74
<i>Hae</i> II	RGCGC↓Y	<i>Haemophilus aegyptius</i>
<i>Hae</i> III	GG↓CC	<i>Haemophilus aegyptius</i>
<i>Hind</i> III	A↓AGCTT	<i>Haemophilus influenzae</i> R _d
<i>Hpa</i> II	C↓CGG	<i>Haemophilus parainfluenzae</i>
<i>Msp</i> I	C↓CGG	<i>Moraxella</i> species
<i>Pst</i> I	CTGCA↓G	<i>Providencia stuartii</i> 164
<i>Pvu</i> II	CAG↓CTG	<i>Proteus vulgaris</i>
<i>Sal</i> I	G↓TCGAC	<i>Streptomyces albus</i> G
<i>Taq</i> I	T↓CGA	<i>Thermus aquaticus</i>
<i>Xho</i> I	C↓TCGAG	<i>Xanthomonas holcicola</i>

^aThe recognition sequence is abbreviated so that only one strand, reading 5' to 3', is given. The cleavage site is represented by an arrow (↓). R, Y, and N represent a purine nucleotide, a pyrimidine nucleotide, and any nucleotide, respectively.

Source: Roberts, R.J. and Macelis, D., REBASE—the restriction enzyme database, <http://rebase.neb.com>.

Gel Electrophoresis



Gel Electrophoresis

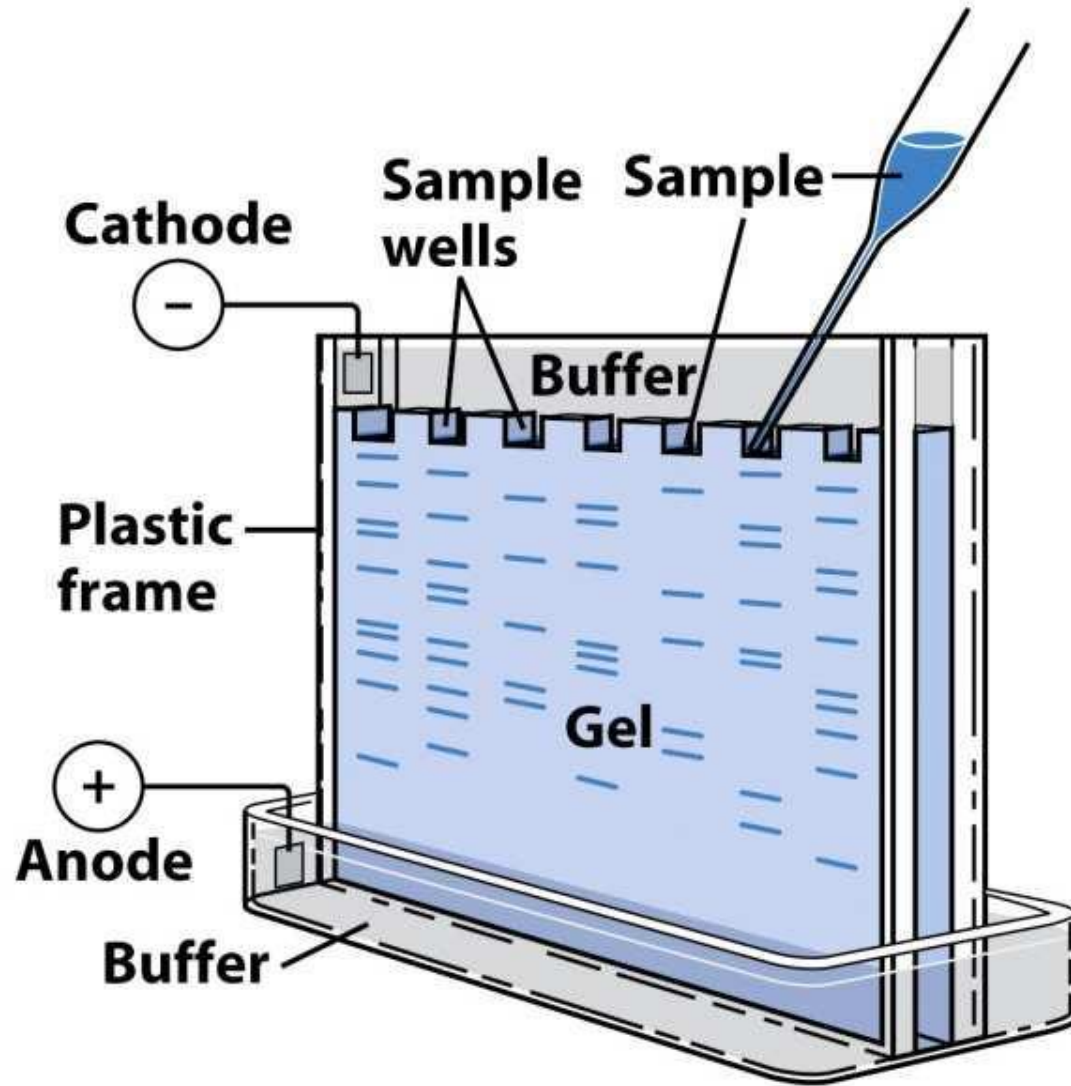
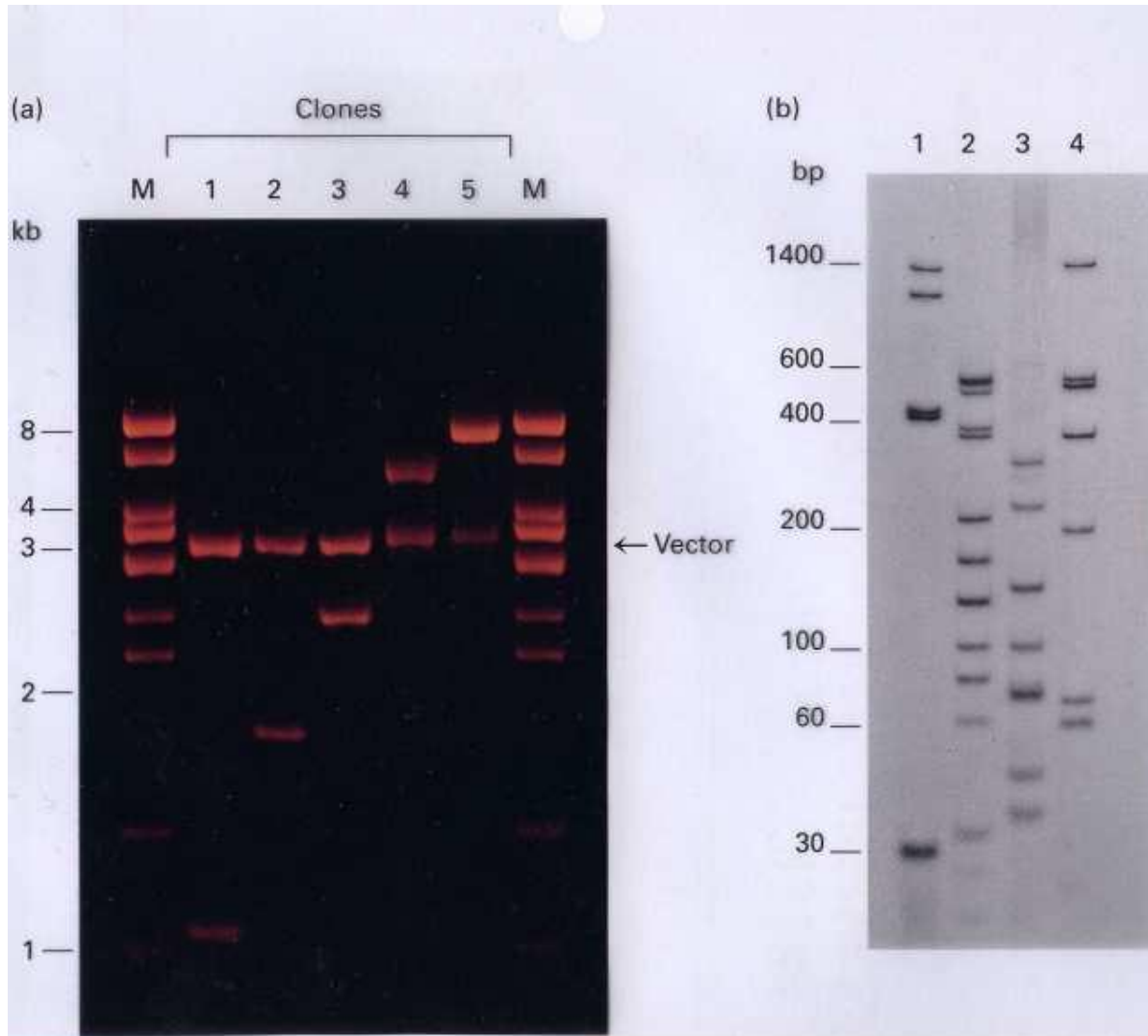
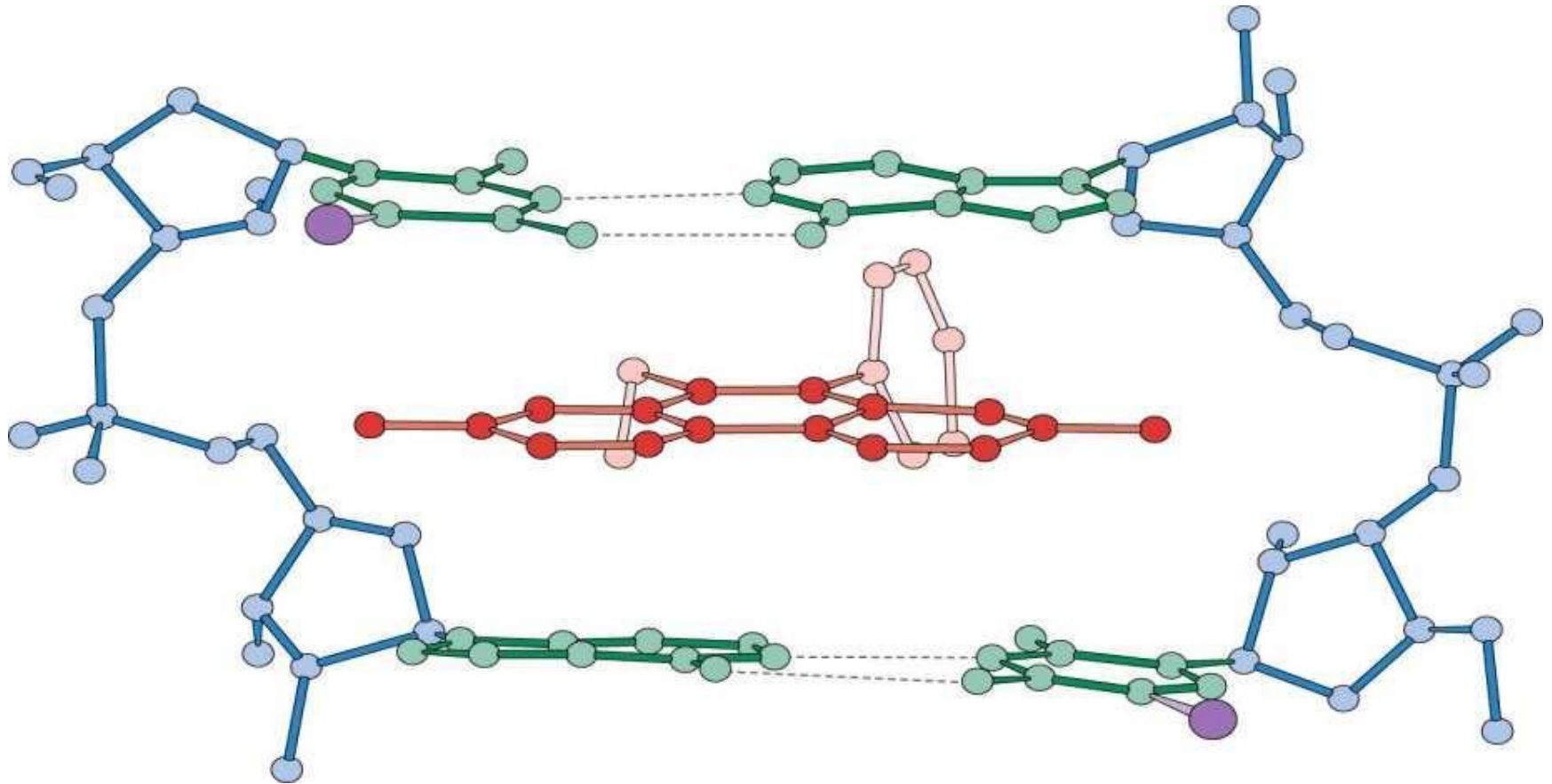


Figure 3-17 Fundamentals of Biochemistry, 2/e
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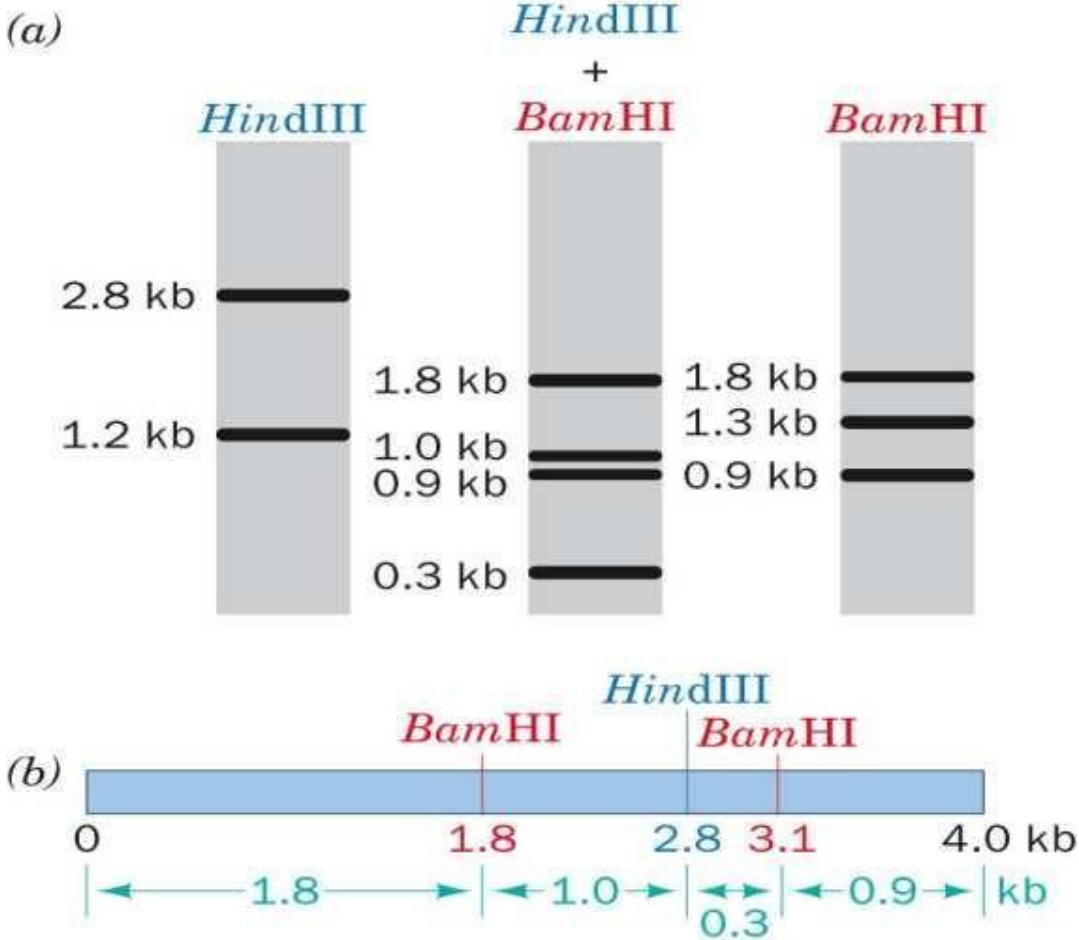
Gel Electrophoresis



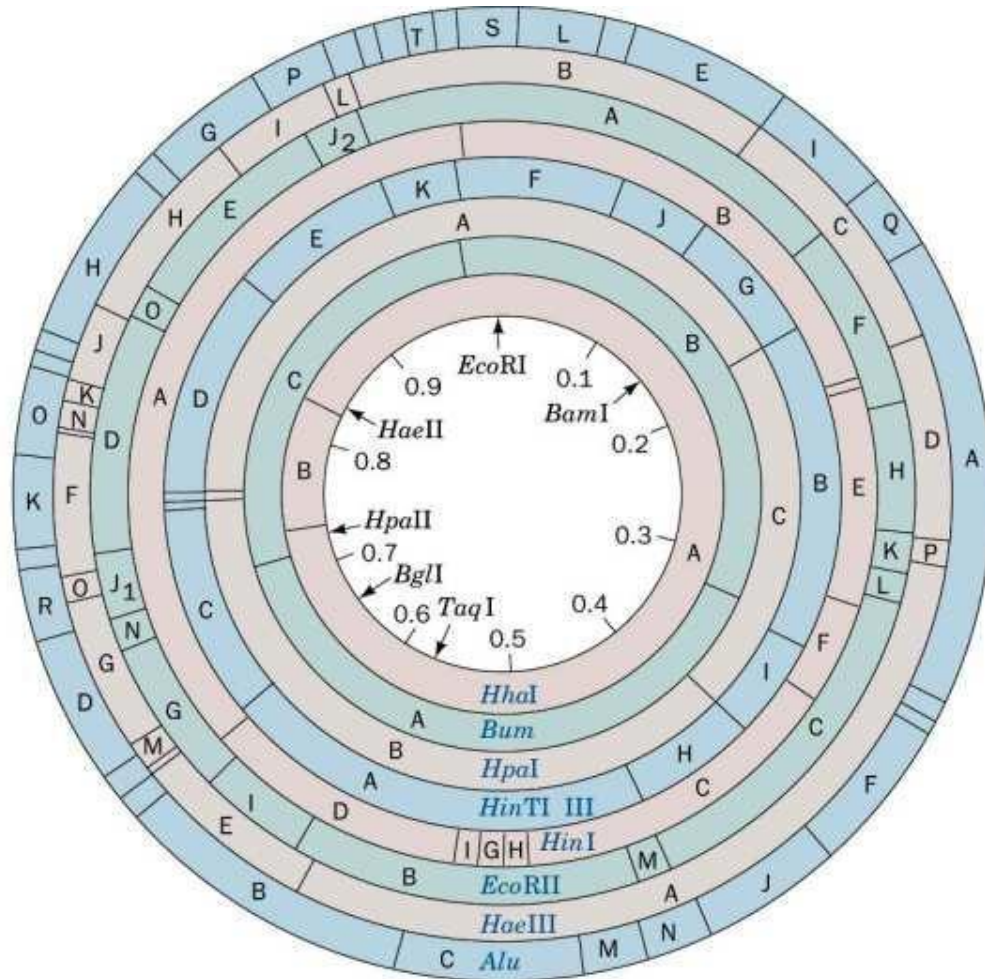
X-Ray structure of a complex of ethidium bromide with DNA.

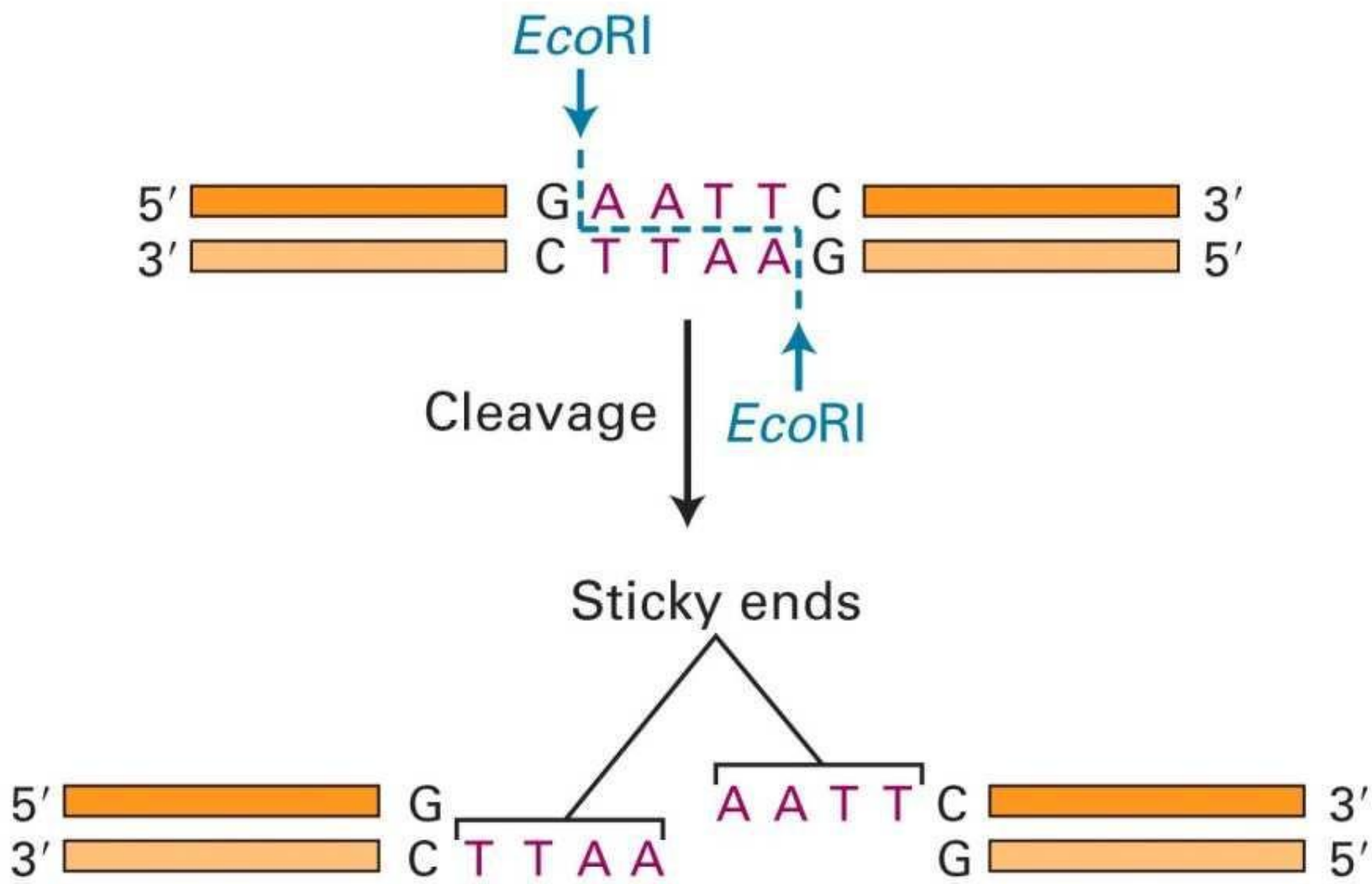


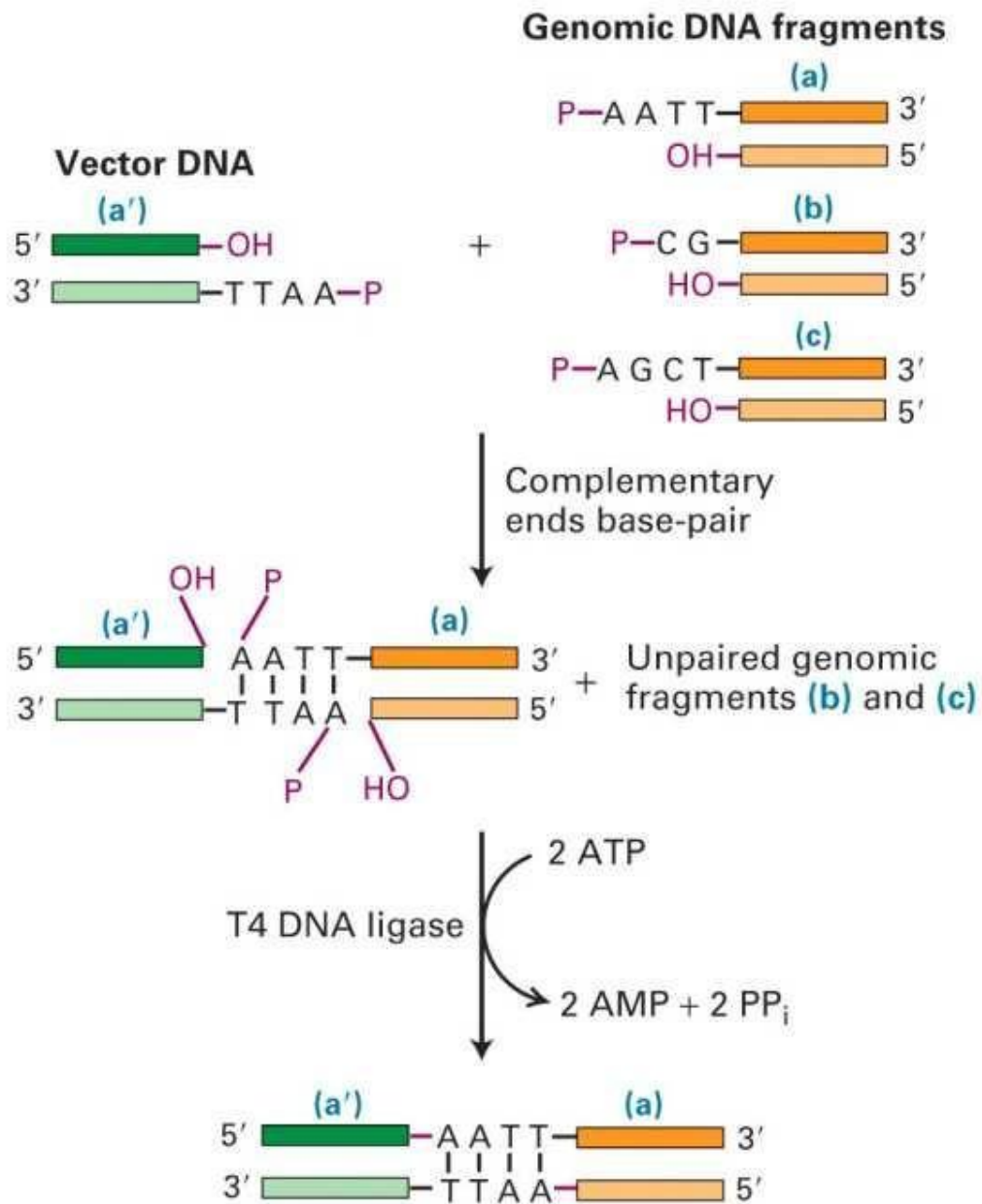
Construction of a restriction map.



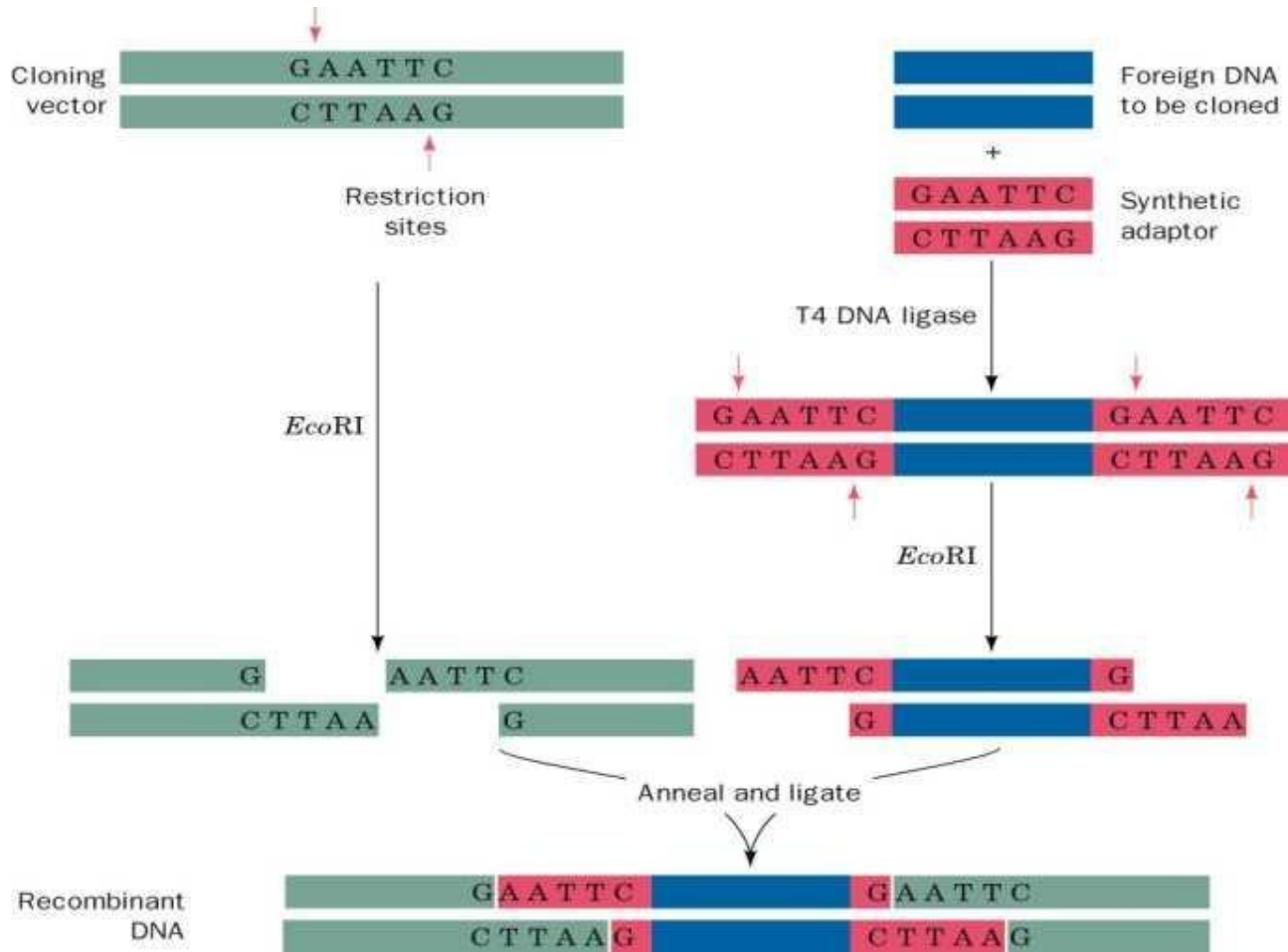
Restriction map for the 5243-bp circular DNA of SV40.

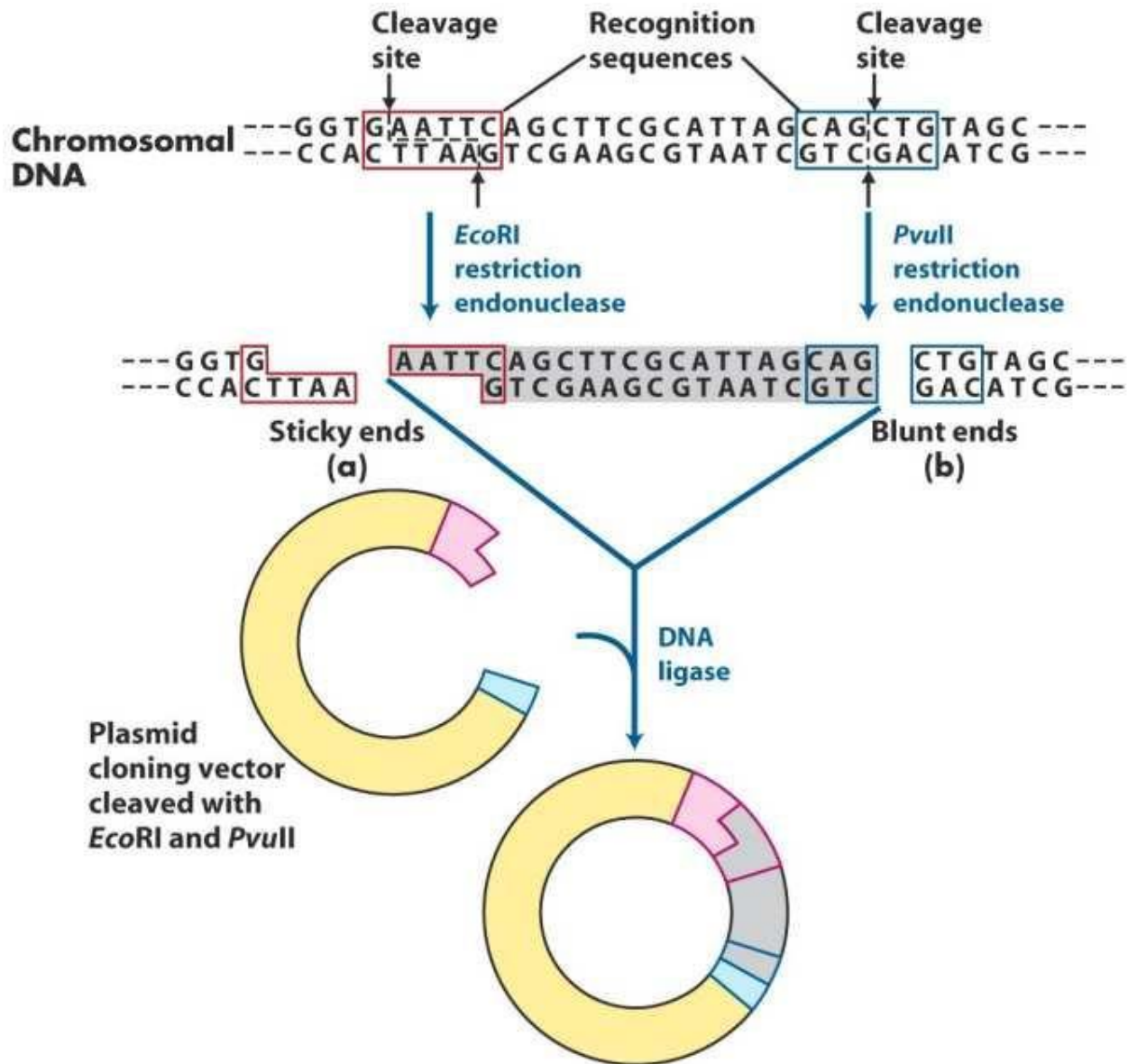




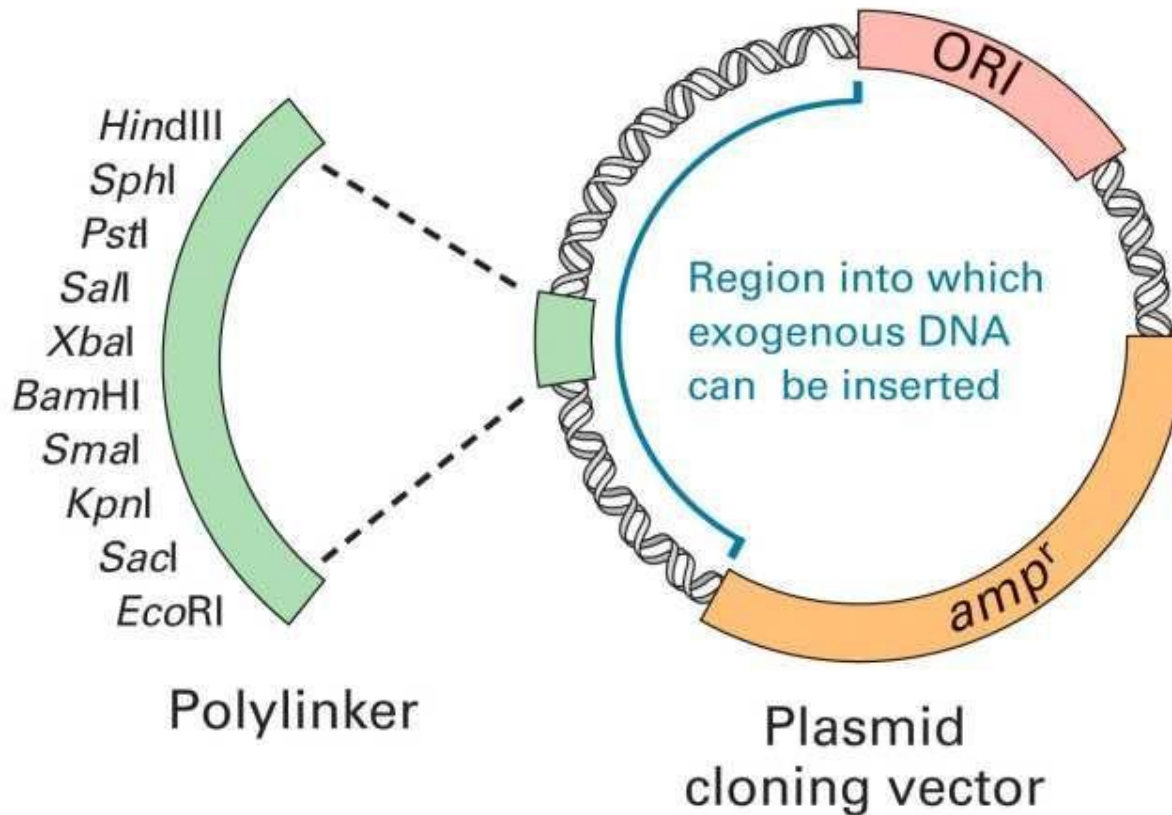


Construction of a recombinant DNA molecule through the use of synthetic oligonucleotide adaptors





Plasmid Cloning Vectors



Plasmid Cloning Vectors

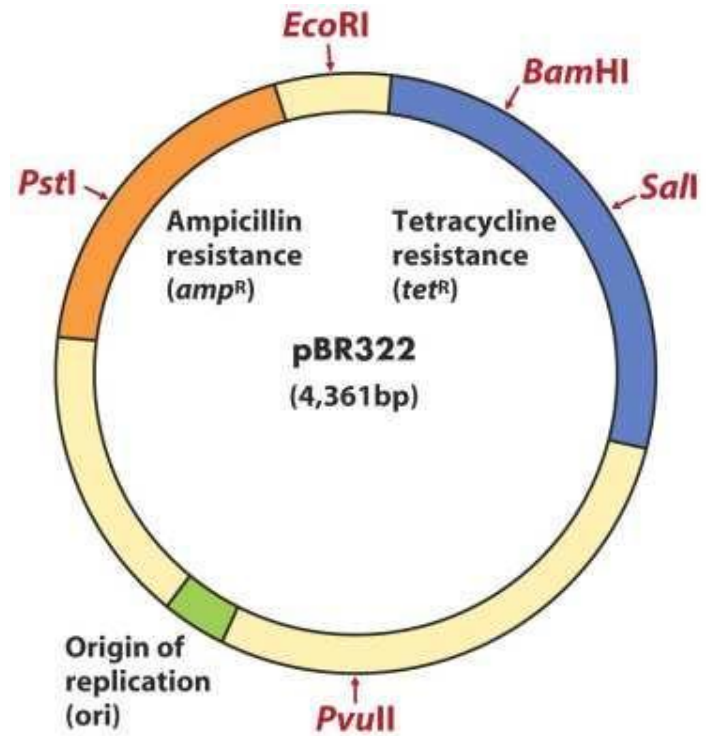
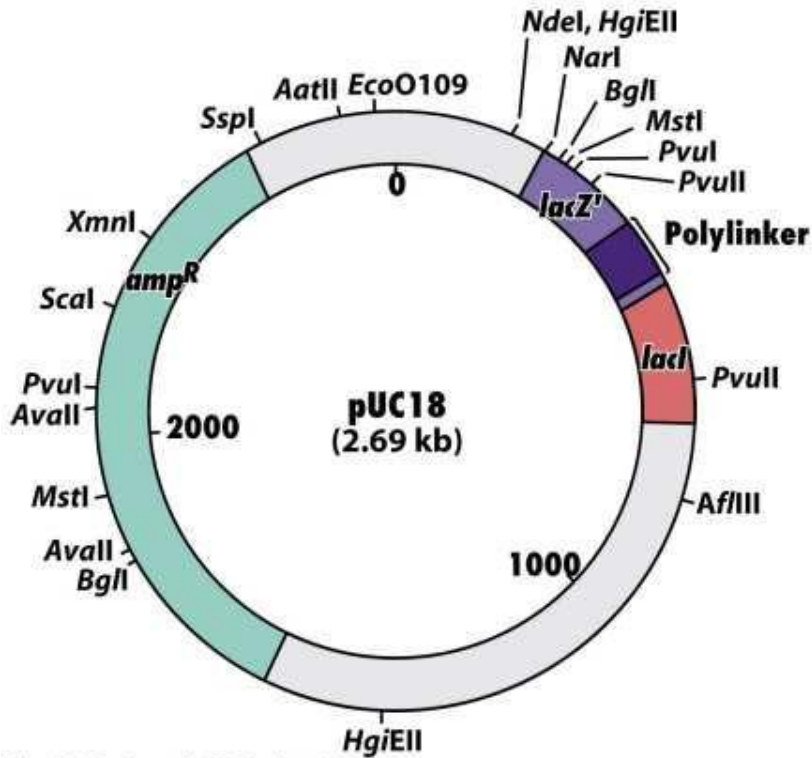
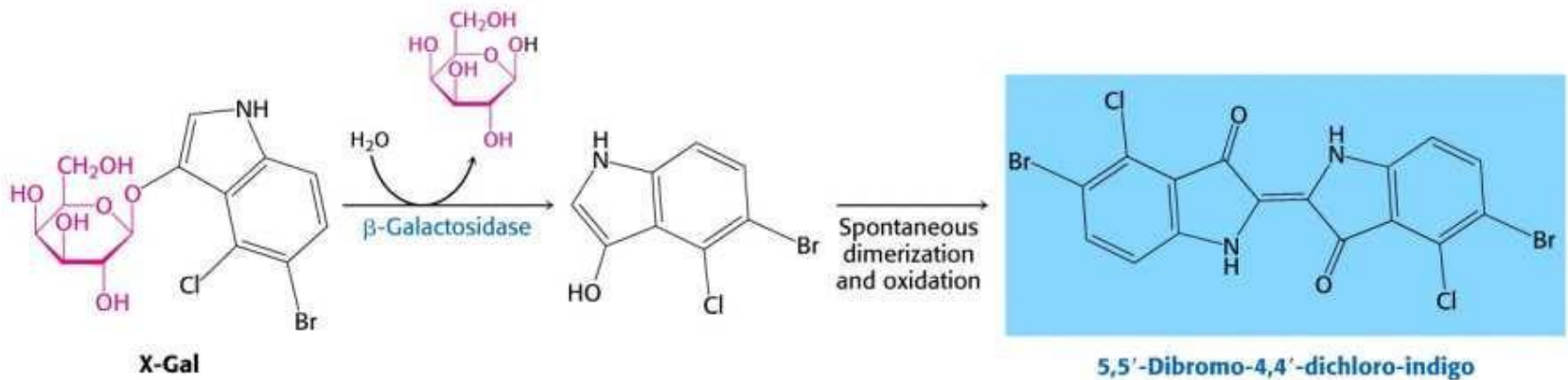
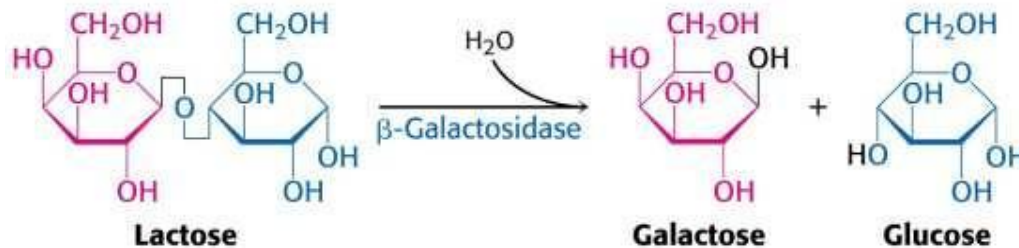


Figure 3-25 Fundamentals of Biochemistry, 2/e
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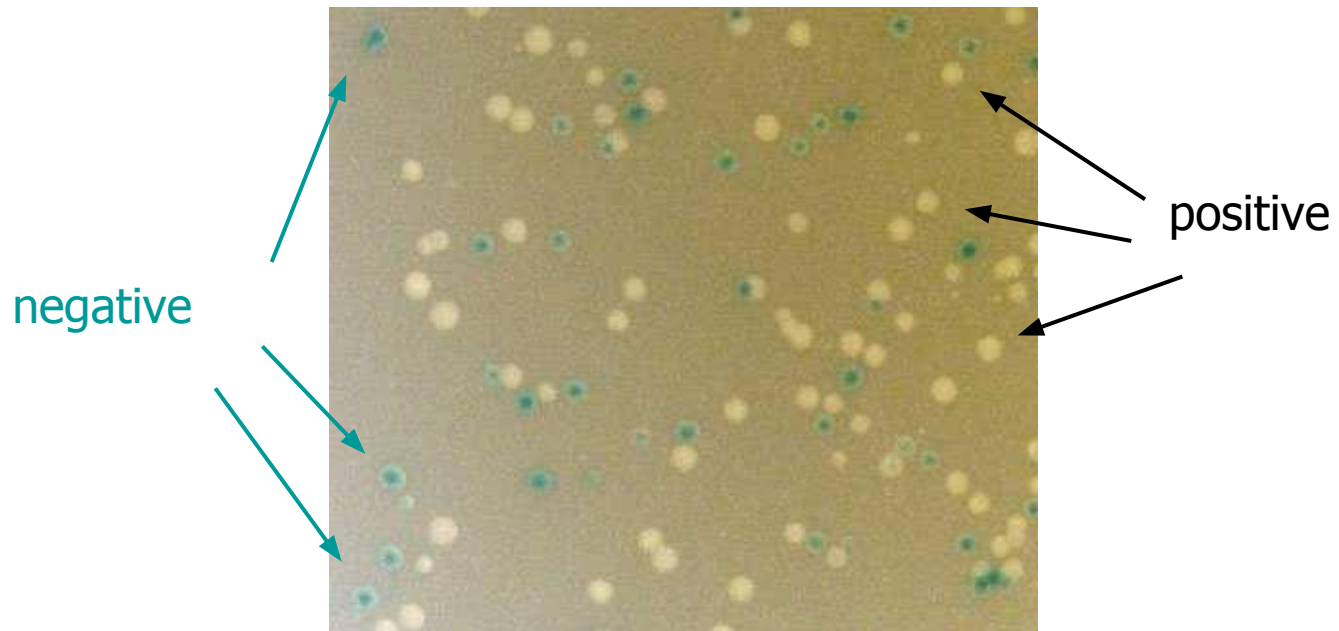
Insertional inactivation

Gene in cloning site:

- *LacZ* → pUC18 (*lacZ* complements the host defect in *lacZ*)
 - pUC18 into host organism → active *lacZ* (β -galactosidase) from plasmid → cleavage of X-gal (blue colonies)
 - gene cloned into polylinker → *lacZ* gene disrupted → no cleavage of X-gal (white colonies)



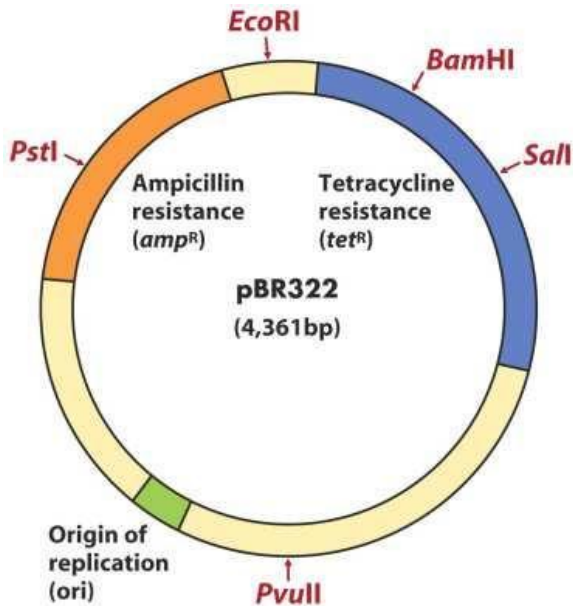
Blue/White Selection



Insertional inactivation

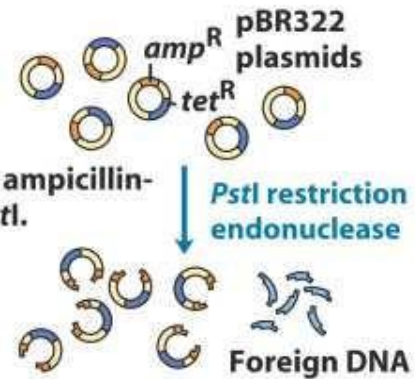
Gene in cloning site:

- Resistance marker -> pBR322 (cloning sites within antibiotic resistance marker)
 - > plasmid into host -> resistance against 2 antibiotics
 - > gene cloned within one resistance marker -> gene for antibiotic resistance marker disrupted -> sensitive against one antibiotic



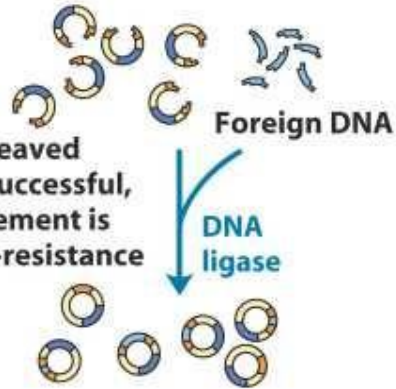
①

pBR322 is cleaved at the ampicillin-resistance element by *PstI*.



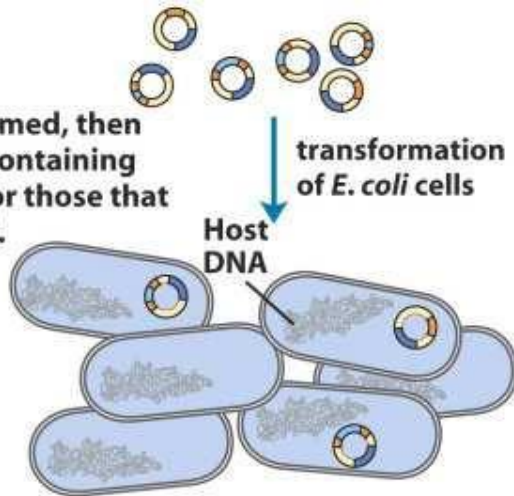
②

Foreign DNA is ligated to cleaved pBR322. Where ligation is successful, the ampicillin-resistance element is disrupted. The tetracycline-resistance element remains intact.



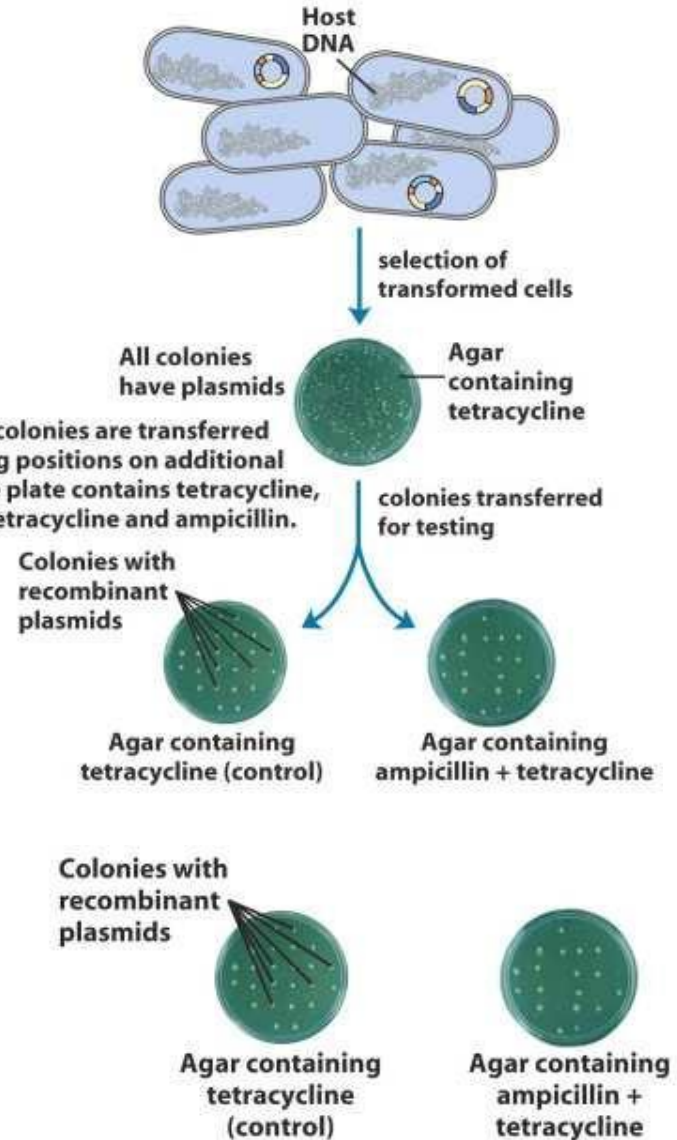
③

E. coli cells are transformed, then grown on agar plates containing tetracycline to select for those that have taken up plasmid.



④

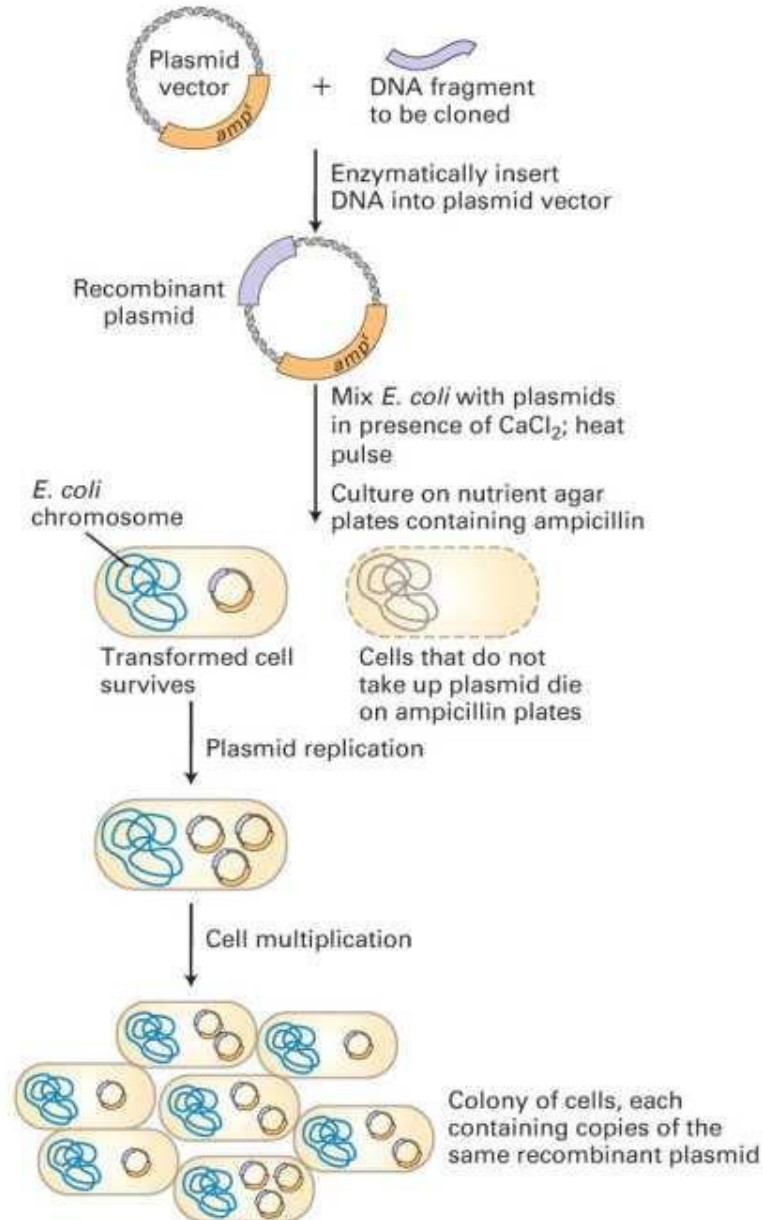
Individual colonies are transferred to matching positions on additional plates. One plate contains tetracycline, the other tetracycline and ampicillin.



⑤

Cells that grow on tetracycline but not on tetracycline + ampicillin contain recombinant plasmids with disrupted ampicillin resistance, hence the foreign DNA. Cells with pBR322 without foreign DNA retain ampicillin resistance and grow on both plates.

Transformation and Selection



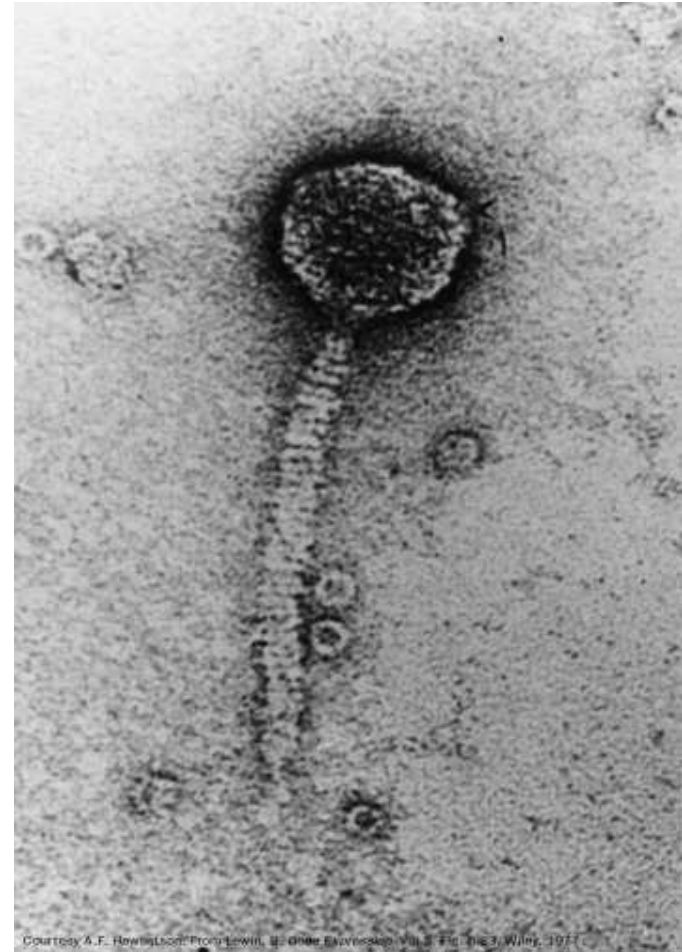
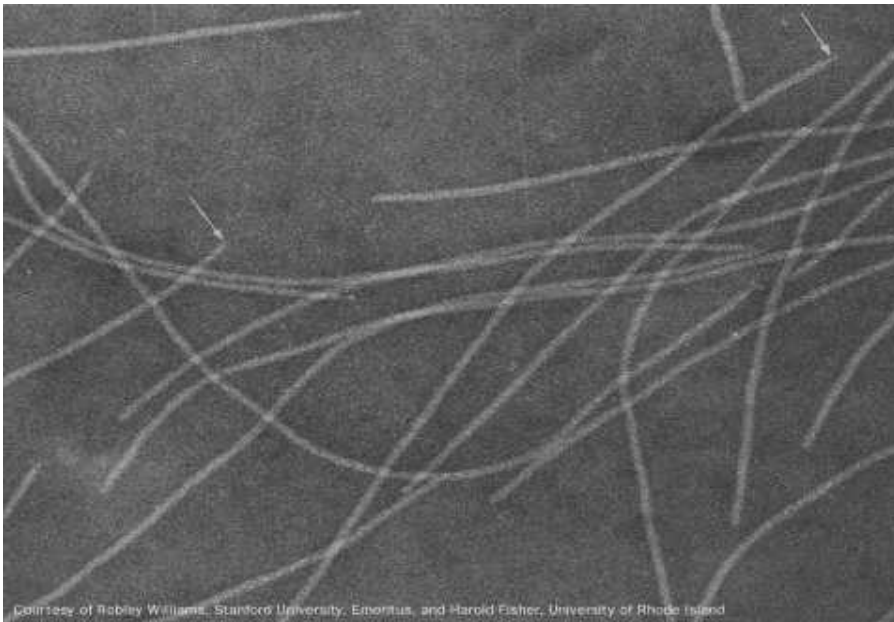
Horizontal gene transfer

- Transformation -> uptake of naked DNA (chemical transformation, electroporation)
- Conjugation -> DNA transfer by cell - cell contact
- Transduction -> DNA transfer by bacteriophage infection

Other methods of Gene transfer -> used with fungi, animal and plant cells:

- Microinjection
- protoplasts

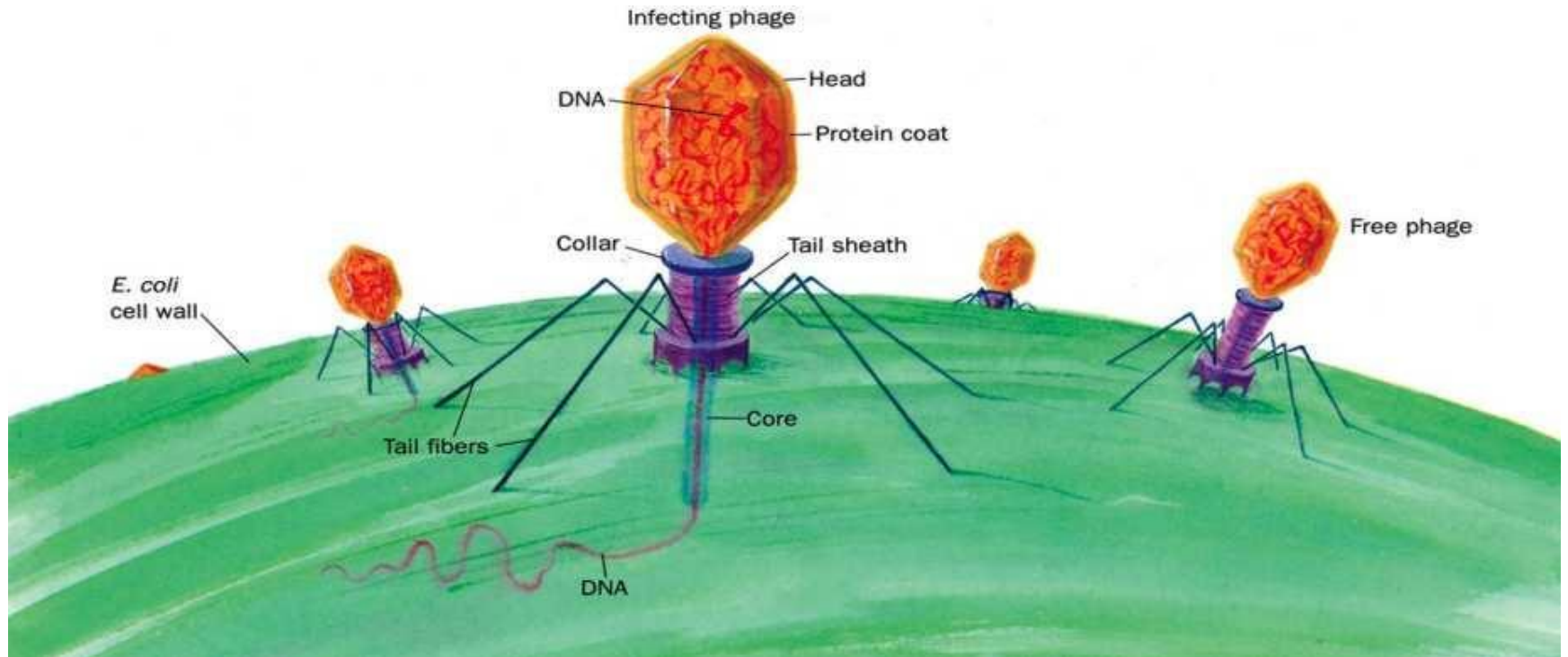
Bacteriophages



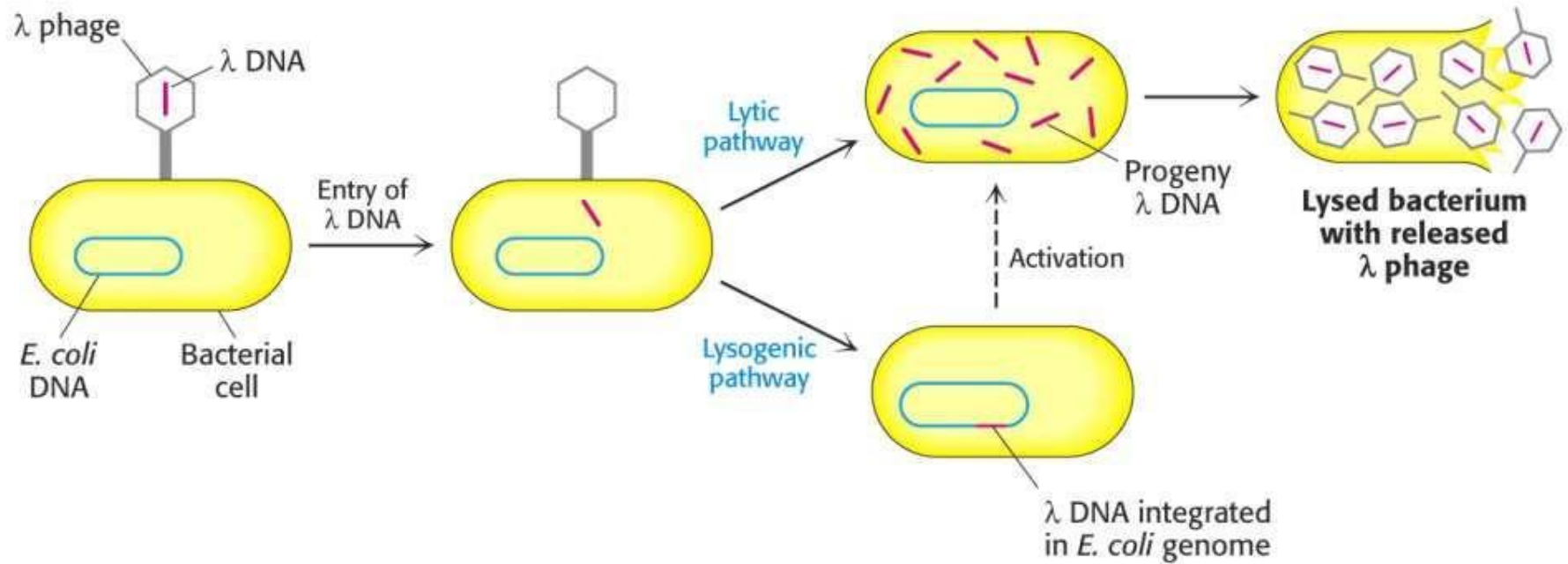
Electron micrograph of the filamentous bacteriophage M13.

Electron micrograph of bacteriophage λ .

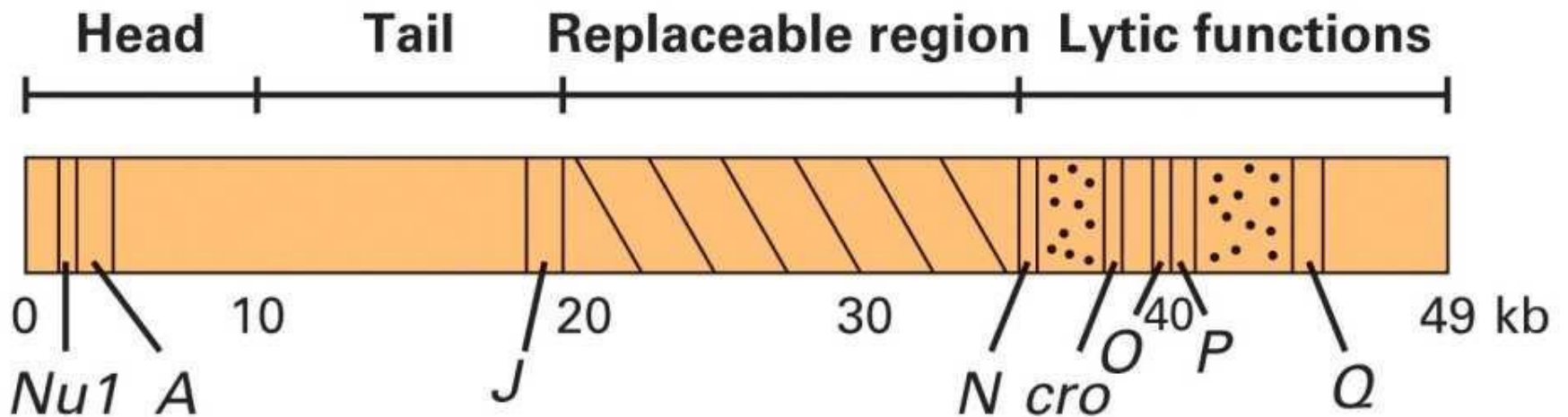
Bacteriophage T2 injecting its DNA into an *E. coli*



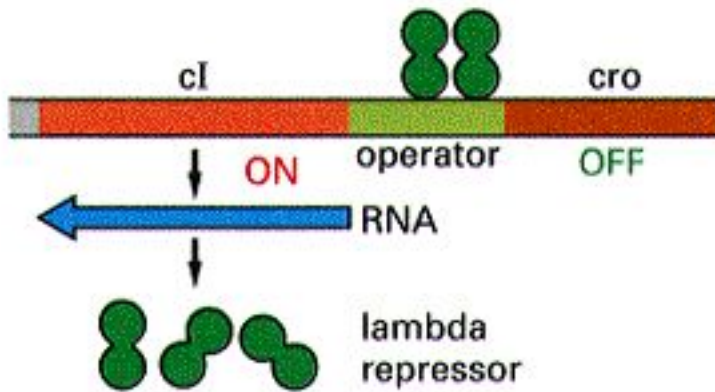
Life Cycle of Bacteriophage



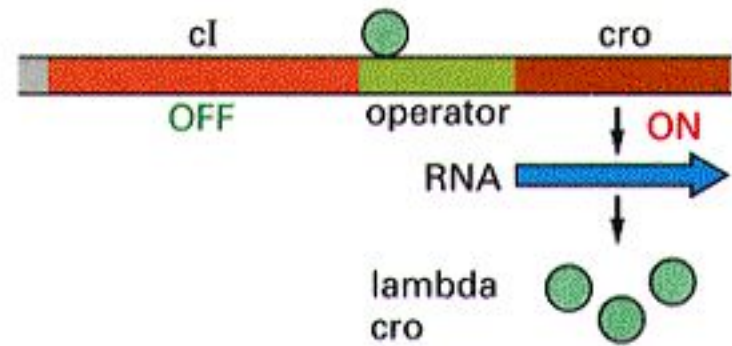
(a) λ Phage genome



stable state 1: the prophage state
lambda repressor protein is made



stable state 2: the lytic state
lambda cro protein is made



Replication of bacteriophage upon infection of a cell

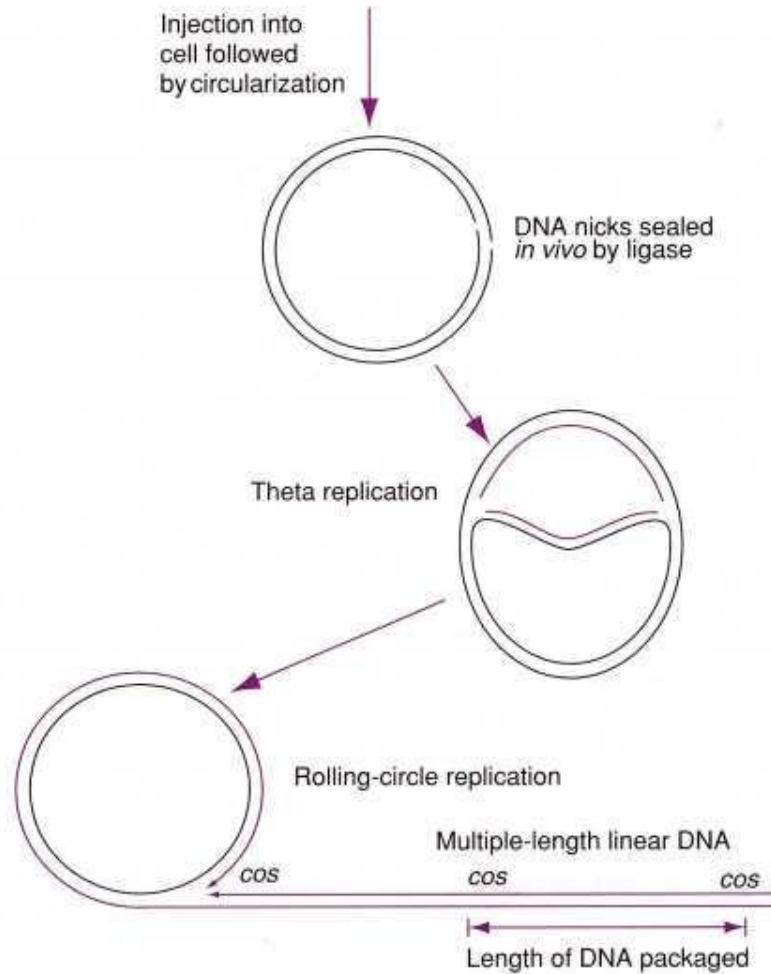
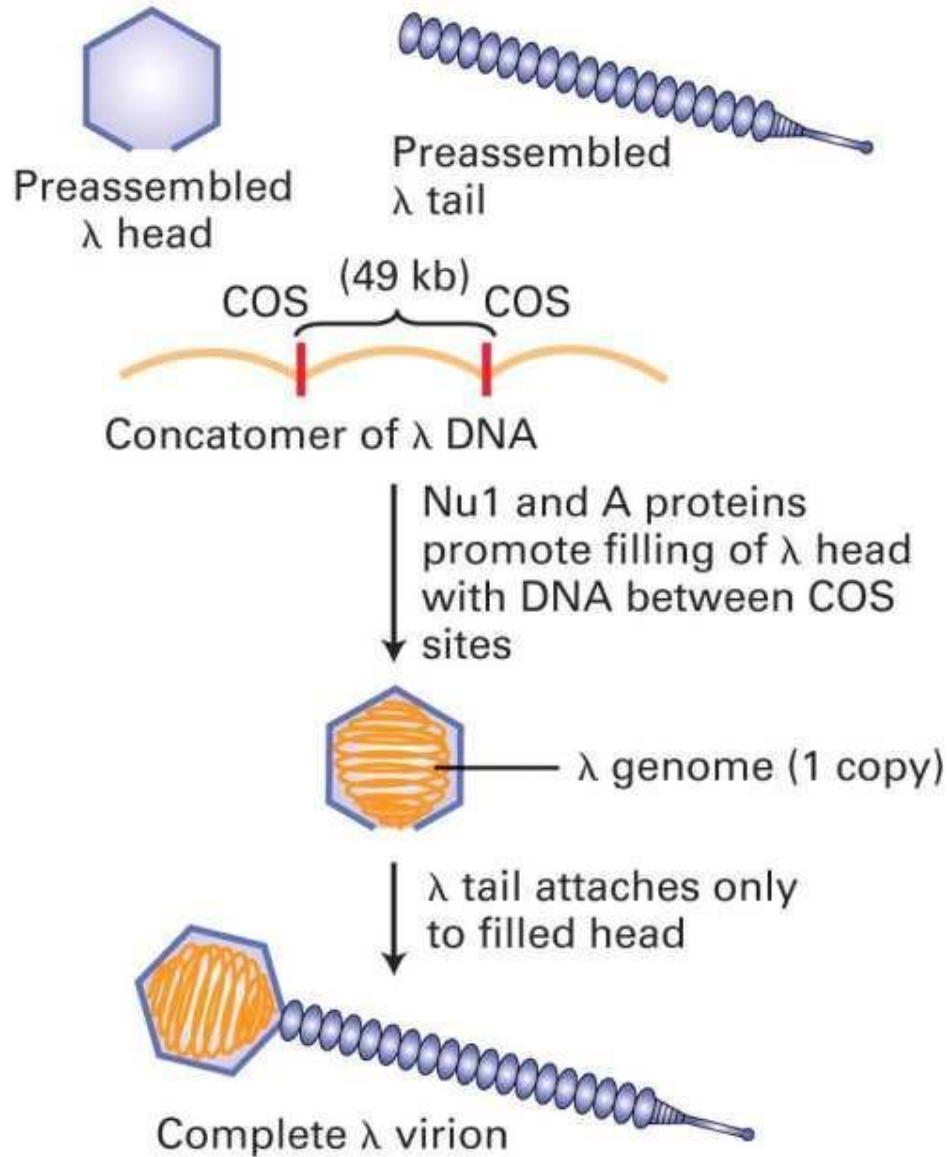
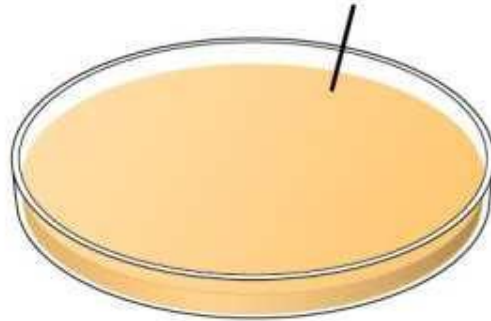


Figure 6.6 Replication of bacteriophage lambda DNA

(b) λ Phage assembly



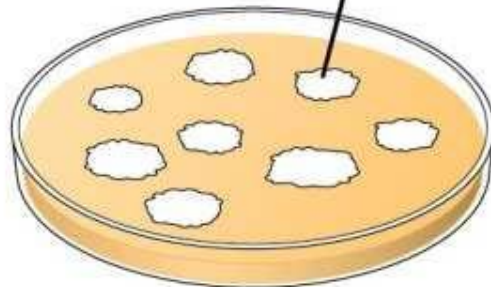
Confluent layer of susceptible host cells
growing on surface of a plate



Add dilute suspension containing virus;
after infection, cover layer of cells
with agar; incubate



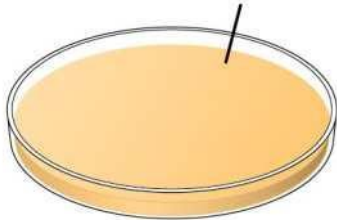
Plaque



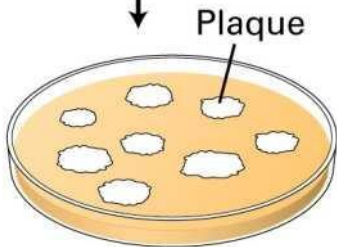
Each plaque represents cell lysis initiated by one viral
particle (agar restricts movement so that virus can
infect only contiguous cells)

Molecular genetics and bacteriophage

Confluent layer of susceptible host cells growing on surface of a plate

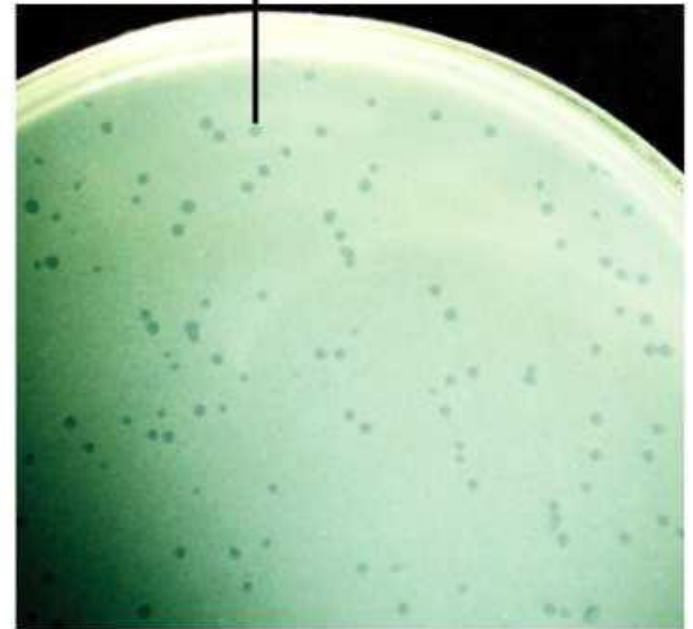


Add dilute suspension containing virus; after infection, cover layer of cells with agar; incubate

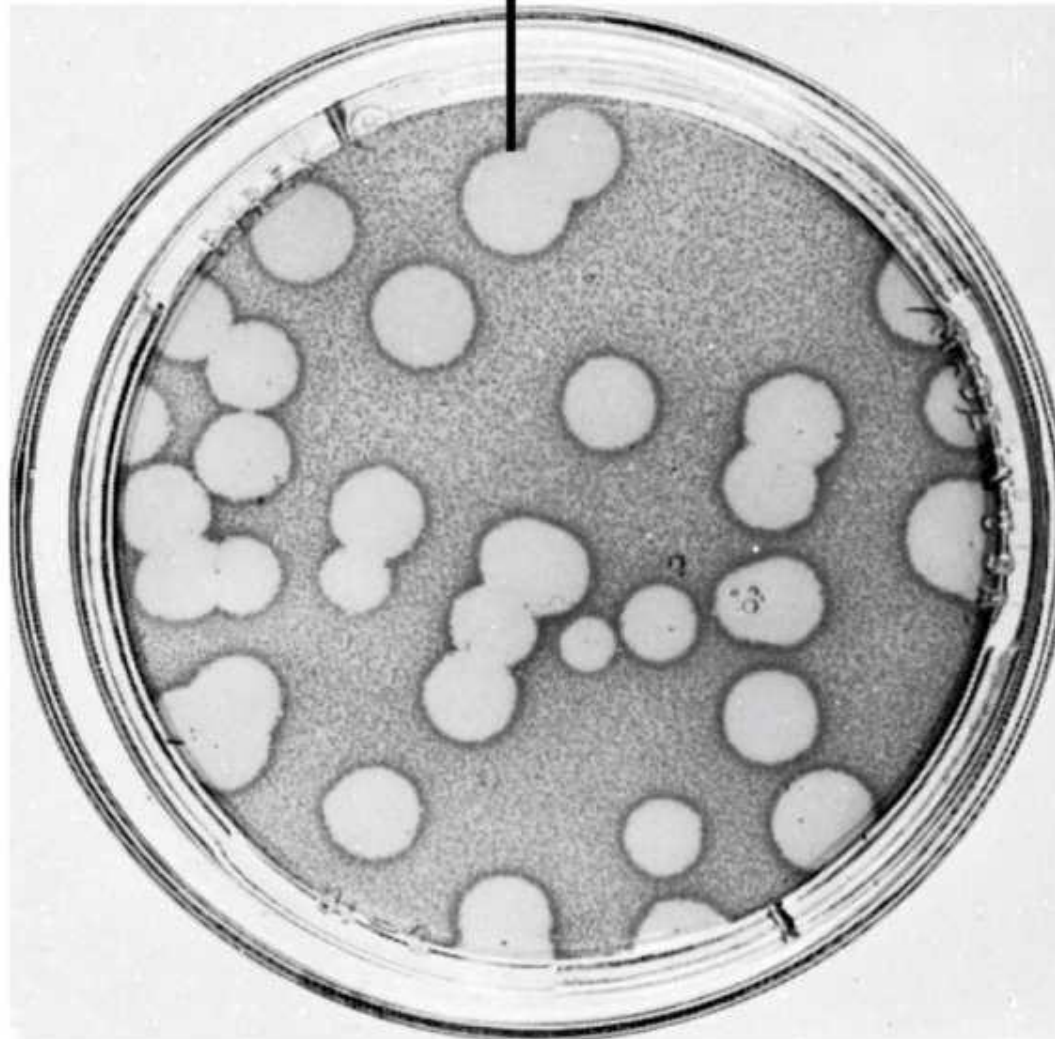


Each plaque represents cell lysis initiated by one viral particle (agar restricts movement so that virus can infect only contiguous cells)

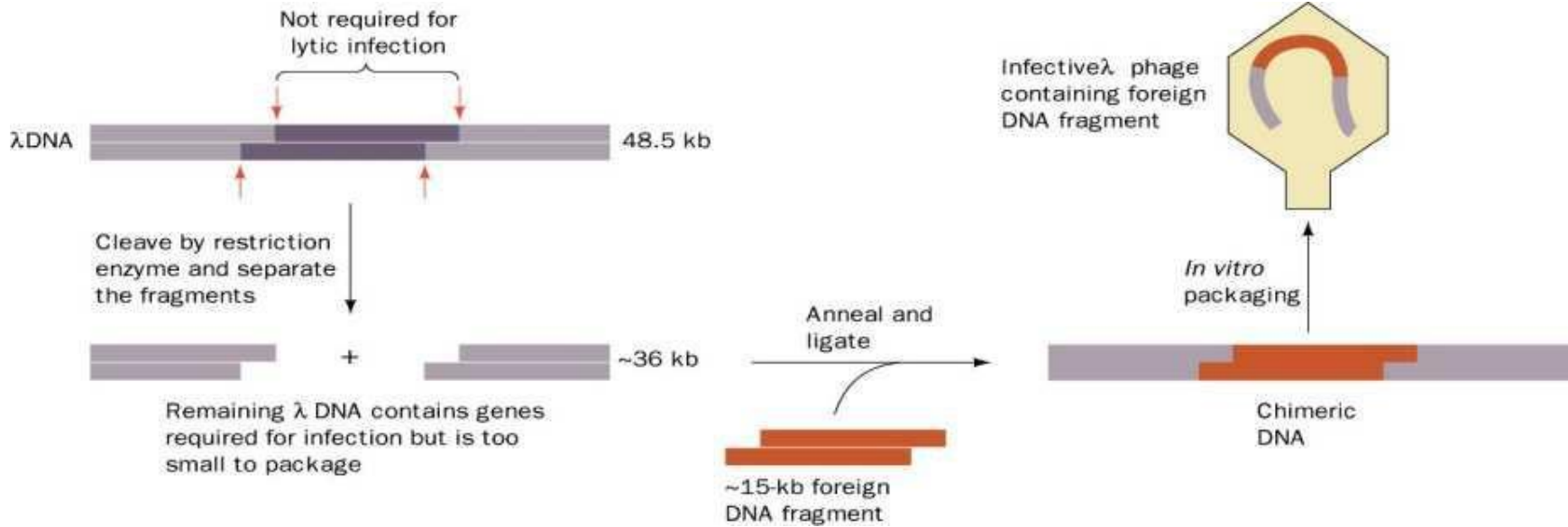
(b) Plaque



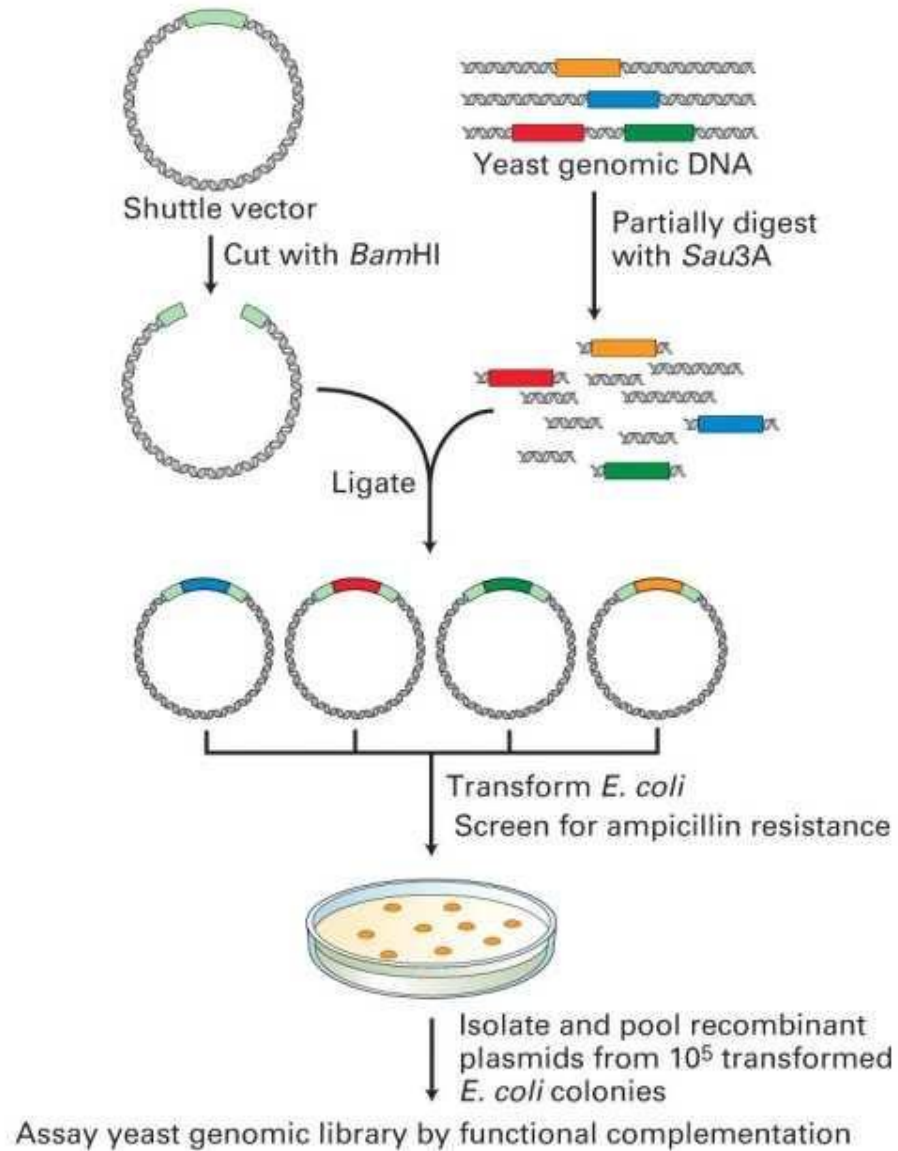
(c) Plaque



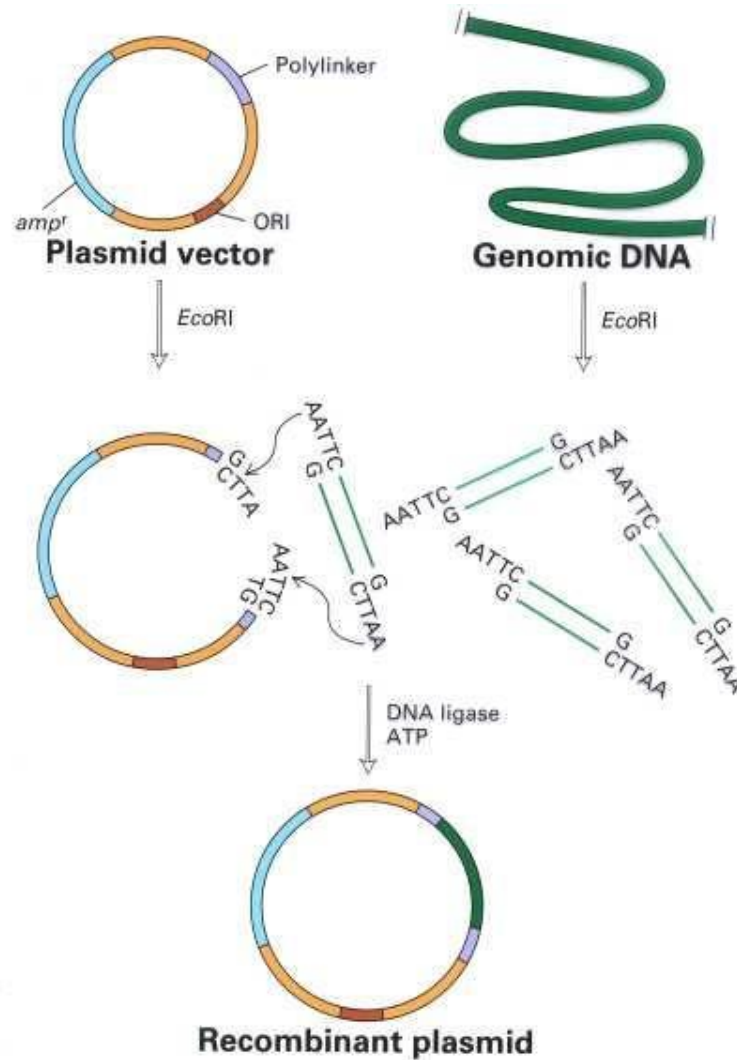
Cloning of foreign DNA in λ phages.



What is a gene library ?



Creation of Libraries



Creation of Libraries

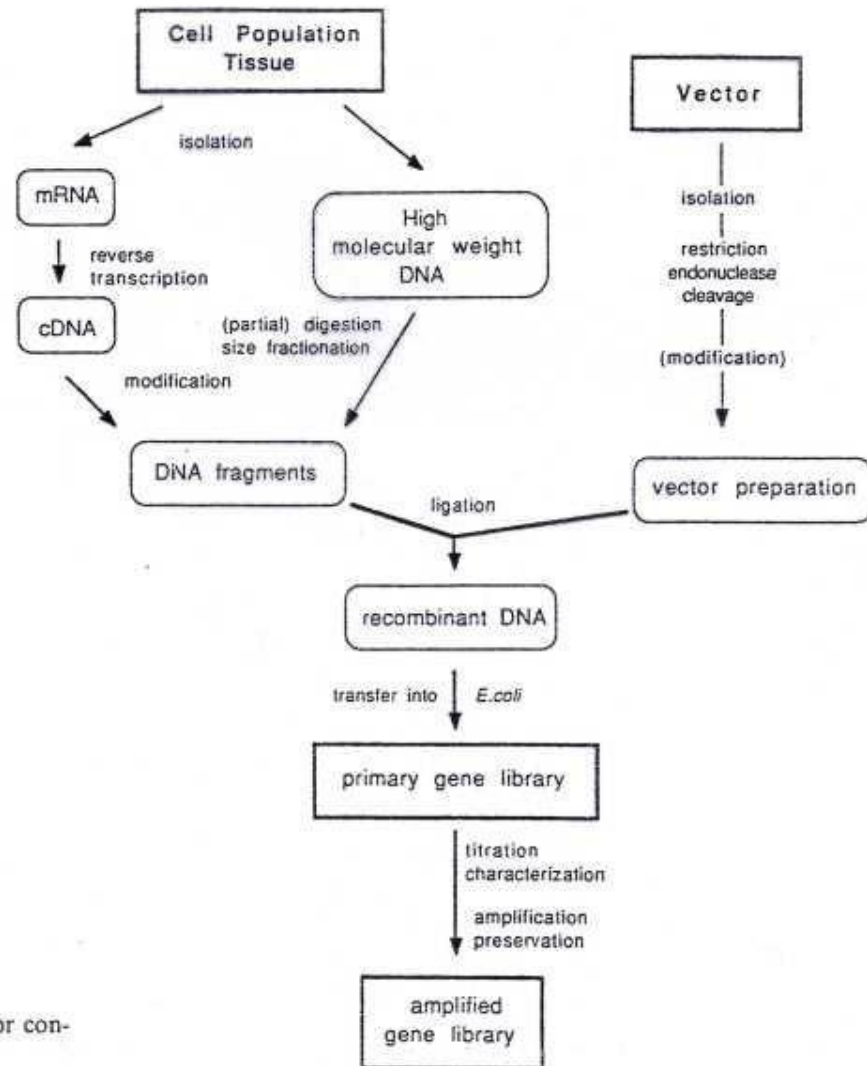


Fig. 6. General scheme for constructing gene libraries.

Sizes of Some DNA Molecules.

Organism	Number of base pairs (kb) ^a	Contour length (μm)
Viruses		
Polyoma, SV40	5.2	1.7
λ Bacteriophage	48.6	17
T2, T4, T6 bacteriophage	166	55
Fowlpox	280	193
Bacteria		
<i>Mycoplasma hominis</i>	760	260
<i>Escherichia coli</i>	4,600	1,600
Eukaryotes		
Yeast (in 17 haploid chromosomes)	12,000	4,100
<i>Drosophila</i> (in 4 haploid chromosomes)	180,000	61,000
Human (in 23 haploid chromosomes)	3,200,000	1,100,000
Lungfish (in 19 haploid chromosomes)	102,000,000	35,000,000

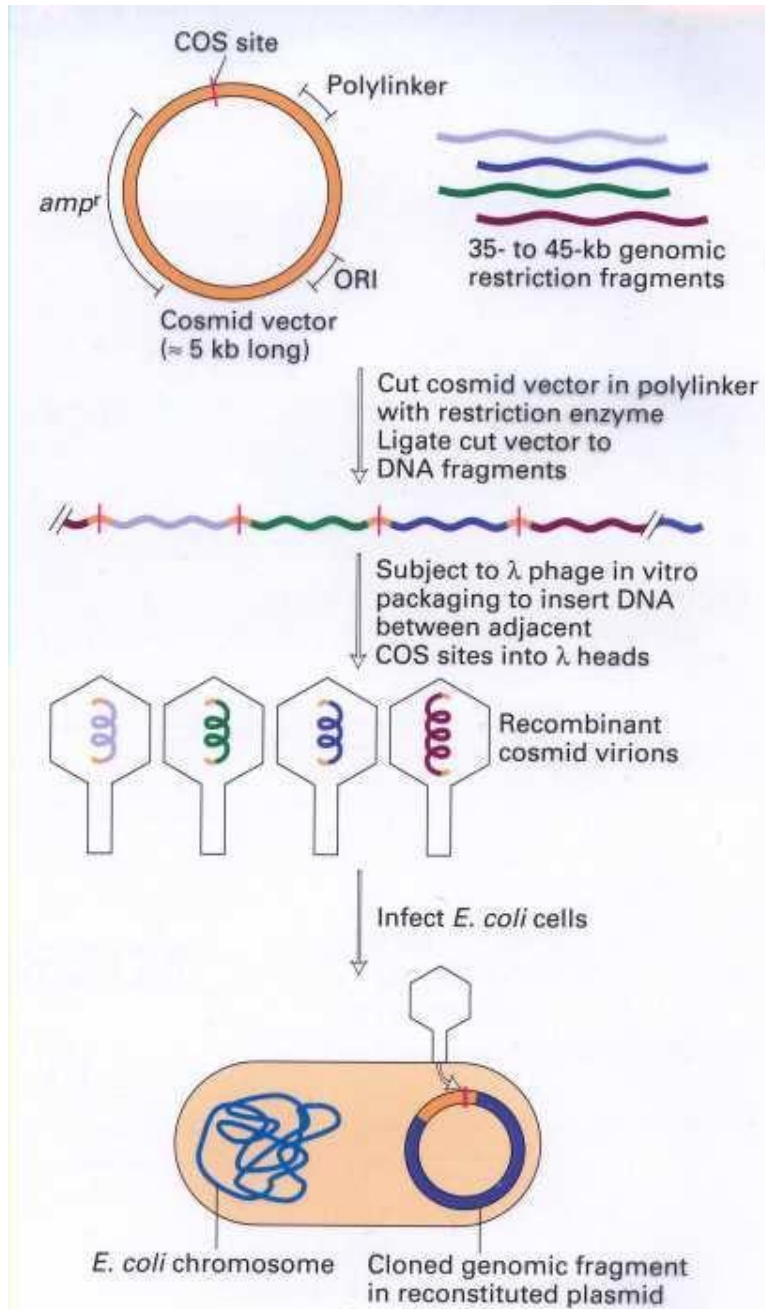
^akb = kilobase pair = 1000 base pairs (bp).

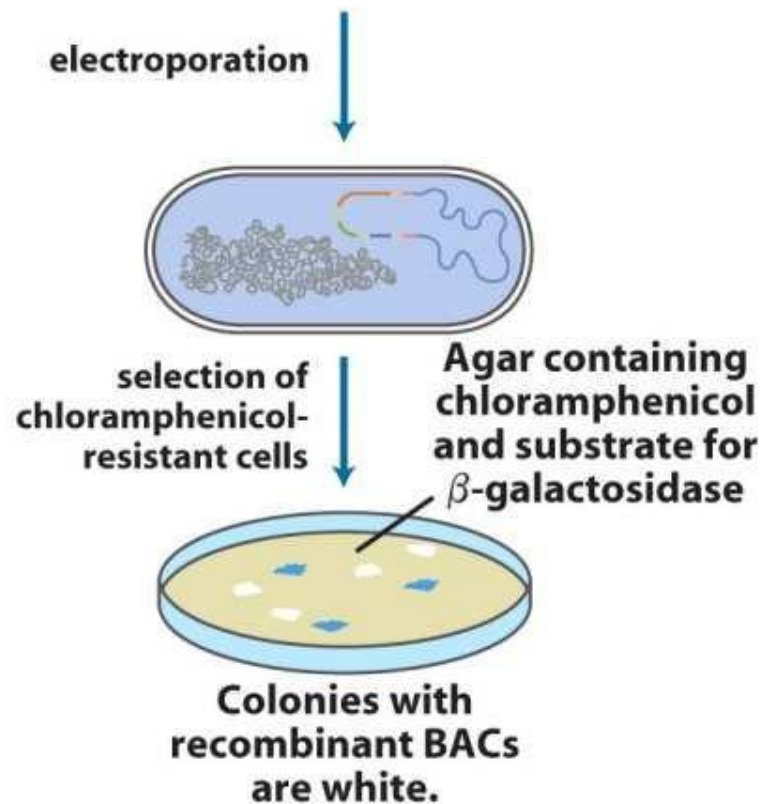
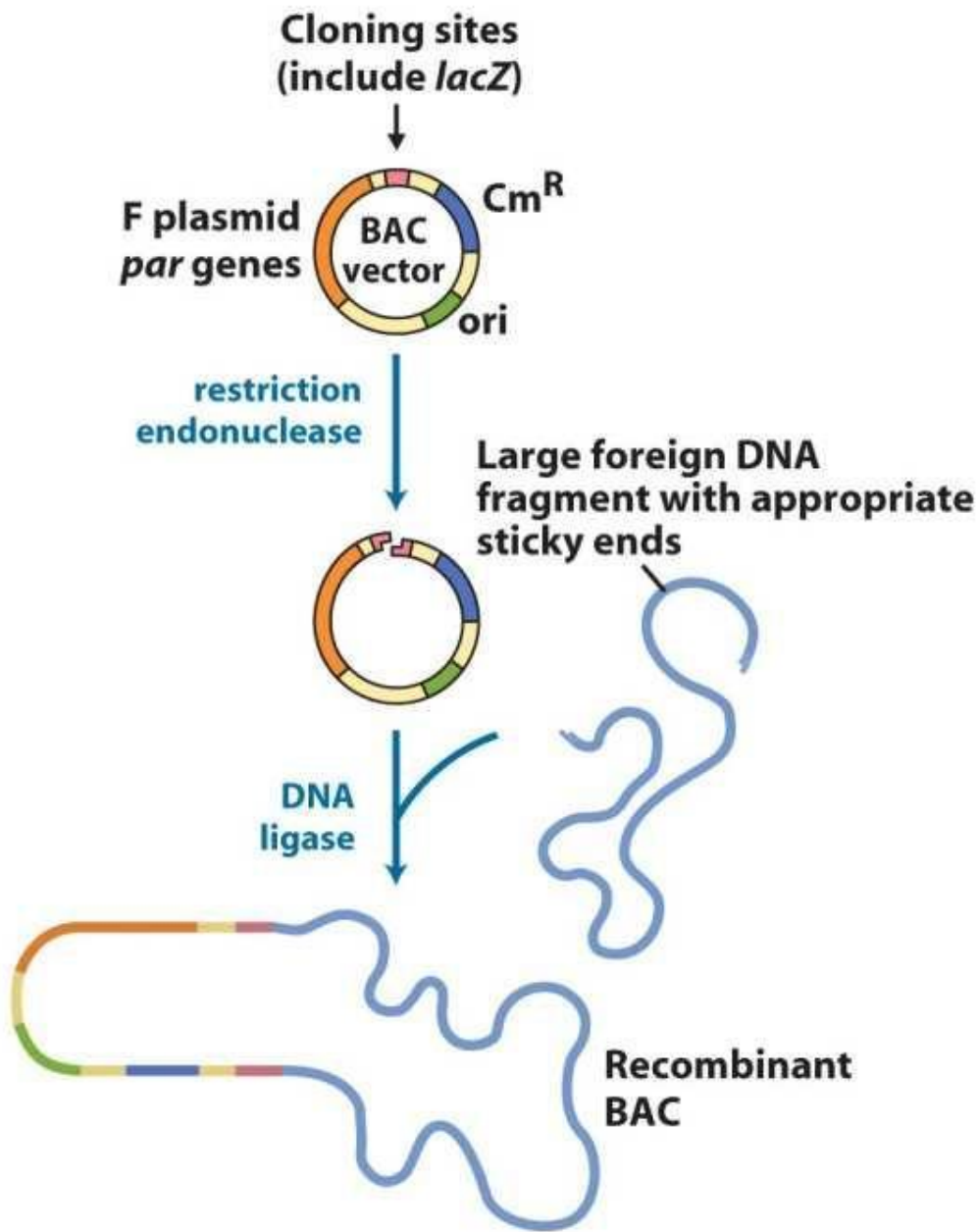
Source: Mainly Kornberg, A. and Baker, T.A., *DNA Replication* (2nd ed.), p. 20, Freeman (1992).

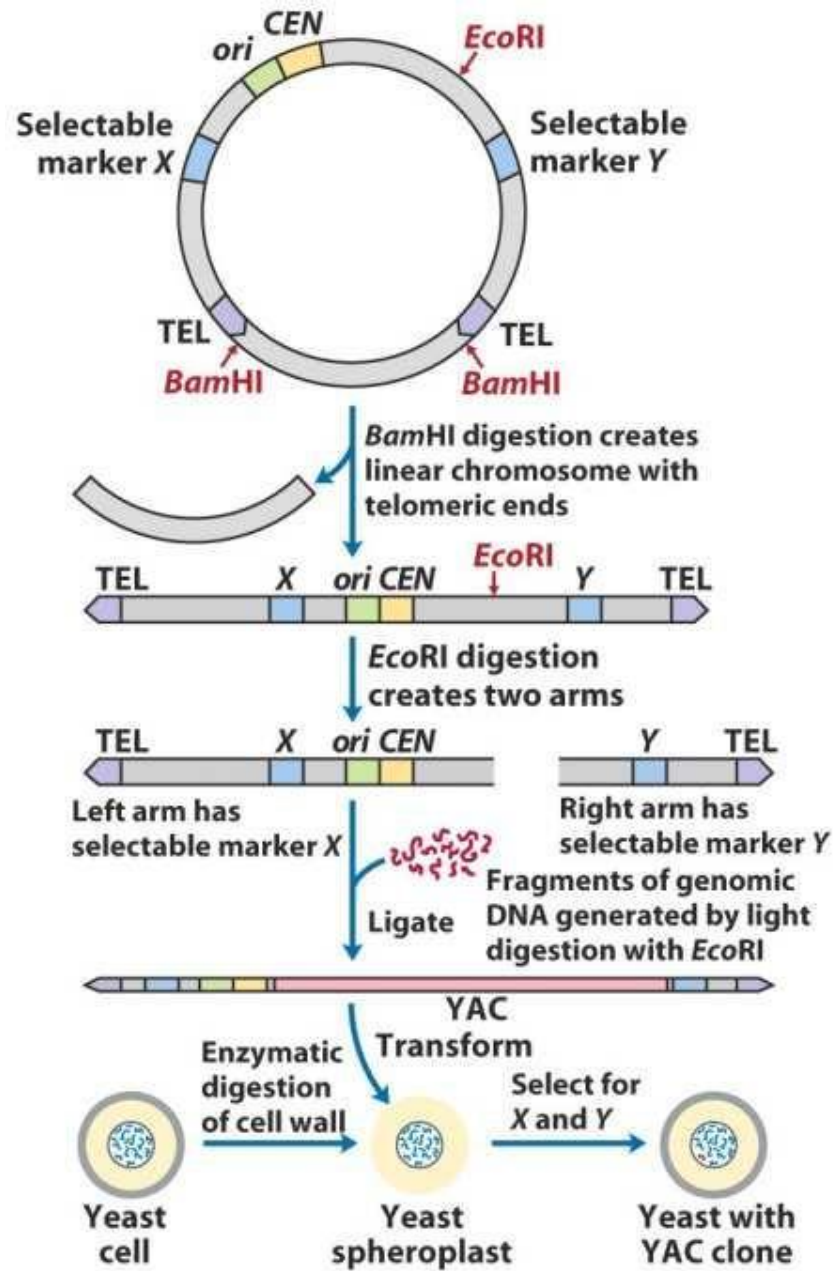
Table 4.6 Insert capacities of some commonly used vector systems

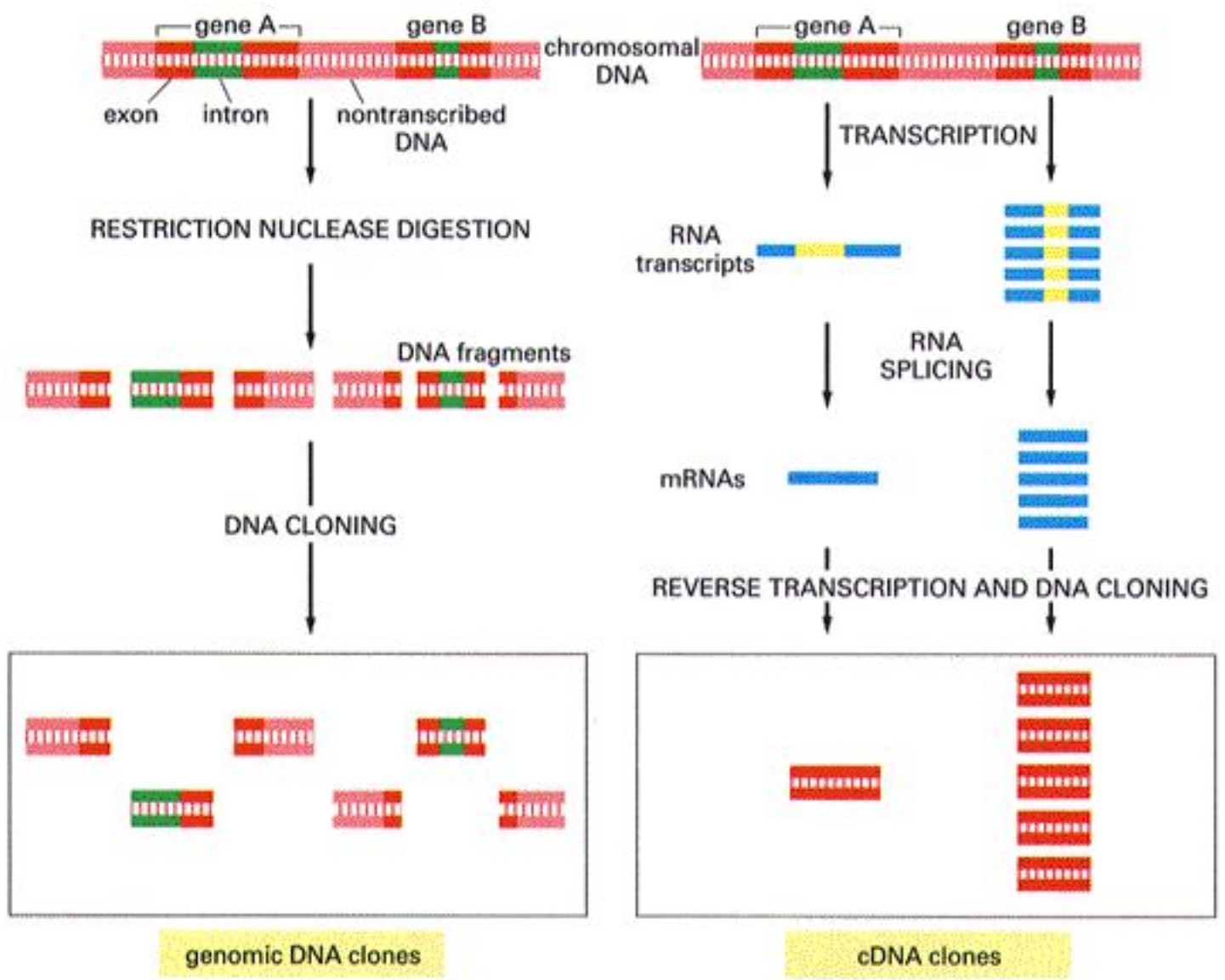
Vector system	Host cell	Insert capacity (kb)
Plasmid	<i>E. coli</i>	0.1–10
Bacteriophage λ	λ / <i>E. coli</i>	10–20
Cosmid	<i>E. coli</i>	35–45
Bacteriophage P1	<i>E. coli</i>	80–100
BAC	<i>E. coli</i>	50–300
P1 bacteriophage-derived artificial chromosome	<i>E. coli</i>	100–300
Yeast artificial chromosome	Yeast	100–2,000
Human artificial chromosome	Cultured human cells	>2,000

Cosmid = Cos - Plasmid

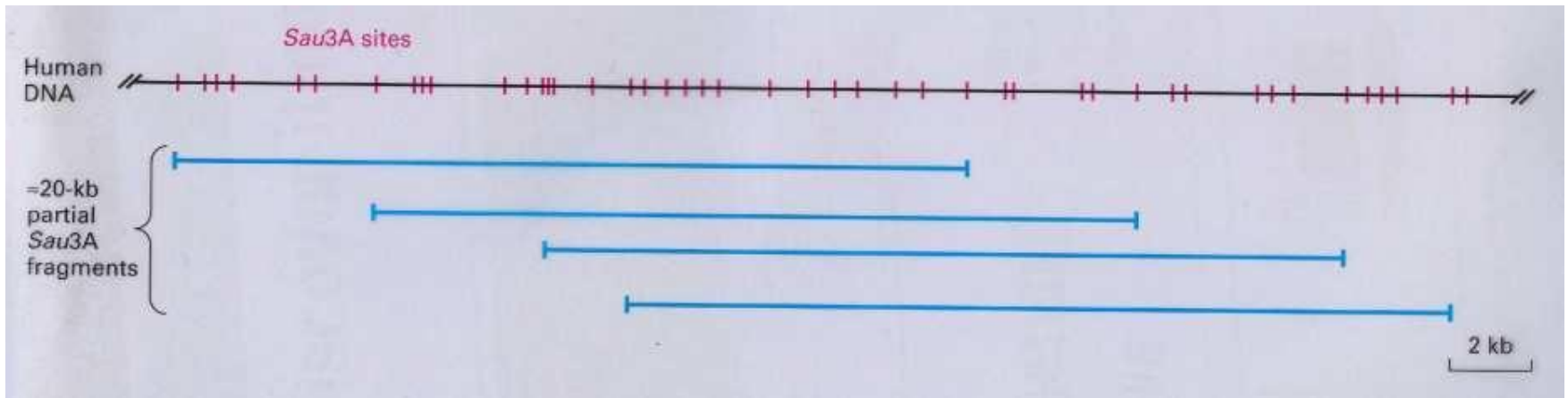


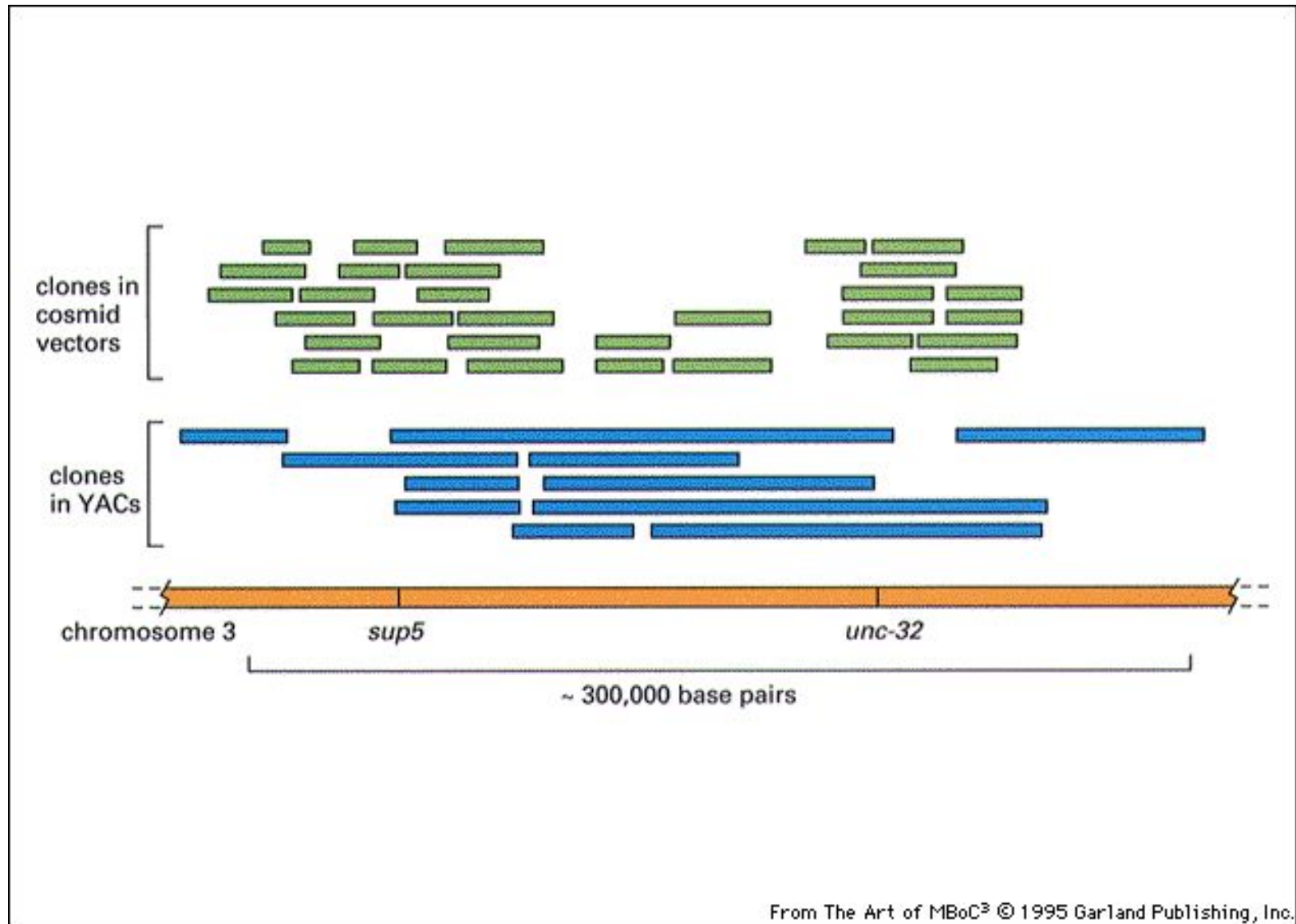






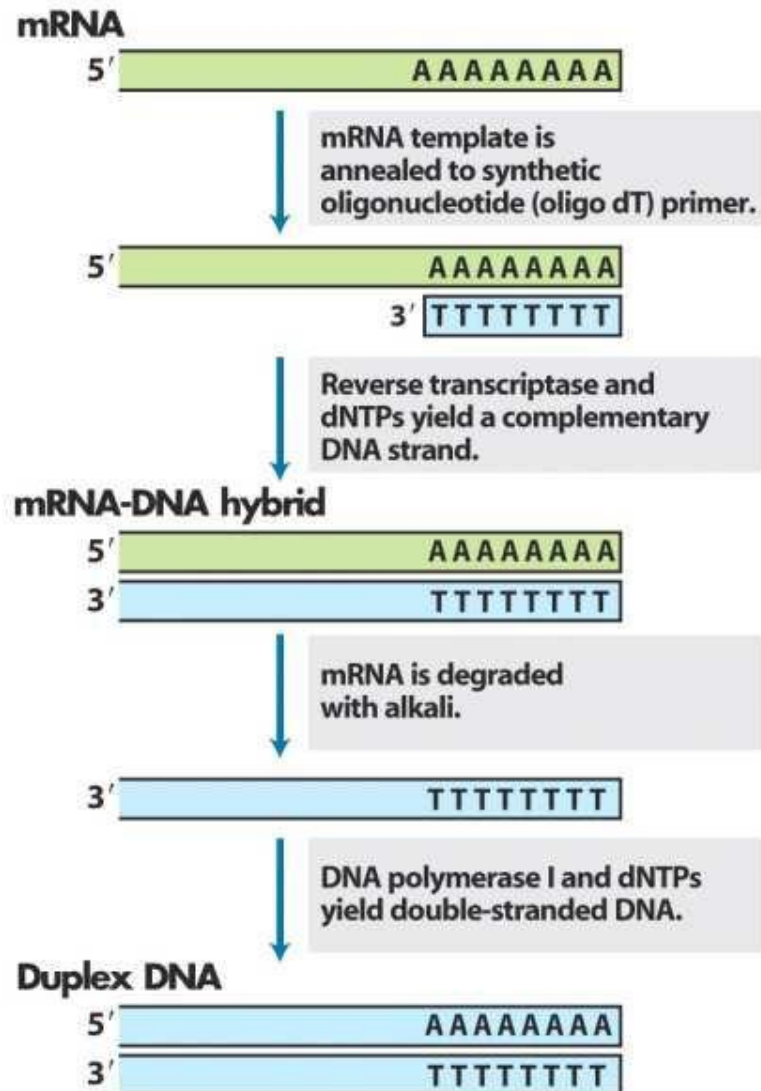
Fragmentation of genomic DNA



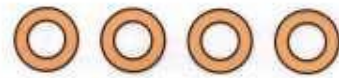


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cDNA synthesis



DNA Library



Plasmid vectors

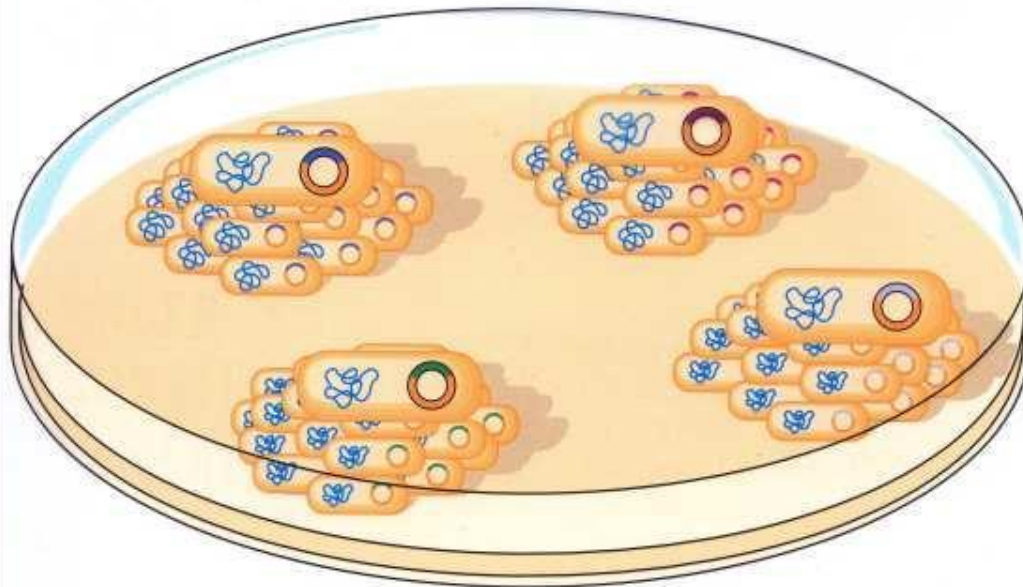


DNA fragments to be cloned

Enzymatically
insert DNA fragments
into plasmid vectors

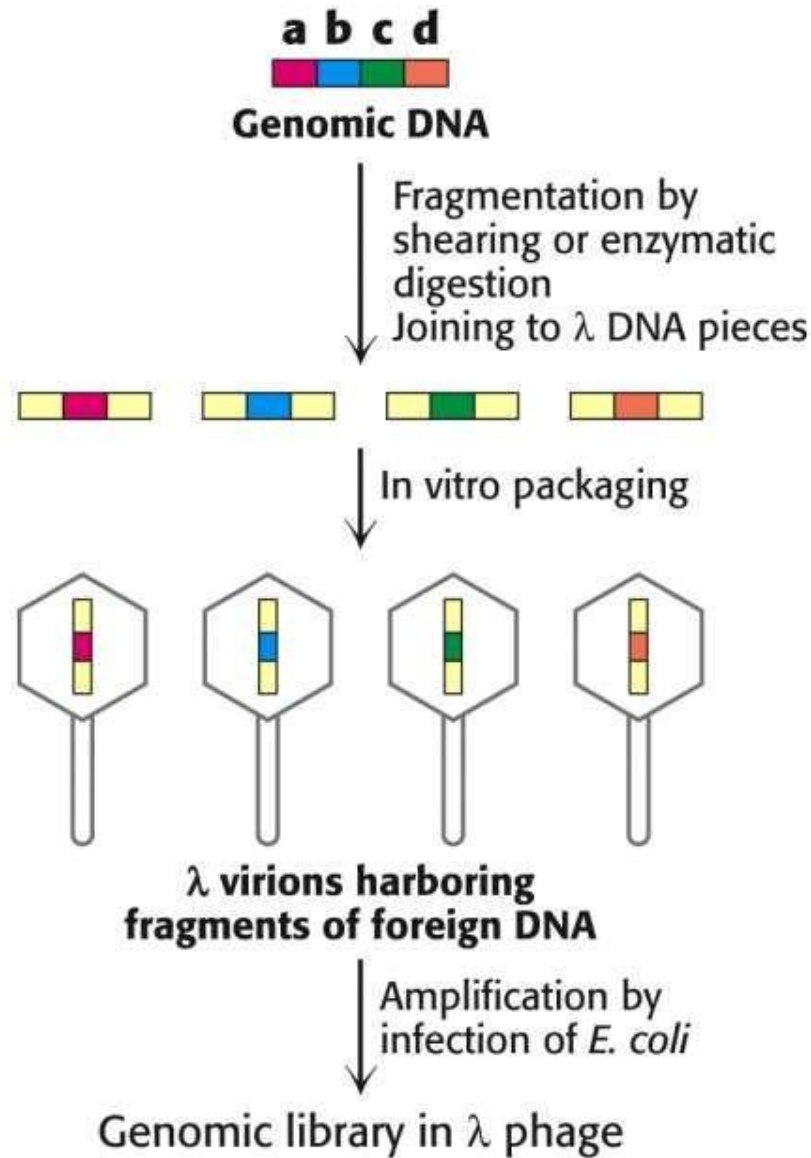


Transform *E. coli* cells
and select for ampicillin-
resistant colonies



Clones ->
genetically
identical

Genomic phage library



Evaluation of library

Box 7.1 Estimates of the required size of genomic libraries

Organism	Genome size	Vector type	Insert size	<i>P</i>	Library size
Bacterium	4×10^6 bases	plasmid	4 kb	0.99	4.6×10^3
		lambda replacement	18 kb	0.99	1.0×10^3
		cosmid	40 kb	0.99	458
		BAC	300 kb	0.99	59
Mammal	3×10^9 bases	plasmid	4 kb	0.99	3.5×10^6
		lambda replacement	18 kb	0.99	7.7×10^5
		cosmid	40 kb	0.99	3.5×10^5
		BAC	300 kb	0.99	4.6×10^4

The values shown for the genome sizes of bacteria and mammals are examples for the purpose of this calculation. The actual genome sizes vary quite widely from one organism to another. The insert sizes for specific vectors will also vary.

Evaluation of library

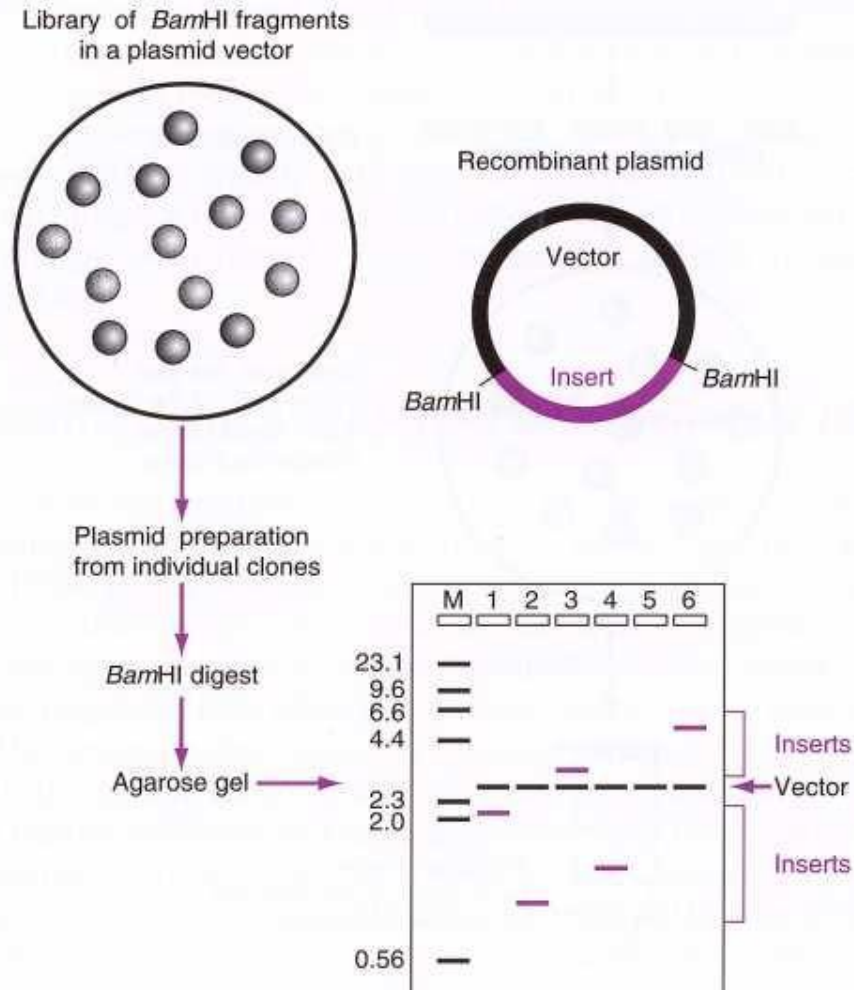
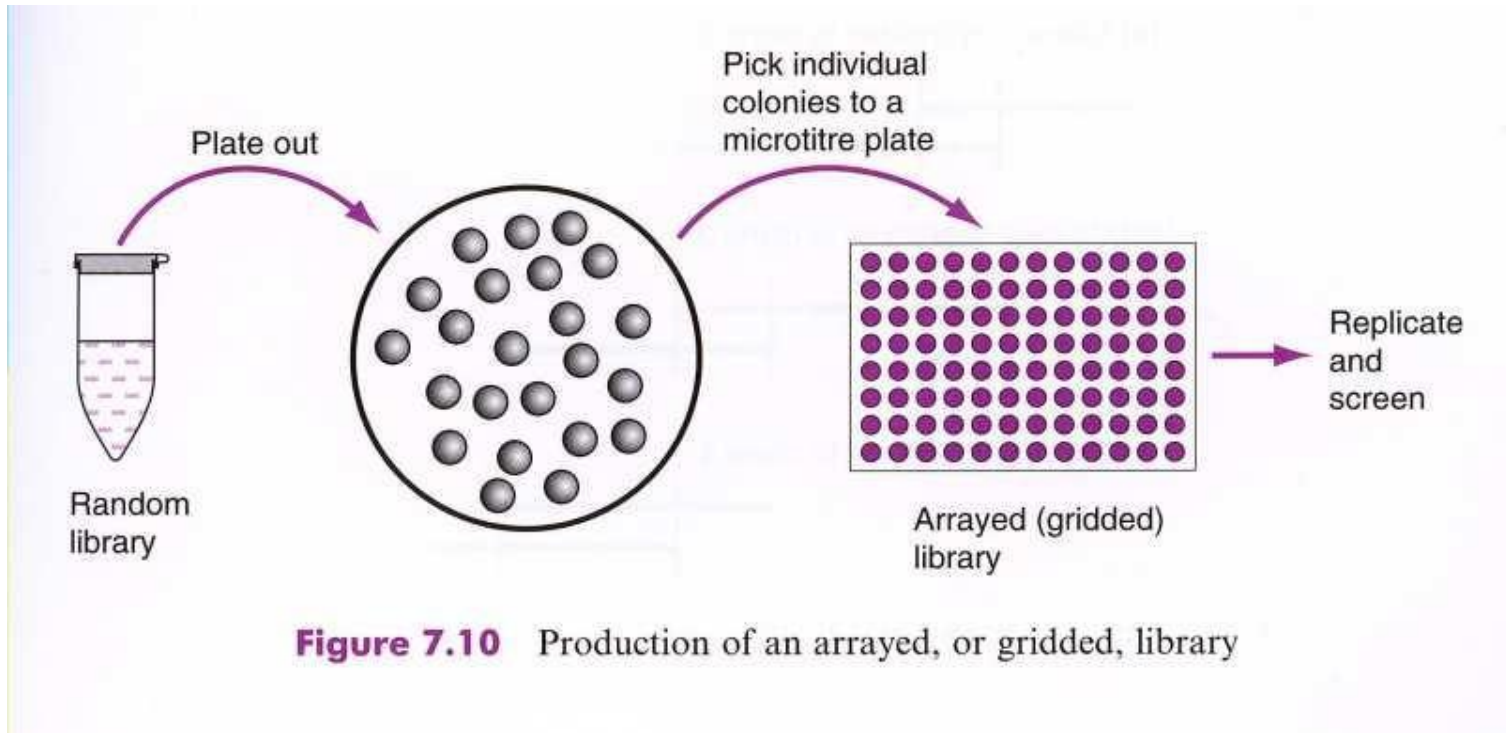


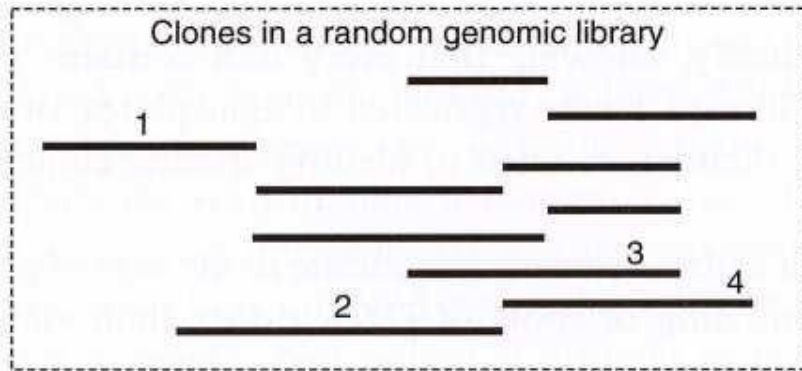
Figure 7.4 Assessing the quality of a gene library

Ordered library



—————> **Microarrays**

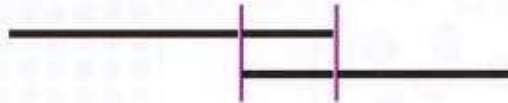
Ordered library



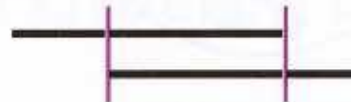
(a) Clone 1 hybridizes to clone 2



(b) Clone 2 hybridizes to clone 3



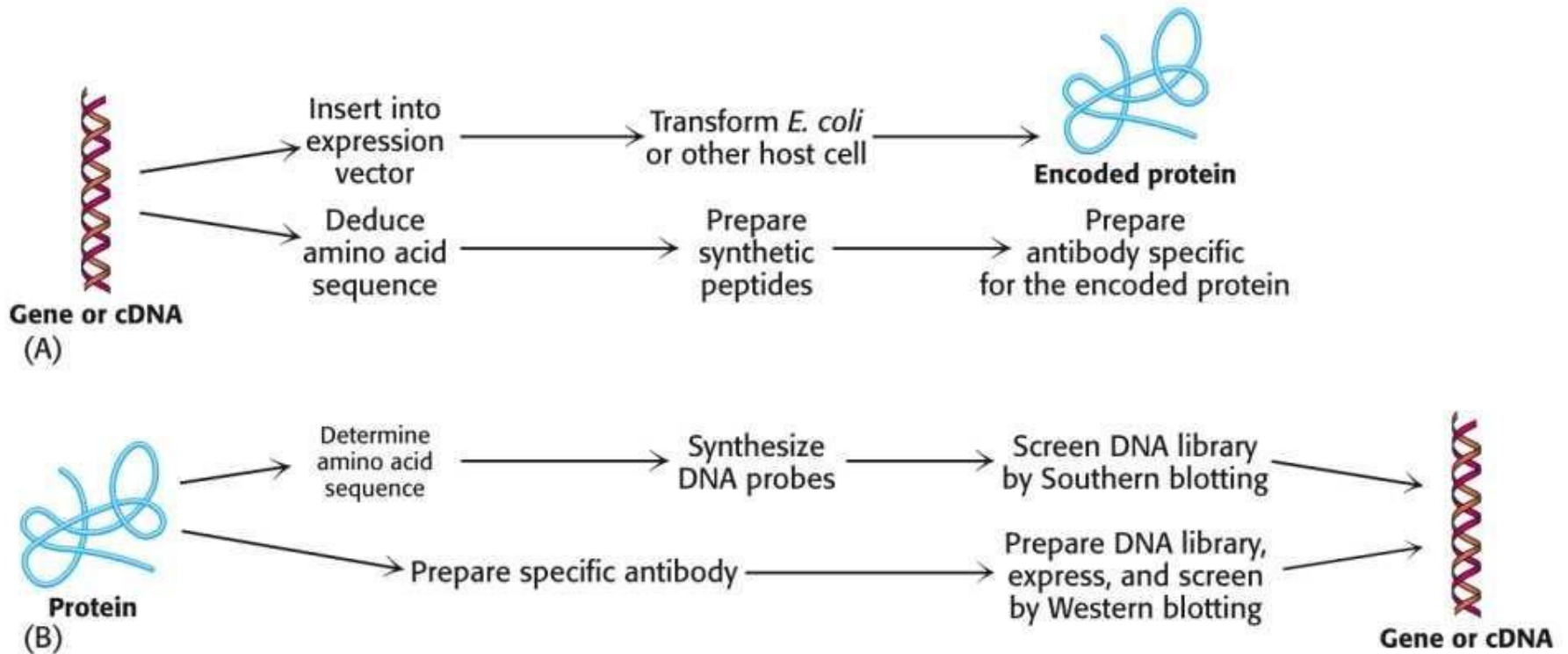
(c) Clone 3 hybridizes to clone 4



→ "Chromosome Walking"
→ also used in "Human Genome Project"

Figure 7.11 Production of an ordered library

Different ways to clone a gene



Bacterial host engineering



Escherichia coli (E. coli) is a type of bacteria normally found in the intestines of people and animals.

Although most strains of E. coli are harmless, some can cause illness or even death.

The most serious form is E. coli **0157:H7**.

E. coli leads to about **73,000 cases of infection** and **61 deaths** each year in the United States.

Genetic and physical maps of the *E. coli* chromosome

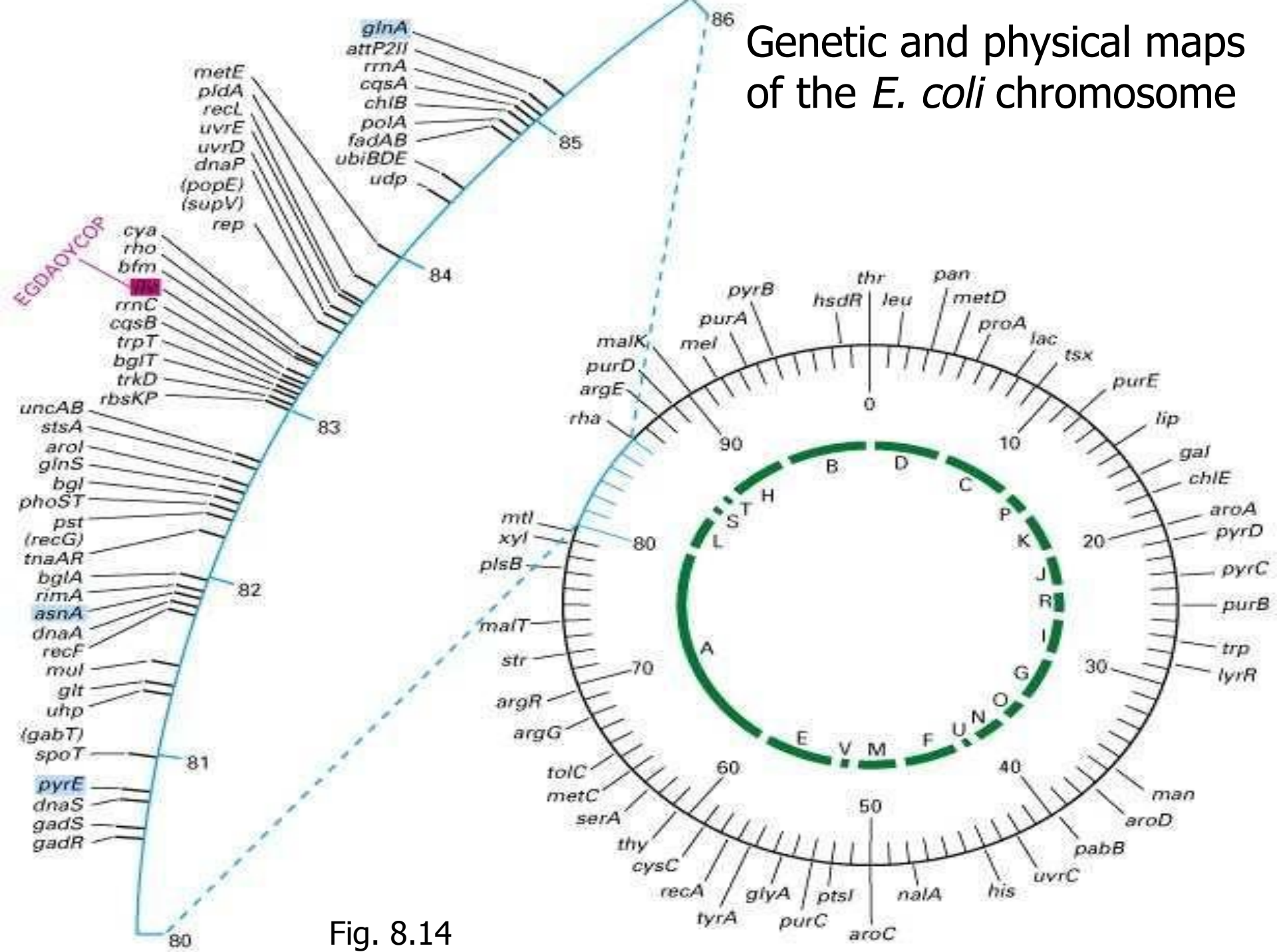


Fig. 8.14

E.Coli K12 strain has been used for further engineering

The K12 strain was first isolated in 1921 from the stool of a malaria patient and it has been maintained in laboratory stocks as a pure strain for the last 75 years.

NIH Recombinant Advisory Committee (RAC) (1973)

Asilomar Conference on Recombinant DNA (February 1975)



Every strain comes with description of its genotype:
DH5alpha (recA-; hsdR-; LacIq; uvrA-; mcrA-.....)

Most strain in molecular biology are *recA⁻ endA⁻ hsdR⁻*

Additional changes in K12 E.coli for ease of the laboratory practice

1. Bacterial restriction modification systems have been removed.

(To prevent its interference with the replication of foreign DNA in bacteria).



hsdR/hsdM/hsdS (EcoK) restriction system

Degrades DNA not methylated
at the sequence 5'-AAC-(N)5-GTGC-3'

- hsdM** recognises unmethylated DNA
- hsdM** is also involved in methylation of DNA
- hsdR** encodes an endonuclease
- hsdS** encodes DNA sequence specific protein

hsdR- or hsdS- mutants facilitate propagation of any foreign DNA

mcrA/mcrB/mrr complex

E.coli DNA is methylated
by **dcm**, **dam** and **hsdM**

mcrA/mcrB/mrr cleaves DNA
methylated by other systems

-mcrA-/mcrB-
strains are good for cloning
eukaryotic DNA

Additional changes in K12 E.coli for ease of the laboratory practice

2. DNA recombination systems are modified to prevent rearrangements (RecA-)

(to prevent deletions and rearrangements)

recA is a core recombination protein

recA- strains allow cloning of repetitive sequences

recA-/recB-/recC- are enhanced strains with very low recombination efficiency

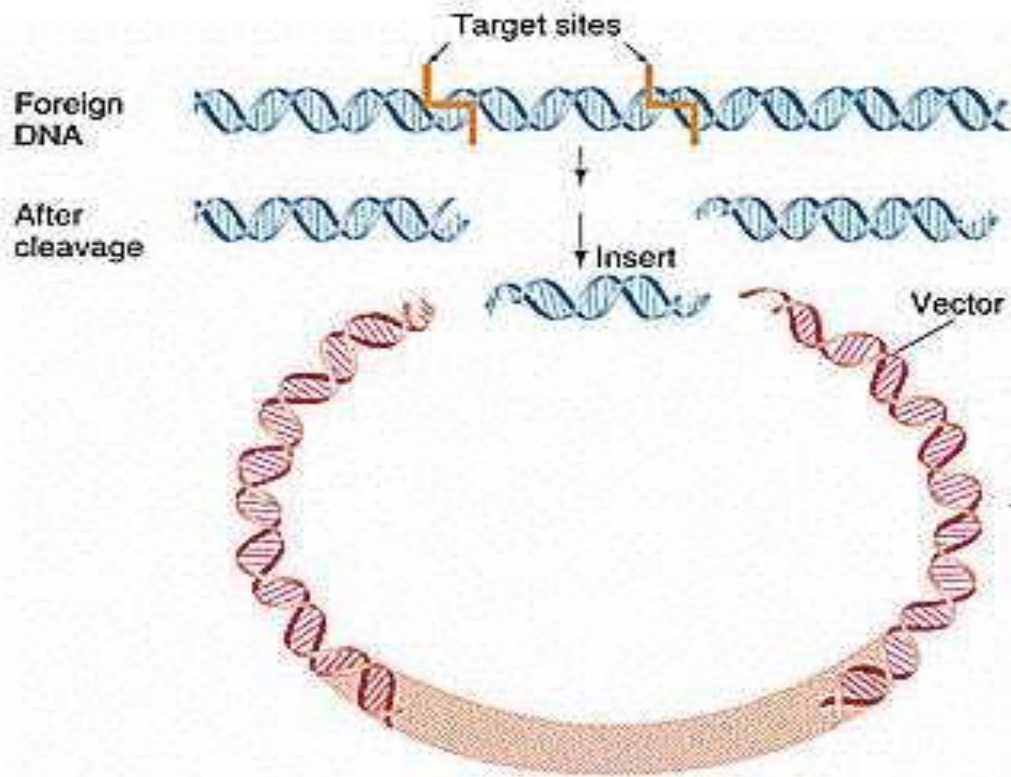
uvrC/umuC are involved in DNA repair

uvrC-/umuC- are good for cloning of inverted repeats

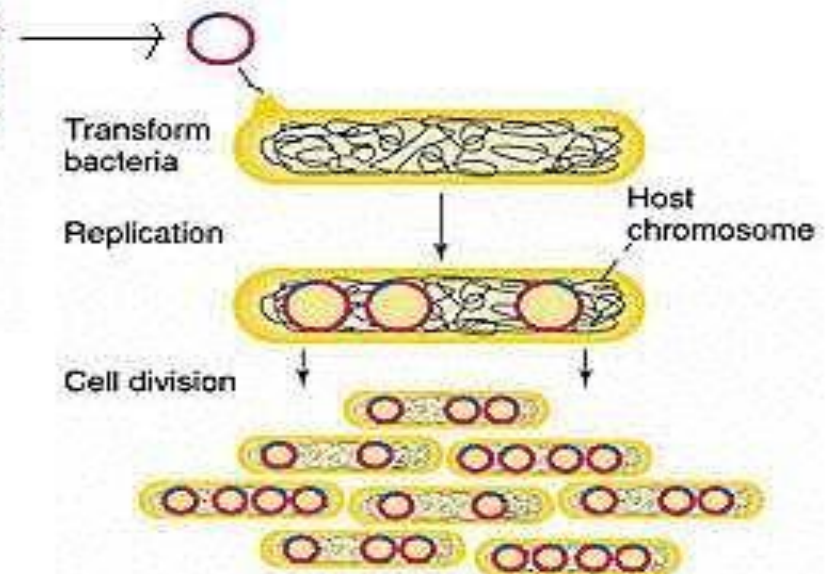
3. Endonuclease activity has been mutated (EndA-)

(to increase plasmid yields and improve the quality of DNA – no nicks)

Transformation of plasmid DNA in competent *E. coli* cells



**Competent (here)
= able to uptake DNA**

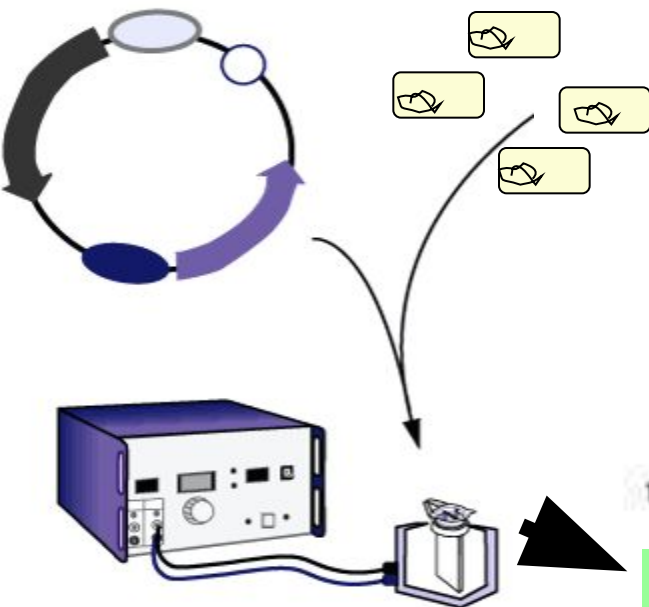


Transformation of plasmid DNA to competent *E. coli* cells

-- Electroporation and electroporation-competent cells

-- Heat shock transformation and chemically competent cells

Electroporation



Chemical transformation

treating *E. coli* with CaCl_2 will batter the membranes and essentially make the bacteria very unhappy.

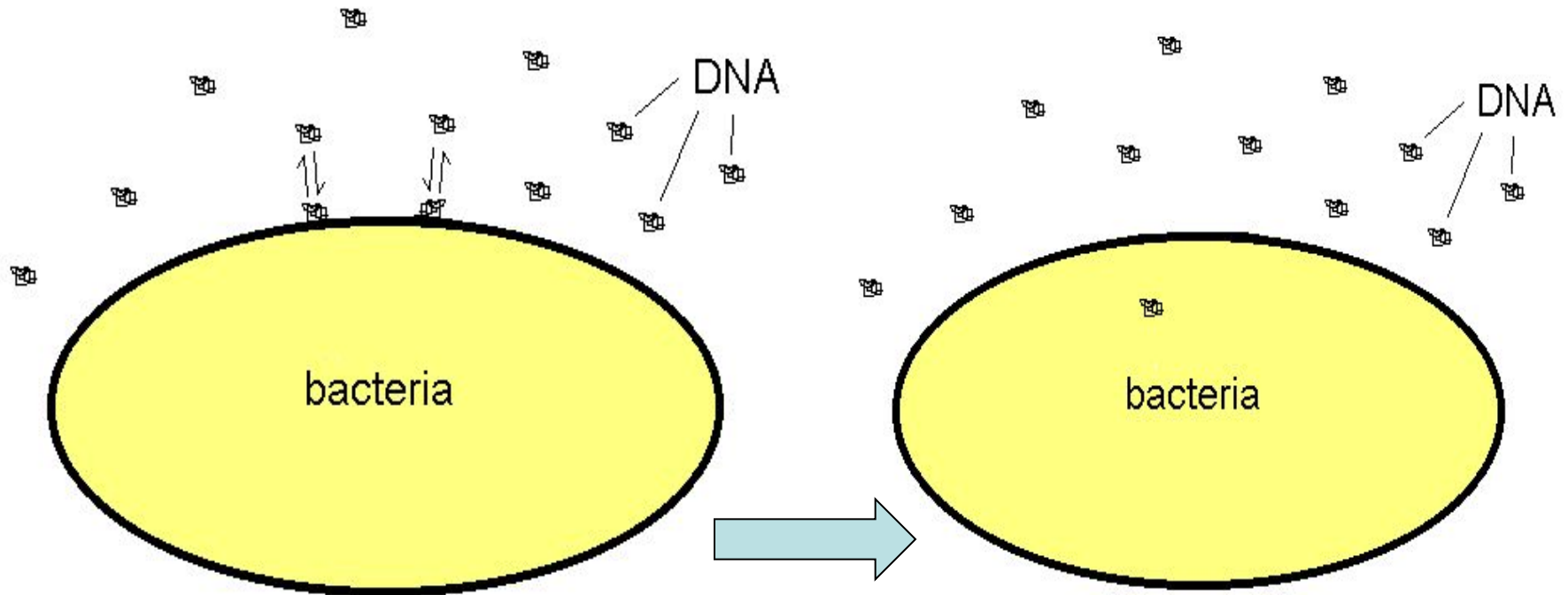
CaCl_2 is gaping holes in the membrane

BRIEF HEAT SHOCK

89

Recovery in rich growing media

Calcium/phosphate (heat shock) method



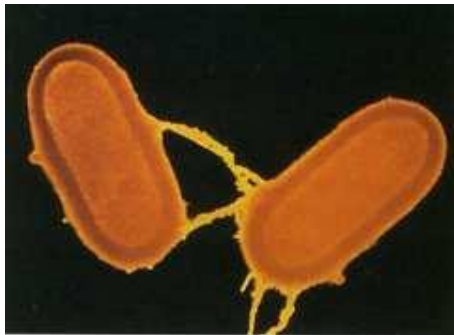
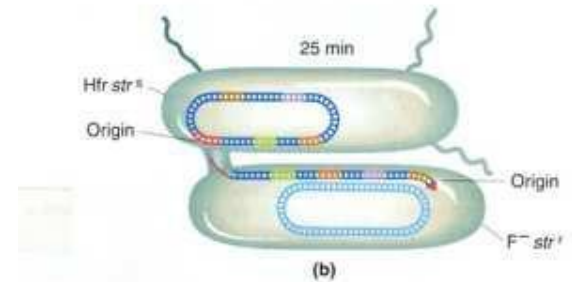
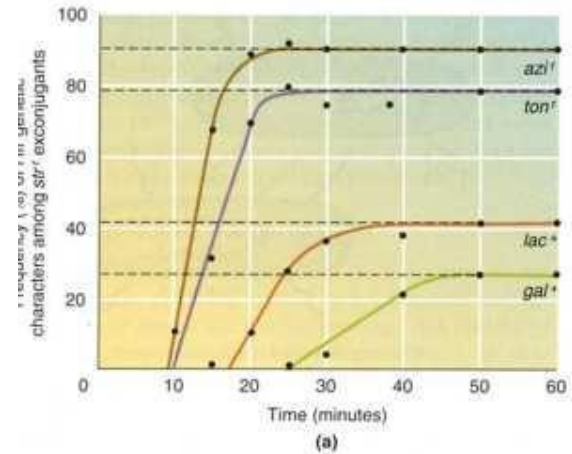
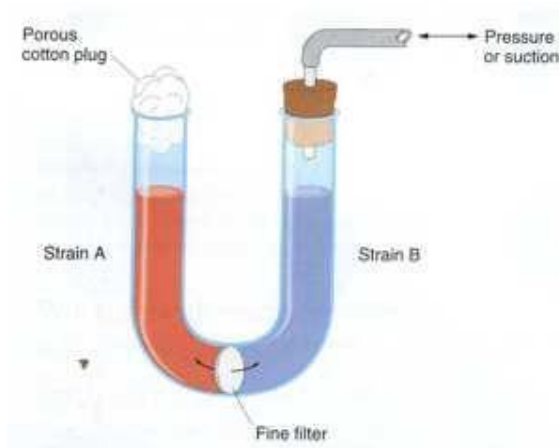
During the incubation on ice, DNA binds to the surface of the bacterium as a calcium-phosphate-DNA complex

Following a sudden increase in temperature, one or more DNA molecules bound to the surface of the cell is taken up by the competent cell.

Conjugation

Lederberg

Monod



- F⁻ to F⁺
- 100 minutes
- 4000 genes